

Role of CD200 and CD43 in Diagnosis and Prognosis of CLL and NHL Patients

*Liqaa Al Sharifi, Haider Abdul Ridha, Ahmed Ebrahim Rashid, Sinan Yahya Muhsin, Teeb Mohammed Jaafer

University of Babylon, Collage of Medicine, Pathology Department

Abstract— Background: Chronic lymphoproliferative disorder (LPD), is a malignant disease of lymphocytes in the blood and lymphatic tissue. Chronic lymphocytic leukaemia, it is the commonest type of chronic lymphoproliferative disorder.

Scoring by immunophenotyping is used to differentiate B-cell chronic lymphocytic leukemia from other B-Non-Hodgkin lymphomas. CD200 (OX2) is a glycoprotein of membrane, it is related to type I superfamily of immunoglobulin .

CD43 (Sialophorin) is a sialoglycoprotein that is present on the surface of T lymphocytes, some B lymphocytes, granulocytes and monocytes, that play important role for immune function and also play a role as physiologic ligand-receptor complex involved in T-cell activation.

Aim of study-Evaluate the role of CD200 and CD43 positive expression and co-expression in diagnosis and prognosis of CLL and NHL.

Subjects and Methods: This cross sectional study on one hundred forty five patients with chronic lymphoproliferative disorders who were attending Baghdad teaching hospital at medical city from beginning of January 2020 to end of December 2020, patients divided in to two groups; chronic lymphocytic leukemia (CLL) and non-Hodgkin lymphoma (NHL) patients At diagnosis, there was assessment of CD200 and CD43. Clinical and laboratory data were done including staging by modified Rai, and Ann arbor staging system (for CLL and NHL respectively), then follow up for about 6-12.

Results: There is significant statistical correlation between CD 200 and CD 43 and often co-expression of both in differentiation between CLL and NHL (p value < 0.001).

Almost all patients of CD 200 positive expression show moderate to bright pattern of expression in CLL in apposite to NHL patients, who showed dim to moderate pattern of expression, while the great majority of CD43 expression was dim to moderate pattern in both CLL and NHL patients. In CLL patients there was no significant correlation between prognostic markers (age, Hb, platelets count, lymphocytes count and CD 38 expression) and CD200, CD 43.

For NHL patients, all markers show no significant correlation except that CD 43 show significant correlation with expression of CD 38.

Conclusion: CD200, CD43 and often co-expression of both have a significant value in diagnosis and differentiation of CLL from B-NHLs.

I. INTRODUCTION

This chronic lymphoproliferative disorder (LPD), is a malignant disorder of lymphocytes in blood and lymphatic tissue. Chronic lymphocytic leukaemia is the commonest chronic lymphoproliferative disorder (LPD) (1). It is frequently diagnosed by combination of clinical, cytogenetic and

immunophenotyping (2). Meanwhile, some of CLL cases still unclear because the cytological and flowcytometric markers can overlay with other lymphoproliferative diseases that present with leukemic phase (3). Many efforts have been made to use immunophenotypic markers by flowcytometry or called scoring system that greatly help to differentiate between CLL and NHL, till now, there is no single test that can clearly differentiate between them (4).

Scoring system by flowcytometry is used to differentiate B-cell chronic lymphocytic leukemia (B-CLL) from B-NHLs. expression of CD23, CD5 and absent or weak expression of CD79b, FMC7, also weak expression of surface IgM make the score is five. However, cases diagnosed as B-NHL had intermediate score of (2–3 points) (3). So scoring by flowcytometry is an important tool in the diagnostics of B-cell non-Hodgkin lymphomas (B-NHLs). The European Research Initiative on CLL implication for flow cytometry, they add another important markers like CD43, CD81, CD200, CD10 and ROR1 for differential diagnosis of CLL and low grade B cell lymphoma (5).

CD200 (OX2) it is a glycoprotein of cell membrane and belongs to the type I superfamily of immunoglobulin. It is usually found or expressed on many of human cells including T and B lymphocytes (6).

CD43 (Sialophorin) it is a sialoglycoprotein on the top of human T lymphocytes, B lymphocytes, granulocytes and monocytes, that play an important for immunity and act as physiologic ligand-receptor complex involved in T-cell activation (7).

The aims of study:

Evaluate the role of CD200 and CD43 positive expression and co expression in diagnosis and prognosis of CLL and NHL patients

II. PATIENT , MATERIALS AND METHOD

A. Patients

1. Study subject

This study including 145 patients diagnosed CLL and leukaemic phase of NHL through January to December 2020. Those patients were collected from hematology department in Baghdad teaching hospital, the immunophenotyping done by 8 colour flowcytometry. The diagnosis depend on clinical data, blood film and bone marrow examination which showed absolute lymphocytosis of mature morphology, then immunophenotypic analysis by flowcytometry with MS scoring

was done for diagnosis of chronic lymphocytic leukaemia and non-Hodgkin lymphoma, CD38 expression was also measured in addition to CD200 and CD43 which are the main markers on the study. At diagnosis, clinical information including age, gender, clinical examination such as organomegaly, generalize or localize lymphadenopathy were obtained from patient laboratory case sheet. According to hematological parameters, physical and clinical findings we made staging for each patient in order to make good categorization for patient stage and the future response to treatment. So for CLL patients we use modified Rai staging system to differentiate low, intermediate and high risk group. For NHL patients we depend on ann arbor staging system, also to make categorization of patient in to four staging group in which there are four groups (I,II,III,IV). All patient were evaluated there after either by face to face collection of data on their attendance to hospital or by phoning, follow up program including all patients (on treatment or those whose physician prefer to not use treatment) for six to twelve months, evaluation of their hematological, clinical and physical response and assessment of their remission state depending on blood film, bone marrow finding and some patient with detection of minimal residual disease by flowcytometry was done.

