

Advanced approaches of the use of circRNAs as a replacement for cancer therapy

Goran Sedeeq Hama Faraj^a, Bashdar Mahmud Hussen^{b,c,**}, Snur Rasool Abdullah^d,
Mohammed Fatih Rasul^e, Yasaman Hajjesmaeili^f, Aria Baniahmad^{g,***}, Mohammad Taheri^{g,h,*}

^a Department of Medical Laboratory Science, Komar University of Science and Technology, Sulaymaniyah, 46001, Iraq

^b Department of Biomedical Sciences, College of Science, Cihan University-Erbil, Erbil, Kurdistan Region, 44001, Iraq

^c Department of Clinical Analysis, College of Pharmacy, Hawler Medical University, Erbil, Kurdistan Region, 44001, Iraq

^d Medical Laboratory Science, Lebanese French University, Erbil, Kurdistan Region, 44001, Iraq

^e Department of Pharmaceutical Basic Science, Faculty of Pharmacy, Tishk International University, Erbil, Kurdistan Region, Iraq

^f Faculty of Health, York University, Toronto, ON, Canada

^g Institute of Human Genetics, Jena University Hospital, Jena, Germany

^h Urology and Nephrology Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

ARTICLE INFO

Keywords:

Cancer
Cancer therapy
CircRNAs
Challenges
Strategies

ABSTRACT

Cancer is a broad name for a group of diseases in which abnormal cells grow out of control and are characterized by their complexity and recurrence. Although there has been progress in cancer therapy with the entry of precision medicine and immunotherapy, cancer incidence rates have increased globally. Non-coding RNAs in the form of circular RNAs (circRNAs) play crucial roles in the pathogenesis, clinical diagnosis, and therapy of different diseases, including cancer. According to recent studies, circRNAs appear to serve as accurate indicators and therapeutic targets for cancer treatment. However, circRNAs are promising candidates for cutting-edge cancer therapy because of their distinctive circular structure, stability, and wide range of capabilities; many challenges persist that decrease the applications of circRNA-based cancer therapeutics. Here, we explore the roles of circRNAs as a replacement for cancer therapy, highlight the main challenges facing circRNA-based cancer therapies, and discuss the key strategies to overcome these challenges to improve advanced innovative therapies based on circRNAs with long-term health effects.

1. Introduction

Cancer continues to be one of the most challenging problems in modern medicine due to its complex molecular pathways and the ongoing evolution of therapeutic resistance [1]. There have been considerable advancements in the treatment of different cancer types using conventional cancer therapies like chemotherapy and radiation [2]. However, these methods frequently have serious side effects and have restrictions on the molecular triggers of cancer growth that they can target [3]. In recent years, circRNAs have attracted much attention as a fresh and promising approach to cancer treatment.

CircRNAs are novel-type non-coding RNA molecules that are covalently closed, single-stranded RNA molecules without 5'–3' end and poly

(A) tails. They are more stable as they resist exonuclease-mediated destruction compared to linear transcripts [4]. Structurally, circRNAs consist of exons or introns, and circRNAs with exons are frequently found in the cytosol, while circRNAs with introns are primarily found in the nucleus [5]. Linking the 3' downstream terminal with the 5' upstream terminal through a back-splicing process creates its circular structure [6]. This circular structure of circRNA helps it to be protected from destruction by exonucleases and makes circRNAs more stable. Therefore, the use of circRNA in the field of disease therapy, especially cancer therapy, is progressing in the right direction [7].

Numerous circRNAs have been found due to the advancement of bioinformatics techniques and high-throughput RNA sequencing (RNA-seq). For instance, in human fibroblasts, Jeck et al. observed over 25,000

* Corresponding author. Institute of Human Genetics, Jena University Hospital, Jena, Germany

** Corresponding author. Department of Biomedical Sciences, College of Science, Cihan University-Erbil, Erbil, Kurdistan Region, 44001, Iraq

*** Corresponding author.

E-mail addresses: Aria.baniahmad@med.uni-jena.de (B.M. Hussen), bashdar.hussen@hmu.edu.krd (A. Baniahmad), mohammad.taheri@uni-jena.de (M. Taheri).