2. Inclusion criteria

- 1-Newly diagnosed mature B- cells neoplasm.
- 2-The patient are randomly collected in relation to gender.
- 3-Adult group patients.
- 4-All patient were not receive any treatment before sample collection.

3. Data Analysis

Statistical analysis was carried out using SPSS version 25. Categorical variables were presented as frequencies and percentages. Continuous variables were presented as (Means \pm SD). Student t-test was used to compare means between two groups. Mann-Whitney Test was used to compare means between two groups when variable was not normally distributed. Pearson chi-square and Fisher's exact test were used to find the association between categorical variables. A *p*-value of ≤ 0.05 was considered as significant.

III. THE RESULTS

A. The study groups

This is across sectional study including 145 LPD patients diagnosed by flowcytometry in to two main subtypes, 98 patients with CLL and 47 patients with NHL have mean age of 62.73 and 62.1 years respectively. Male is more than female percentage as in table (1).

B. Age and sex distribution

Table 4.1: The association between socio-demographic characteristics and diagnosis (N=145)

Study variables	CLL (N=98)	NHL (N=47)	P-value
Age (years)	(62.73 \pm 9.20)	(62.10 \pm 10.25)	0.711
Gender			
Male	61 (62.2)	32 (68.1)	0.493
Female	37 (37.8)	15 (31.9)	
Total	98 (100.0)	47 (100.0)	

IV. ROLE OF CD200 AND CD43 IN DIFFERENTIATION BETWEEN CLL AND NHL.

A. The relevance of each CD marker in diagnosis of CLL and NHL

There was significant association between study variables including (CD200 and CD43) and diagnosis, CD200 and CD43 are positive in high percentage of CLL patients, *p* value below 0.001, as shown in table (2)

Table 4.2 Association between study variables (CD200, CD43 and CD 305) and diagnosis (N=145)

Study variables	Diagnosis		Total	X ²	P-value
	CLL	Non Hodgkin Lymphoma			
CD 200					
Positive	98 (100.0)	29 (61.7)	127 (87.6)	42.85	<0.001*
Negative	0 (0.0)	18 (38.3)	18 (12.4)		
Total	98 (100.0)	47 (100.0)	145 (100.0)		
CD 43					
Positive	82 (83.7)	15 (31.9)	97 (66.9)	38.42	<0.001*
Negative	16 (16.3)	32 (68.1)	48 (33.1)		
Total	98 (100.0)	47 (100.0)	145 (100.0)		

B. The significance of co-expression CD 200 and CD 43 in CLL and NHL cases

There is significant positive co-expression of both CD200 and CD43 and diagnosis of CLL, *p* value below 0.001 as shown in table 3.

Table 4.3. Association between CD200 and CD43 co-expression and diagnosis (N=145)

Study variables	Diagnosis		Total	P-value
	CLL (98)	NHL (47)		
Positive CD 200 and CD 43 co-expression	82 (83.7)	9 (19.1)	91 (62.8)	<0.001*

C. Association of pattern of expression and diagnosis

There is significant correlation between pattern (bright, moderate and dim) expression of CD200 and the provisional diagnosis with moderate to bright expression in CLL, *p* value below 0.001, as in figure 1

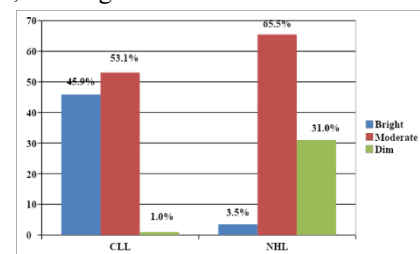


Figure 4.1: Distribution of patients with positive CD200 according to flow cytometry pattern of positive (N=127, P<0.001*)

There is no significant correlation between pattern (bright, moderate and dim) expression of CD43 and the provisional diagnosis, *p* value 1.000.as in figure 2.

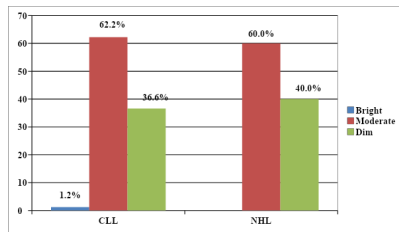


Figure 4.2: Distribution of patients with positive CD43 according to flow cytometry pattern of positive (N=97, P=1.000)

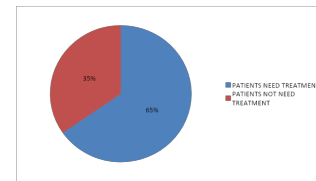


Figure 4.4. Show number of CLL patients who need treatment 64 (65%), and those who not need 34 (35%) total number 98 patients.

D. Role of CD 200 and CD 43 in prognosis of CLL patients

There is no significant association between CD200, and CD43 with age, Hb. Level, platelets, lymphocytes count and CD38 in CLL patients as in table (4)

Table 4.4. Association between study variables (CD200, CD43) and laboratory prognostics factors in CLL patients (N=98)

Prognostic factors	CD 200			CD43		
	+(98)	-(0)	P	+(82)	-(16)	P
Age						
< 60	36 (36.7)	0 (0.0)	-	30 (36.6)	6 (37.5)	0.945
≥ 60	62 (63.3)	0 (0.0)		52 (63.4)	10 (62.5)	
Hb						
< 11	46 (46.9)	0 (0.0)	-	40 (48.8)	6 (37.5)	0.408
≥ 11	52 (53.1)	0 (0.0)		42 (51.2)	10 (62.5)	
Platelet count						
<100	11 (11.2)	0 (0.0)	-	8 (9.8)	3 (18.8)	0.381
≥ 100	87 (88.8)	0 (0.0)		74 (90.2)	13 (81.2)	
CD 38						
Positive	13 (13.3)	0 (0.0)	-	9 (11.0)	4 (25.0)	0.218
Negative	85 (86.7)	0 (0.0)		73 (89.0)	12 (75.0)	
Lymphocyte						
< 30	56 (57.1)	0 (0.0)	-	47 (57.3)	9 (56.2)	0.937
≥ 30	42 (42.9)	0 (0.0)		35 (42.7)	7 (43.8)	

Relation to clinical stage of disease

. CD200 and CD43 has no significant relation to clinical stage as shown in table (5).

Table 4.5. Association between study variables (CD200, CD43) and clinical stage among CLL patients (N=98)

Study variables	Clinical Stage					Total	X ²	P-value
	Low risk (0) (8.2%)	Intermediate risk		High risk				
		I	II (56%)	III	IV (35.7%)			
CD 200								
Positive	8 (100.0)	31 (100.0)	24 (100.0)	23 (100.0)	12 (100.0)	98 (100.0)	-	-
Negative	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)		
Total	8 (100.0)	31 (100.0)	24 (100.0)	23 (100.0)	12 (100.0)	98 (100.0)		
CD 43								
Positive	8 (100.0)	26 (83.9)	19 (79.2)	20 (87.0)	9 (75.0)	82 (83.7)		0.652 f
Negative	0 (0.0)	5 (16.1)	5 (20.8)	3 (13.0)	3 (25.0)	16 (16.3)		
Total	8 (100.0)	31 (100.0)	24 (100.0)	23 (100.0)	12 (100.0)	98 (100.0)		

Relation to state of remission

35% of CLL patients not need treatment, as shown in figure (4) CD200 and CD43 had neither association to clinical stage of disease nor to remission state after treatment.

Table 4.6. Association between study variables (CD200, CD43) and state of remission in CLL patients (N=98)

Study variables	State of remission				Total	P-value
	Patients die while on treatment	Patients have treatment without remission	Patients have treatment with remission	Patients not need treatment		
CD 200						
Positive	9 (100.0)	14 (100.0)	41 (100.0)	34 (100.0)	98 (100.0)	-
Negative	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	
Total	9 (100.0)	14 (100.0)	41 (100.0)	34 (100.0)	98 (100.0)	
CD 43						
Positive	7 (77.8)	11 (78.6)	33 (80.5)	31 (91.2)	82 (83.7)	0.468
Negative	2 (22.2)	3 (21.4)	8 (19.5)	3 (8.8)	16 (16.3)	
Total	9 (100.0)	14 (100.0)	41 (100.0)	34 (100.0)	98 (100.0)	

E. Role of CD 200, CD43 in prognosis of NHL patients

Association with laboratory factors

There is no significant association between CD200, CD43 with age, platelets, lymphocytes count in NHL patients, p value 0.03 as in table (7).

Table 4.7. Association between study variables (CD200, CD43) and laboratory prognostics factors in NHL patients (N=47)

Prognostic factors	CD 200		P	CD43		P
	+(29)	-(18)		+(15)	-(32)	
Age						
< 60	12 (41.4)	4 (22.2)	0.178	6 (40.0)	10 (31.3)	0.555
≥ 60	17 (58.6)	14 (77.8)		9 (60.0)	22 (68.7)	
Hb						
< 11	14 (48.3)	11 (61.1)	0.391	9 (60.0)	16 (50.0)	0.522
≥ 11	15 (51.7)	7 (38.9)		6 (40.0)	16 (50.0)	
Platelet count						
<100	8 (27.6)	3 (16.7)	0.492	5 (33.3)	6 (18.8)	0.292
≥ 100	21 (72.4)	15 (83.3)		10 (66.7)	26 (81.2)	
CD 38						
Positive	6 (20.7)	2 (11.1)	0.692	6 (40.0)	2 (6.3)	0.009*
Negative	23 (79.3)	16 (88.9)		9 (60.0)	30 (93.7)	
Lymphocyte count	17.18	30.11	0.347	12.58	26.61	0.268

Relation to clinical stage of disease

CD200 and CD43 has no significant role as shown in table (8).

Table 4.8. Association between study variables (CD200, CD43 and CD305) and clinical stage among non-Hodgkin lymphoma patients (N=47)

Study variables	Clinical Stage				Total	P-value
	Low risk		High risk			
	I	II (27.7%)	III	IV (72.3%)		
CD 200						
Positive	1 (100.0)	10 (83.3)	12 (50.0)	6 (60.0)	29 (61.7)	0.215
Negative	0 (0.0)	2 (16.7)	12 (50.0)	4 (40.0)	18 (38.3)	
Total	1 (100.0)	12 (100.0)	24 (100.0)	10 (100.0)	47 (100.0)	
CD 43						
Positive	0 (0.0)	3 (25.0)	7 (29.2)	5 (50.0)	15 (31.9)	0.577
Negative	1 (100.0)	9 (75.0)	17 (70.8)	5 (50.0)	32 (68.1)	
Total	1 (100.0)	12 (100.0)	24 (100.0)	10 (100.0)	47 (100.0)	

Relation to state of remission

CD43 negative expression correlated with achievement of remission state and with patients on no treatments, as shown in table (9) keeping in mind that 89% of NHL patients in this study were on treatment as in figure (5).