<https://doi.org/10.1016/j.ncrna.2024.03.012>

Received 14 December 2023; Received in revised form 18 March 2024; Accepted 29 March 2024

Available online 30 March 2024

2468-0540/© 2024 The Authors. Published by KeAi Communications Co., Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

circRNAs that have not been broken down by exonucleases [8]. Similarly, in samples of juvenile acute lymphoblastic leukemia, spliced gene transcripts make up a large percentage of the circRNA map, as Salzman and his colleagues explored [9].

Synthetic circRNAs are being investigated for application in disease therapy, and recently, there has been an increase in interest in developing technology for their synthesis [10]. Synthetic circRNAs have also been used as biosensors and therapies, such as the replacement of therapeutic proteins and peptides [11] and vaccinations [12].

Despite their unique properties as potential cancer treatments, such as their circular structure and stability, several obstacles still stand in the way of their extensive therapeutic applications. This review highlights the functions of circRNAs as an alternative for cancer therapy, identifies the most critical challenges plaguing circRNA-based cancer therapies, and addresses the essential techniques to overcome these challenges to enhance advanced innovative therapeutics based on circRNAs with long-term health impacts.

2. CircRNA biogenesis and characterization

CircRNAs are created through non-canonical splicing processes called back-splicing, which is classified as an alternative splicing process. Cellular spliceosomal machinery is necessary to synthesize circRNAs in humans and animals [13]. A back-splicing process joins an upstream 3' splice site with a downstream 5' splice site to make a single-strand, covalently closed-loop structure [14]. This is how most circRNAs are made from pre-mRNAs. Different circRNAs can be made from identical sequences using alternate back-splicing [10].

Despite extensive study over many years, the specific mechanism underlying circRNA synthesis remains unknown. According to their cycling mechanisms and composition, circRNAs are typically categorized into three kinds: exonic circRNAs (ecircRNAs) [15], intronic circRNAs (ciRNAs) [16], and exon-intron circRNAs (EicirRNAs) [17]. Exonic circRNAs are primarily found in the cytoplasm and have one or more exons, with two or three exons coming from alternative splicing [18]. There are many types of nuclear back-splicing and linear splicing processes that have been studied. Some of these are exon skipping,