Table 4.9. Association between study variables (CD200, CD43) and state of remission in NHL (N=47)

Study variables	State of remission				Total	P-value
	Patients die while on treatment	Patients have treatment without remission	Patients have treatment with remission	Patients not need treatment		
CD 200						
Positive	5 (55.6)	12 (54.5)	7 (63.6)	5 (100.0)	29 (61.7)	0.336
Negative	4 (44.4)	10 (45.5)	4 (36.4)	0 (0.0)	18 (38.3)	
Total	9 (100.0)	22 (100.0)	11 (100.0)	5 (100.0)	47 (100.0)	
CD 43						
Positive	3 (33.3)	11 (50.0)	0 (0.0)	1 (20.0)	15 (31.9)	0.019*
Negative	6 (66.7)	11 (50.0)	11 (100.0)	4 (80.0)	32 (68.1)	
Total	9 (100.0)	22 (100.0)	11 (100.0)	5 (100.0)	47 (100.0)	

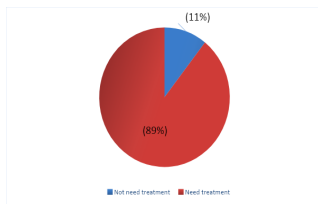


Figure 4.5. Show number of NHL patients who need treatment 42 (89%), and those who not need 5 (11%) total number 47 patients

DISCUSSION

A. Age and sex distribution in patients with CLL and NHL

In this study, mean age of CLL patients is 62.73 and NHL is 62.1, this goes with the fact that LPDs can affect elderly age group and has clear age preference (3). LPD has no sex predilection, and in this study, no significant sex difference between CLL and NHL patients, although most were male that account for (62%) in CLL, (68%) for NHL patients. This may be attributed to the sample size and random chance.

B. Expression of CD200, CD43 in diagnosis of CLL and NHL

We investigate the value of expression of CD200, CD 43 in diagnosis of LPDs

- CD200 was significantly positive in almost all patients in this study, with moderate to bright expression in all CLL (98/98) (100%), while in (NHL) patients CD 200 was mostly dim to moderate expression in 29 out of 47 patients (61.7%) (p value < 0.001) as in table 2, figure 1.

The explanation of the difference in expression patterns of CD200 in BCLPDs, literature partially mentioned this to the different activation of the AKT and MEK/ERK pathways in these different disorders (4).

- CD43 was expressed in (82/98) (83.7%) in CLL patients with dim to moderate expression while it was almost always dim - moderate expression in 15 out of 47 NHL patients (31.9%) (p value < 0.001) as show in table 2, figure 2, these findings are consistent with El Desoukey NA et al, Sandes AF et al, Alapat D et al, Brunetti L et al, Pillai V et al, McWhirter JR et al and Kohnke T et al; all those series show that almost all CLL cases show moderate to bright expression of CD200 and CD 43 with lower expression in those with NHL patients. (4, 5,40,48, 49, 50,51).

In this series of chronic lymphoproliferative disorders with exclusive PB and/or BM affection, CD43-positivity increased with increasing Matutes score of CLL, suggesting that this CD marker could be of help in the distinctive role in diagnosis of complex or borderline cases. The frequent CD43-positivity in CLL has been well recognized both by FC and our results agree with Sorigue M et al, Quijano S et al, Kostopoulos IV et al, Jung G et al. (7,52,53,58).

Other series show that CD43 improved the classification accuracy of the classical scoring system. However, the inclusion of CD43 offers a better performance that affect discriminant functions rather than the classical scoring system. So CD43, together with classical markers, is useful in

differentiation between CLL and non-CLL, and should be systematically used in addition to well-known methods to improve the diagnosis and classification of the B-cell lymphoproliferative disorders (58).

-Co-expression of both 200 and 43 in differentiation between CLL and NHL; 82 with CLL out of 98(83.6%) of patients show co-expression of both CD200 and CD43 while 9 with NHL out of 47(19.1) of patients show both CD200 and CD43 markers expression, p value was significant (< 0.001) as show in table 3.

Some study state that the combination of CD200 and CD43 offers a highest accuracy to differentiate between CLL and non-CLL LPDs. However borderline LPDs, which constitute a small proportion of cases, remain a diagnostic challenge and cannot be dependably identified or readily diagnosed with our proposed combination of CD200 and CD43. but further studies will be necessary to determine a precise score (36).

We confirmed that the introduce CD200 and CD 43 to CLL matutes score system had a very high accuracy to distinguish between CLL and NHL patients(3).

C. Role of CD 200 and CD 43 as a prognostic parameter in CLL patients:

In this study, also we evaluate whether the use of CD200 and CD43 as an innovative routine marker could be helpful to improve the utility of flow cytometry in prognostic significance of CLL patients.

Prognostic factors can help us to sort CLL patients who need immediate therapy soon after diagnosis in which include certain clinical and laboratory features, cytogenetic, molecular, and biochemical characteristics of the neoplastic cell.

-CD 200 there is no significant statistical correlation between CD 200 expression and age, Hb, platelet count, lymphocytes count and CD38. Meanwhile about 86.7% of same group have negative expression of CD 38, which indicate good prognostic group (p value 0.037) as in table 4. the above parameters consider a prognostic laboratory features, these finding is consistent with El Desoukey NA et al and D'Arena G et al in whom state that CD200 expression in B-CLL was not correlated with these laboratory features (4,19); while in contrast to our results, Dorfman DM et al, El Din Fouad et al, Wang et al., Zahedi M et al and Wong KK conclude that positive CD200 expression was associated with these prognostic variable.(19,55,56,59,72) .

In present study we also notify that correlation of CD200 positive expression to the clinical stage of the disease that 35 out of 98 patients (35.7%) have high risk group (stage III-IV) while 63patients (64%) of them low and intermediate group (0-I-II) according to RIA staging system (there is no p value because all CLL patients was CD200 positive as in table 5, similarly El Desoukey NA et al and D'Arena G et al that consider the CD200 expression in B-CLL was not correlated with the clinical staging system (RAI staging) (4,34) while in contrast to our results, Dorfman DM et al, El Din Fouad et al, Wang et al., Zahedi M et al and Wong KK conclude that positive CD200 expression was associated with advanced stage and earlier time to progression. (19,55,56,59,72) .

There is no significant clinical association between CD200 expression and the state of remission as in table 6, this finding is consistent with El Din Fouad et al that conclude that there is

no correlation with response to treatment and overall survival (55), as in table 6.

-CD 43 there is no significant statistical correlation between CD 43 expression and age, Hb, platelet count, lymphocytes count and CD 38, as in table 4.

also notify there is no significant correlation between CD43 expression and the clinical stage of the disease, as show in table 5. There are no significant clinical association between CD43 expression and the state of remission (p value 0.468) as show in 6. No other study correlate the expression of CD43 as a prognostic factor.

D. Role of CD200 and CD43 as a prognostic parameter in NHL patients

-CD200 there is no significant correlation between CD200 expression and the prognostic factors (age, Hb, platelet count, CD 38 expression and lymphocytes count) as in table 7, similarly El Desoukey NA et al, Sandes AF et al and Rodrigues et al (4, 5 and 68) .

Also there is no significant correlation between CD200 positive expression and clinical stage of the disease (p value 0.215) as in table 8.

Moreover There are no significant clinical association between CD200 expression and the state of remission (p value 0.336) as show in table 9, unfortunately there are no similar studies found to correlate these findings.

-CD43 there is no significant statistical correlation between CD43 expression and age, Hb, platelet count and lymphocytes count. While there is a significant correlation with presence of CD38 (which suggest poor prognostic value) (p value 0.009) as in table 7.

There is no significant association between CD43 and clinical stage of disease as in table 8, further studies are required to correlate CD43 with clinical stage.

There is significant correlation between CD 43 expression and state of remission, p value (0.019) so high percentage of patients 100%, 80% of patients achieved remission, not need treatment respectively show negative expression of CD43 as in table 9. so we conclude that CD 43 has negative expression in those with good prognostic history, this agreed with Ma XB et al that showed CD43 is an independent prognostic factor for DLBCL, NOS and other types of NHL that foresees poor prognosis, further studies are also recommended (26). further studies are required to verify our conclusions in other subtypes of NHL

CONCLUSION

From the present study, we may conclude the following:

1. CD 200 and CD43 expression and most commonly co-expression of both markers can significantly aid the diagnosis and discrimination of CLL from other B-LPDs.
2. CD200 has always moderate to bright expression in CLL while CD43 has dim to moderate expression.
3. CD200 and 43 expressions had no prognostic value for both CLL and NHL.
4. About 34.6% of CLL patients not need treatment.