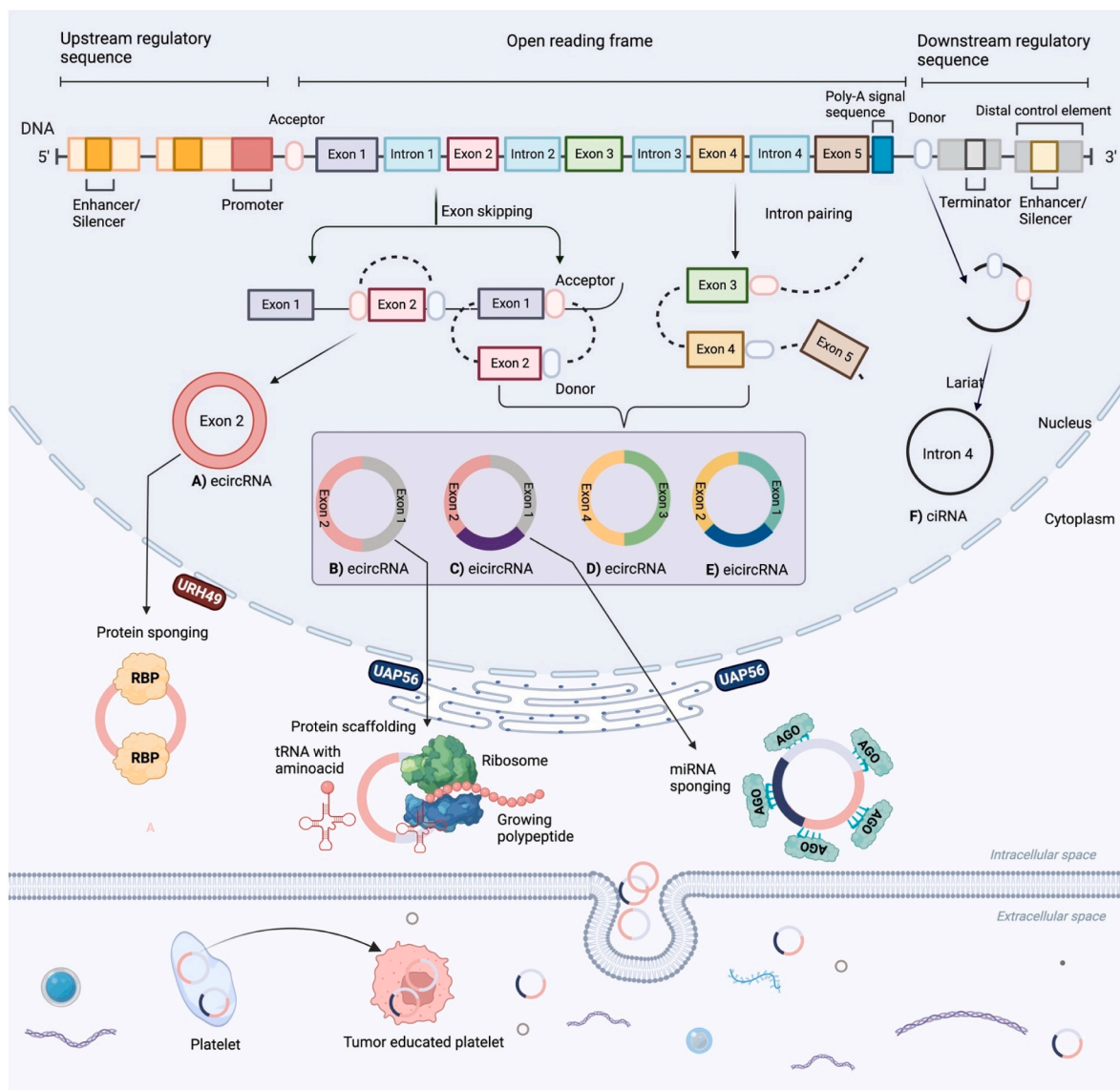


Fig. 1. Formation of circRNA and the three main hypotheses: an ecircRNA is produced when introns are removed through intron-pairing-driven circularization, also called direct back-splicing. In this process, the pre-flanking mRNA's intronic complementary sequences create a lariat through direct base pairing. A process of circularization known as exon-skipping or lariat-driven produces pre-ecircRNA by removing transcript introns. RBP-mediated circularization regulates circRNA production by managing activators and inhibitors. RBP RNA binding proteins, URH49 uridine-ribosyltransferase 49, tRNA transfer RNA, AGO Argonaute protein.

intron pairing, and RNA-binding proteins (RBPs) [19]. Recently, it has been discovered that a novel class of mitochondrial-encoded circRNAs (meccRNAs) acts as molecular chaperones to assist the mitochondrial entrance of nuclear-encoded proteins [20].

Three possible mechanisms for circRNA formation have been proposed: First, ecircRNAs are primarily formed through direct back-splicing, also known as intron-pairing-driven circularization, in which the pre-flanking mRNA's intronic complementary sequences form a lariat by directly base-pairing, which results in an ecircRNA upon intron removal [17]. Secondly, RNAs fold close to exons and connect the downstream splicing donor to the upstream splicing acceptor through 3', 5' phosphodiester bonds to create a lariat structure, secondary splicing, or intron splicing to make a loop, and then cut introns to make an exon-containing lariat. This is also known as exon-skipping. Furthermore, intron lariats with a branching point rich in cytosine and a 5' splice site rich in 7-nucleotide guanine uracil can produce ciRNAs without being impacted by debranching enzymes [21]. Lastly, in RBP-mediated circularization, the production of circRNA is significantly regulated by RBPs that act as managing activators or inhibitors (Fig. 1). RNA editing enzyme ADR1 stops the output of circRNA by directly inverting ALU repeats. This is mediated by A-to-I editing of RNA pairing circularized exons [22,23]. This reduces the complementarity and stability of intron-base pairing interactions [18]. Currently, numerous studies have revealed the aberrant circRNA expression patterns and their regulatory roles in the development and spread of cancer [24,25].