REFERENCES

1. Chan JKC, Jaffe ES KY-H. WHO classification of tumors of hematopoietic and lymphoid tissues (Revised 4th edition) IARC. WHO classification of tumors of hematopoietic and lymphoid tissues (Revised 4th edition) IARC. 2017. 353–4 p.
2. Falay M, Öztürk BA, Güneş K, Kalpakçı Y, Dağdaş S, Ceran F, et al. The role of CD200 and CD43 expression in differential diagnosis between chronic lymphocytic leukemia and mantle cell lymphoma. *Turkish J Hematol.* 2018;35(2):94–8.
3. Hoffmann J, Rother M, Kaiser U, Thrun MC, Wilhelm C, Gruen A, et al. Determination of CD43 and CD200 surface expression improves accuracy of B-cell lymphoma immunophenotyping. *Cytom Part B - Clin Cytom.* 2020;98(6):476–82.
4. El Desoukey NA, Afify RAA, Amin DG, Mohammed RF. CD200 Expression in B-cell chronic lymphoproliferative disorders. *J Investig Med.* 2012;60(1):56–61.
5. Sandes AF, De Lourdes Chauffaille M, Oliveira CRMC, Maekawa Y, Tamashiro N, Takao TT, et al. CD200 has an important role in the differential diagnosis of mature B-cell neoplasms by multiparameter flow cytometry. *Cytom Part B - Clin Cytom.* 2014;86(2):98–105.
6. Iova A, Vlădăreanu A, Bumbea H, Begu M, Vasile D, Andruş E. CD 200 - A useful marker in chronic B lymphoproliferative disorders. *J Med Life.* 2012;5(Spec Issue):66–70.
7. Sorigue M, Juncà J, Sarrate E, Grau J. Expression of CD43 in chronic lymphoproliferative leukemias. *Cytom Part B - Clin Cytom.* 2018;94(1):136–42.
8. Meynard L. The inhibitory collagen receptor LAIR-1 (CD305). *J Leukoc Biol.* 2008;83(4):799–803.
9. Hoffbrand AV, Higgs D R, Keeling D M. *Postgraduate Haematology*, seventh edition, wiley blackwell, 2016, p 500–523.
10. Devilta VT, Laurence T S, Rosenberg SA. *Cancer principles and practices of oncology*, ninth edition, Lippincot, Williams and wilkins, 2011, p1973–1988.
11. Abdulridha RH, Jawad NK, Numan AT. Prevalence and risk of leukemia reported cases, observational descriptive statistic from Iraqi center for hematology in Baghdad province. *Indian J Forensic Med Toxicol.* 2021;15(1):2428–33.
12. GREER J P, Arber DA, Appelbaum FR, Wintrob clinical hematology, fourteen edition, Wolters Kluwer, 2019, 1888-1929.
13. Rawstron AC, Kreuzer KA, Soosapilla A, Spacek M, Stehlikova O, Gambell P, et al. Reproducible diagnosis of chronic lymphocytic leukemia by flow cytometry: An European Research Initiative on CLL (ERIC) & European Society for Clinical Cell Analysis (ESCCA) Harmonisation project. *Cytom Part B - Clin Cytom.* 2018;94(1):121–8.
14. Hallek M, Cheson BD, Catovsky D, Caligaris-Cappio F, Dighiero G, Döhner H, et al. Guidelines for the diagnosis and treatment of chronic lymphocytic leukemia: A report from the International Workshop on Chronic Lymphocytic Leukemia updating the National Cancer Institute-Working Group 1996 guidelines. *Blood.* 2008;111(12):5446–56.
15. Hussain I S, Mufti G J, *Advances in Malignant Hematology*, first edition, wiley- blackwell, 2011, p 211-227.
16. Hussain AMA, Lafta RK. Cancer trends in Iraq 2000–2016. *Oman Med J.* 2021 Jan 1;36(1):1–8.
17. Swerdlow SH, Campo E, Pileri SA, Lee Harris N, Stein H, Siebert R, et al. The 2016 revision of the World Health Organization classification of lymphoid neoplasms. *Blood.* 2016;127(20):2375–90.
18. Kaushansky, Lichtman M A, Prchal J Tet al. *Williams Hematology 9th Edition*, Mc Graw Hill, 2016, p1527-1551.
19. Dorfman DM, Shahsafaei A. CD200 (OX-2 membrane glycoprotein) expression in B cell-derived neoplasms. *Am J Clin Pathol.* 2010;134(5):726–33.
20. Longo D L, Fauci AS, BRAUNWALD E et al. *HARRISON'S Hematology and Oncology*, 17TH edition, McGraw-Hill Companies; 2010.182-204.
21. Gatter K C, Delsol G, Warnke R A, et al. *The Diagnosis of Lymphoproliferative Diseases*, second edition. Wiley- Blackwell; 2012. 90-94.
22. van den Brand M, Han J, van Krieken JM. Recognizing nodal marginal zone lymphoma: Recent advances and pitfalls. A systematic review. *Haematologica.* 2013;98(7):1003–13.
23. Bain B J, Clark D M, Wilkins B S. *BONE MARROW PATHOLOGY*, Fifth edition, Wiley-Blackwell, 2019, p 436-486.
24. Gorodetskiy VR, Probatova NA, Radenska-Lopovok SG, Ryzhikova NV, Sidorova YV, Sudarikov AB. Clonal relationship of marginal zone