3. Innovative advances and therapeutic potential of circRNA-based cancer therapy

Single-stranded and covalently closed circRNA molecules were initially described in the genomes of virions by Sanger et al., in 1976 [26]. Later, in 1979, Hsu et al. proved circRNAs without free terminals and the need for companion protein [27]. In 1991, Nigro et al. became the first to describe how non-canonical splicing led to the unexpected isolation of isoforms from the deleted colon cancer gene ("scrambled exons") [28]. Further, Cocquerelle and his team presented consistent findings for the human EST-1 gene in 1992, and they established a link between the presence of many nearby introns and the synthesis of these transcripts [29]. They showed that the circularized RNAs remained stable in the cytoplasm over two days after actinomycin D treatment [30]. Later studies by Cape et al. found that the scrambled product of gene Sry RNA was circular; this circRNA is primarily intracellular,

tissue-specific, and present in three different mouse strains [31]. When these hypothetical steps have been completed, nuclear extracts have been shown in several studies to be useful for the *in vitro* generation of circRNAs, the target molecules for this chemical [32,33].

From 1996 to the beginning of the 2000s, scientists discovered that other human genes could make circRNAs. For example, cytochrome P450 2C24 in rats [34], cytochrome P450 in humans, androgen-binding protein in the rat [35], dystrophin in humans [36], and cyclin-dependent kinase 4 (INK4/ARF)-associated ncRNA were used to make other types of circRNA [37]. Even though these early studies proved that RNA molecules can fold back on themselves, their significance was not immediately recognized. Advances in RNA-seq technologies and specialized computational workflows prompted a new wave of circRNA research in 2010. Later, circRNAs were found to affect the actions of particular miRNAs and were subsequently used as a biomarker in diagnosing cancer such as lung cancer [38]. Recently, circRNAs were studied in clinical trials, such as circPUM1 targets in renal cell carcinoma tissue to sponge miRNA [39] (Fig. 2). These circular RNA molecules have proven to be capable of controlling gene expression, interacting with different cellular elements, and acting as valuable biomarkers. As this research field develops, circRNA-based therapeutics have the potential to offer cancer patients more effective and individualized therapy alternatives.

4. CircRNAs' potential roles in carcinogenesis

CircRNAs, a family of long non-coding RNAs, are involved in several critical biological processes that promote or inhibit cancer [40–42]. More evidence suggests that circRNAs play a crucial role through different mechanisms in several malignancies, including esophagus cancer (EC) [43], lung cancer (LC) [44], gastric cancer (GC) [45], breast cancer (BC) [46], and colorectal cancer (CRC) [47,48].

The physiological functions of circRNAs are mediated by the miRNA sponge in cancers. For instance, circBCAR3, a molecular sponge for miR-27a-3p, increases tumorigenesis and metastasis in EC patients [43]. Likewise, hsa-circ-0013958 has been upregulated in plasma and tissues of LC patients, and sponging miR-134 leads to elevated levels of cyclin D1, a known carcinogenic protein. Further, the hsa-circ-0013958 expression level was connected with lymphatic metastasis and TNM stage [49]. Moreover, overexpression of hsa_circRNA_102958 increases the proliferation of GC, although its expression level was associated with the TNM stage [50]. Similarly, hypoxia-inducible circWSB1 was

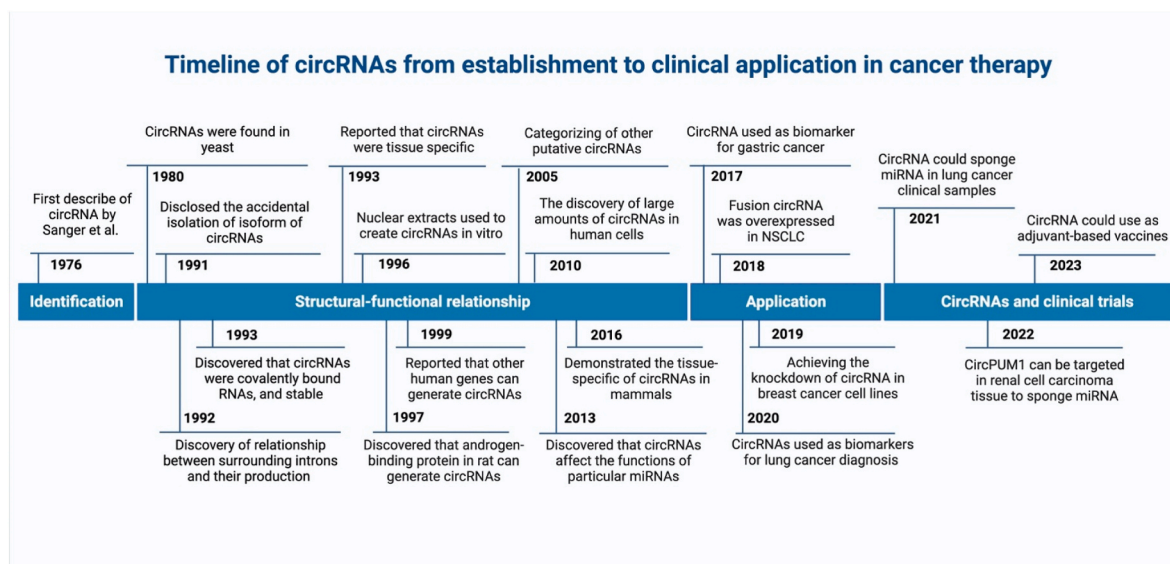


Fig. 2. A timeline outlining the most significant findings made about circRNA-based cancer therapy.

significantly upregulated and interacts with USP10 to reduce the stability of p53 caused by USP1 and promote the progression of BC tissues [46]. Furthermore, circTDRD3 was upregulated under hypoxic conditions and accelerates the progression and spread of CRC by affecting a positive feedback loop through the HIF1 α /PTBP1/circTDRD3/miR-1231/HIF1 α pathway [47]. The molecular mechanisms that lead to tumors progressing to a malignant state may be better understood in light of these results.

Consequently, the abundance of stable circRNAs constitutes a new class of RNA species that may distinguish between cancer cells and healthy cells, proving its significant potential as a circulating biomarker for diagnostic cancers. The exact role of circRNAs in carcinogenesis remains unknown despite recent progress in this field.

According to growing data, circRNAs may be employed as prospective genetic markers for diagnostic techniques, prognosis, early cancer recognition, and even therapy response monitoring. The following are the principal applications of circRNAs in cancer.

4.1. CircRNAs as a biomarker in cancer diagnosis and therapy

CircRNAs can serve as cancer biomarkers due to their dysregulation and association with cancer morphologies. CircRNAs have a high degree of tissue- and disease-specificity, making them a potential choice for cancer diagnostics [51] (Fig. 3). More evidence suggests that circRNAs are involved as a biomarker in diagnosing several cancers. For instance, circ-ITCH acts as a sponge for miR-7/17/214 in ESCC, which may increase the level of ITCH circRNA. ITCH overexpression promotes the ubiquitination and degradation of phosphorylated Dvl2, which in turn limits the progression of ESCC by inhibiting the Wnt/ β -catenin axis [52]. Likewise, hsa_circ_0013958 up-regulates oncogenic cyclin D1 through sponging of miR-134, which plays an essential role in the progression of NSCLC. According to these findings, hsa_circ_0013958 might be

employed as a non-invasive biomarker for early diagnosing and screening LAC [49]. Moreover, Li et al. revealed that the sensitivity and specificity of identifying hsa_circ_0001649 between GC and normal samples are satisfactory. This means that it could be used as a biomarker for non-invasive screening of GC by making comparisons of the expression profiles in tissue and serum samples [53].

CircRNAs can act as a competitive endogenous RNA (ceRNA) to inhibit specific target genes in several types of cancer, which might be used as a diagnostic biomarker. For example, circGFRA1 acts as a ceRNA to control *GFRA1* production by sponging miR-34a in TNBC and could be used as a diagnostic biomarker and a potential target for TNBC treatment [54]. Similarly, circ-PDE8A regulates MACC1 and promotes invasive development via the MACC/MET/ERK or AKT pathways in PDAC by acting as a ceRNA for miR-338. It is suggested that exosomal circ-PDE8A could be a valuable diagnostic for PDAC diagnosis or prognosis, and circ-PDE8A plays a crucial role in tumor invasion [55]. These data suggest that tumor cell-derived circRNAs are released into the tumor milieu surrounding the circulation, supporting the use of circRNAs as biomarkers for patients with malignant tumors [56].