- lymphoma and diffuse large B-cell lymphoma in Sjogren's syndrome patients: case series study and review of the literature. *Rheumatol Int* [Internet]. 2020;40(3):499–506. Available from: <https://doi.org/10.1007/s00296-019-04470-x>
25. PARTEC.Cy flow@cube:high performance multilaser flowcytometry analyser and cell sorter .article at www.partec.com/applications/healthcare.html accessed at December,10,2020.
 26. Ma XB, Zheng Y, Yuan HP, Jiang J, Wang YP. CD43 expression in diffuse large B-cell lymphoma, not otherwise specified: CD43 is a marker of adverse prognosis. *Hum Pathol*. 2015;46(4):593–9.
 27. Carvalheiro T, Garcia S, Pascoal Ramos MI, Giovannone B, Radstake TRDJ, Marut W, et al. Leukocyte Associated Immunoglobulin Like Receptor 1 Regulation and Function on Monocytes and Dendritic Cells During Inflammation. *Front Immunol*. 2020;11(August):1–13.
 28. Perbellini O, Falisi E, Giaretta I, Boscaro E, Novella E, Facco M, et al. Clinical significance of LAIR1 (CD305) as assessed by flow cytometry in a prospective series of patients with chronic lymphocytic leukemia. *Haematologica*. 2014;99(5):881–7.
 29. Innova biosciences.first for antibody labelling. Article at www.innovabiosciences.com/technical-guides.html accessed at December, 4, 2020.
 30. McCoy Jr J P. Basic principles of flow cytometry. *Hematol Oncol Clin N Am* 2002; 16: 229– 243.
 31. Sun T. *Flow Cytometry and Immunohistochemistry for Hematologic Neoplasms*. 1st Edition. Denver, Colorado: Lippincott Williams & Wilkins; 2008.
 32. Van Dongen JJM, Lhermitte L, Böttcher S, Almeida J, Van Der Velden VHJ, Flores-Montero J, et al. EuroFlow antibody panels for standardized n-dimensional flow cytometric immunophenotyping of normal, reactive and malignant leukocytes. *Leukemia*. 2012;26(9):1908–75.
 33. Bain B J, Lewis M. Preparation and staining methods for blood and bone marrow films. In: Bain B J, Bates I, Laffan M A et al editors. *Practical Haematology*. Philadelphia: Elsevier Limited; 2012.P: 57-68.
 34. D'Arena G, De Feo V, Pietrantonio G, Seneca E, Mansueto G, Villani O, et al. CD200 and Chronic Lymphocytic Leukemia: Biological and Clinical Relevance. *Front Oncol*. 2020;10(November).
 35. Morilla R, Morilla AM. Immunophenotyping by Flow Cytometry. In: Bain B J, Bates I, Laffan M A et al editors. *Practical Haematology*. Philadelphia: Elsevier Limited; 2017.P:330-346.
 36. Sorigue M. CD43 in the malignant flow cytometry laboratory in 2020. *Expert Rev Hematol* [Internet]. 2021;14(1):123–36. Available from: <https://doi.org/10.1080/17474086.2021.1856653>.
 37. Bain BJ, Clark DM, Wilkins BS. *Bone Marrow Pathology*, fourth edition. West Sussex:Wiley- Blackwell;2010.54-99 .
 38. Swerdlow SH, Campo E, Harris NL, et al. WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues. Franc 2008; IARC, Lyon.
 39. Stevenson M S, Yuan C M. Flow cytometry.In: Erber W N, editor. *Diagnostic Techniques in Hematological Malignancies*. The Edinburgh Building, Cambridge CB2 8RU, UK; 2010.p.51-70.
 40. Alapat D, Coviello-Malle J, Owens R, Qu P, Barlogie B, Shaughnessy JD, Lorschach RB. Diagnostic usefulness and prognostic impact of CD200 expression in lymphoid malignancies and plasma cell myeloma. *Am J Clin Pathol* 2012;137:93–100.
 41. Dunphy CH. Applications of Flow Cytometry and Immunohistochemistry to Diagnostic Hematopathology. *Arch Pathol Lab Med* 2004 ; 128 : 1004-1022.
 42. Olteanu H, Harrington AM, Kroft SH. Immunophenotypic stability of CD200 expression in plasma cell myeloma. *Am J Clin Pathol* 2012;137:1013–1014.
 43. Olteanu H, Harrington AM, Hari P, Kroft SH. CD200 expression in plasma cell myeloma. *Br J Haematol* 2011;153:408–411.
 44. Palumbo GA, Parrinello N, Fargione G, Cardillo K, Chiarenza A, Berretta S, Conticello C, Villari L, Di Raimondo F. CD200 expression may help in differential diagnosis between mantle cell lymphoma and B-cell chronic lymphocytic leukemia. *Leuk Res* 2009;33:1212– 1216.
 45. Tonks A, Hills R, White P, Rosie B, Mills KI, Burnett AK, Darley RL. CD200 as a prognostic factor in acute myeloid leukaemia. *Leukemia* 2007;21:566–568.
 46. Moreaux J, Hose D, Reme T, Jourdan E, Hundemer M, Legouffe E, Moine P, Bourin P, Moos M, Corre J, et al. CD200 is a new prognostic factor in multiple myeloma. *Blood* 2006;108:41
 47. Costa ES, Pedreira CE, Barrena S, Lecrevisse Q, Flores J, Quijano S, Almeida J, del Carmen Garcia-Macias M, Bottcher S, Van Dongen JJ, et al. Automated pattern-guided principal component analysis vs expert-based immunophenotypic classification of B-cell chronic lymphoproliferative disorders: A step forward in the standardization of clinical immunophenotyping. *Leukemia* 2010;24:1927–1933.
 48. Brunetti L, Di Noto R, Abate G, et al. CD200/OX2, a cell surface molecule with immuno-regulatory function, is consistently expressed on hairy cell leukaemia neoplastic cells. *Br J Haematol*. 2009;145:665-667.
 49. Pillai V, Pozdnyakova O, Charest K, et al. CD200 flow cytometric assessment and semiquantitative immunohistochemical staining distinguishes hairy cell leukemia from hairy cell leukemia-variant and other B-cell lymphoproliferative disorders. *Am J Clin Pathol*. 2013;140:536-543.
 50. McWhirter JR, Kretz-Rommel A, Saven A, et al. Antibodies selected from combinatorial libraries block a tumor antigen that plays a key role in immunomodulation. *Proc Natl Acad Sci U S A*. 2006;103:1041-1046.
 51. Kohnke T, Wittmann VK, Bucklein VL, et al. Diagnosis of CLL revisited: increased specificity by a modified five-marker scoring system including CD200. *Br J Haematol*. 2017;179(3):480-487 .
 52. Quijano S, López A, Rasillo A, Sayagués JM, Barrena S, Sánchez ML, Teodosio C, Giraldo P, Giralto M, Pérez MC, Romero M, Perdiguero L, Orfao A. Impact of trisomy 12, del(13q), del(17p), and del(11q) on the immunophenotype, DNA ploidy status, and proliferative rate of leukemic B-cells in chronic lymphocytic leukemia. *Cytometry B Clin Cytom* 2008;74:139-149.
 53. Kostopoulos IV, Paterakis G, Papadimitriou K, Pavlidis D, Tsitsilonis OE, Papadimitriou SI. Immunophenotypic analysis reveals heterogeneity and common biologic aspects in monoclonal B-cell lymphocytosis. *Genes Chromosomes Cancer* 2015;54:210-221.
 54. Kretz-Rommel A, Bowdish KS. Rationale for anti-CD200 immunotherapy in B-CLL and other hematologic malignancies: new concepts in blocking immune suppression. *Expert opinion on biological therapy*. 2008;8(1):5-15.
 55. El Din Fouad NB, Ibrahim NY, Abdel Aziz RS, Ibrahim SK. CD200 expression in diagnostic and prognostic assessment of mature B cell lymphoproliferative neoplasms. *Asian Pac J Cancer Prev* (2018) 19:3383–92. doi: 10.31557/APJCP.2018.19.12.338
 56. Wang X, Zhang Z, Liu Y, Wang L, Yuan H, Xie P, et al. Expression of CD200 in the bone marrow of chronic lymphocytic leukemia patients and its correlations with clinical prognosis. *Chin J Cell Mol Immunol* (2014) 30:75–8.
 57. Miao Y, Fan L, Wu Y-J, Xia Y, Qiao C, Wang Y, et al. Low expression of CD200 predicts shorter time-to treatment in chronic lymphocytic leukemia. *Oncotarget* (2016) 7:13551–62. doi: 10.18632/oncotarget.6948.
 58. Jung G, Eisenmann JC, Thiébault S, et al. Cell surface CD43 determination improves diagnostic precision in late B-cell diseases. *Br J Haematol*. 2003;120(3):496–9.
 59. Zahedi M, Saeede S, Khorasani K, Nasirzadeh A, Abouali R, Abounoori M, et al. Promising Role of CD200 Expression in Diagnosis and Prognosis of Chronic Lymphocytic Leukemia: A Review. *Int J Med Invest* [Internet]. 2019;8(4):16–22. Available from: <http://www.intjmi.com>.
 60. Hammad R. Role of Leukocyte Associated Immunoglobulin Like Receptor-1 (CD305) in Predicting Clinical Variables of Chronic Lymphocytic Leukemia. 2020;1(March).
 61. Poggi A, Catellani S, Bruzzone A, Caligaris-Cappio F, Gobbi M, Zocchi MR. Lack of the leukocyte-associated Ig-like receptor-1 expression in high-risk chronic lymphocytic leukaemia results in the absence of a negative signal regulating kinase activation and cell division. *Leukemia*. 2008; 22(5):980-8.
 62. Thornton PD, Fernandez C, Giustolisi GM, Morilla R, Atkinson S, A'Hern RP, et al. CD38 expression as a prognostic indicator in chronic lymphocytic leukaemia. *Hematol J* 2004; 5:145–151.
 63. Ahmady I A, Alhassanin SA, Moussa NS. Role of CD305 and CD38 in Chronic Lymphocytic Leukemia Clinical Relevance. *Menoufia Med J*. 2018;31(2):564.
 64. van der Vuurst de Vries AR, Clevers H, Logtenberg T, Meyaard L. Leukocyte-associated immunoglobulin-like receptor-1 (LAIR1) is differentially expressed during human B cell differentiation and inhibits B cell receptor-mediated signaling. *Eur J Immunol*. 1999;29(10):3160-7.
 65. Pedraza-Alva G, Rosenstein Y. CD43: One molecule, many tales to recount. *Signal Transduct* 2007;7:372-85.
 66. Smal C, Lisart S, Maerevoet M, et al. s2-Chloro-2_-deoxyadenosine (CdA) in the B-cell leukemia cell line EHEB. *Biochem Pharmacol*. 2007;73:351Y358.