According to recent studies, circRNAs can be utilized to predict the metastasis of cancer cells. For instance, in a study comparing the circRNA regulation profiles of six patients, Xu et al. showed that circRNA 0001178 and circRNA 0000826 were considerably overexpressed in CRC metastatic tissue samples, allowing them to reliably identify between CRC and liver metastasis patients [57]. Moreover, circASAP1 was markedly upregulated in the HCC tissues of patients with lung metastasis after surgery. Downward regulation of circASAP1 was associated with decreased overall survival and an increased recurrence rate and was positively correlated with CSF1, MAPK1, and CD68⁺ tumor-associated macrophage levels [58]. Thus, circASAP1 could have a potential role in the prognosis of HCC metastasis.

Based on other studies, circRNAs are essential for predicting

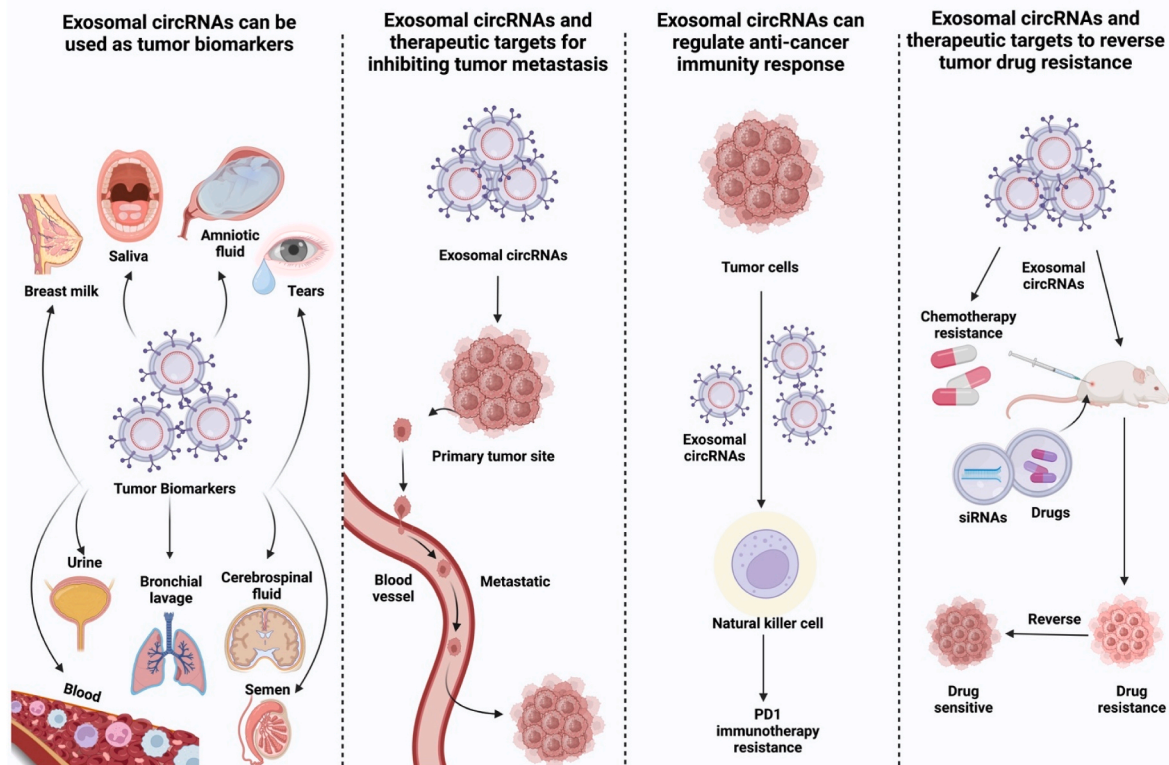


Fig. 3. Use of circular RNAs (circRNAs) as diagnostic and prognostic biomarkers and as possible therapeutic interventions for human diseases and disorders, including cancer, biological samples that contain circRNAs are tears, saliva, cerebrospinal fluid, bronchial lavage, breast milk, blood, amniotic fluid, urine, and semen.

therapeutic resistance in a specific type of cancer. For instance, cisplatin is one of the most effective chemotherapeutics for treating GC [59]. However, Huang et al. showed that circAKT3 upregulates *PIK3R1*, which promotes cisplatin resistance via sponging of miR-198, in 105 GC patients [60]. This clarified that circAKT3 is a highly reliable prognostic biomarker in GC patients, which makes them resistant to cisplatin. Similarly, circ_0076305 has been proven to increase *ABCC1* expression by sponging miR-186-5p, driving resistance to cisplatin in NSCLC [61]. CircRNAs are more specifically described in Table 1 as prognostic and diagnostic biomarkers in several cancer types.