67. Jares P, Colomer D, Campo E. Genetic and molecular pathogenesis of mantle cell lymphoma: perspectives for new targeted therapeutics. *Nat Rev Cancer*. 2007;7:750Y762.
68. Rodrigues CA, Gonçalves MV, Ikoma MRV, et al (2016). Diagnosis and treatment of chronic lymphocytic leukemia: recommendations from the Brazilian Group of Chronic Lymphocytic Leukemia. *Rev Bras Hematol Hemoter*, 38, 346-57.
69. Mitrovic Z, Ilic I, Nola M, et al. CD43 expression is an adverse prognostic factor in diffuse large B-cell lymphoma. *Clin Lymphoma Myeloma* 2009;9:133-7.
70. Misawa Y, Nagaoka H, Kimoto H, et al. CD43 expression in a B cell lymphoma, WEHI 231, reduces susceptibility to G1 arrest and extends survival in culture upon serum depletion. *Eur J Immunol* 1996;26:2573-81.
71. Challagundla P, Medeiros LJ, Kanagal-Shamanna R, Miranda RN, Jorgensen JL. Differential expression of cd200 in b-cell neoplasms by flow cytometry can assist in diagnosis, subclassification, and bone marrow staging. *Am J Clin Pathol*. 2014;142(6):837-44.
72. Wong KK, Khatri I, Shaha S, et al. The role of CD200 in immunity to B cell lymphoma. *J Leukoc Biol*. 2010;88:361- 372.
73. Chuksina JJ, Kataeva E V, Mitina TA. Features of immunophenotypic finding B-cell lymphoproliferative diseases by flow cytometry. *Kazan Med J*. 2020;101(1):145-52.
74. Sales MM, Ferreira SIACP, Ikoma MRV, Sandes AF, Beltrame MP, Bacal NS, Silva MCA, Malvezzi M, Lorand-Metze IGH, Orfao A, Yamamoto M. Diagnosis of Chronic Lymphoproliferative Disorders by Flow Cytometry Using Four-Color Combinations for Immunophenotyping: A Proposal of the Brazilian Group of Flow Cytometry (GBCFLUX). *Cytometry Part B* 2017; 92B: 398-410.
75. Benedetti D, Shabani M, Perini C, Tissino E, Bo MD, Bulian P, et al. The B-Cell Receptor Signaling Inhibitor Molecules CD305 and CD307b Are Markers of Favorable Prognosis in Chronic Lymphocytic Leukemia with Both Mutated and Unmutated IGHV Gene Status. *Blood* [Internet]. 2016;128(22):4358. Available from: [http://dx.doi.org/10.1182/blood.V128.22.4358.43582016;128\(22\):4358](http://dx.doi.org/10.1182/blood.V128.22.4358.43582016;128(22):4358). Available from: <http://dx.doi.org/10.1182/blood.V128.22.4358.4358>