Overall, developing novel, non-invasive, extremely precise, and accurate biomarkers for early cancer screening, diagnosis, therapy, and prognosis monitoring is necessary.

4.2. CircRNAs as inhibitors in cancer therapy

Almost every human malignancy has circRNAs that are involved in its pathogenesis. In our previous discussion, we established that circRNAs compete with target genes for miRNA-binding sites, allowing them to regulate gene expression in a manner similar to that of miRNA sponges [158]. Current research has shed light on how circRNAs, which function as miRNA inhibitors, regulate the migration and prognosis of cancer cells. For instance, circ-ZKSCAN1 inhibits BC progression by miR-1178-3p sponging through the miR-1178-3p/p21 axis and serves as a predictive factor for recurrence [159]. Similarly, Zhang et al. demonstrated that circRNA-104075 can inhibit miRNA-104075 in HCC tissue and serve as a new diagnostic biomarker [138]. Therefore, targeting circ 104075 might lead to novel HCC diagnosis and therapy approaches.

4.3. CircRNAs as therapeutic targets in cancer therapy

In the area of therapeutic targets, tumor-promoting, and tumor-suppressive circRNAs have the potential to be used as therapeutic targets in cancer. CircRNAs have a unique back-splicing junction sequence that allows them to be targeted selectively without modifying the parent mRNA. As a result, carcinogenic circRNAs can be a target for siRNAs and undergo Ago2-mediated destruction [160]. In contrast, tumor-suppressing circRNAs may be expressed ectopically utilizing expression vectors with lengthy reverse complementary sequences that allow the enhanced sequence to circulate [161]. A potential therapeutic approach could be to target circRNAs or associated pathways, as their dysregulation can lead to cancer.

Several biomolecules, such as proteins, shRNA, and siRNA, inhibit or enhance circRNAs. For instance, Fang et al. demonstrate that circAGO2 is enhanced with HuR protein and overexpressed in several malignant tissue samples. Stimulation of circAGO2 led to boosted growth, invasion, and metastasis in cell culture and animal models [162]. In contrast to stimulation, the inhibition of circAGO2 by shRNA spurred tumor growth in vivo since circAGO2 activity may be controlled by the inhibition of circAGO2 and HuR response; this can be accomplished with the assistance of a HuR suppressor peptide delivery [163]. Moreover, Zhang et al. investigated that circCUX1 could significantly be overexpressed and enhanced tumor cell growth, invasion, and migration by inhibiting miR-16-5p in a neuroblastoma sample, while inhibited circCUX1 with shRNA lowered tumor proliferation in a mouse model [164].

In addition, antisense oligonucleotides have also been applied to downregulate circular RNAs. For example, in colorectal cancer (CRC), the connection between circLONP2 and DDX1 led to the overexpression of cells that produce metastases, which facilitated invasion and propagation. Conversely, the in vivo downregulation of circLONP2 by antisense oligonucleotides prevented CRC metastases [165]. Furthermore, in patients with acute myeloid leukemia, the expression of circ_0004277 was reduced after chemotherapy treatment but increased again when patients relapsed after full remission, demonstrating the relationship between upregulation and treatment success [166].

Moreover, according to the findings of the study conducted by Fang and his colleagues, it was proposed that the administration of FUS-mediated circ_0002099 might potentially enhance the EMT process in the progression of BLCA by means of the miR-217-5p/miR-103a-3p/KRAS axis-WNT/ β -catenin axis. Additionally, it has the potential to serve as a therapeutic target and a promising predictive biomarker for BLCA [167]. Similarly, CiRS-7 is a promising biomarker for predicting the prognosis of CRC patients, and it has the potential to be used as a therapeutic target for lowering the activity of EGFR-RAF1 in CRC patients [168]. Additionally, in a patient-derived CRC xenograft model, Yang and his team demonstrated that the injection of shRNA into the tail vein precisely targets circPTK2 and reduces the spread of tumor metastases. Thus, they identified that circPTK2 is a promising biomarker for early diagnosis of CRC metastasis and provides a potential therapeutic target for CRC [169].

Recently, there are developing in clinical research that are focusing on various elements of circRNAs as therapeutic targets in cancer therapy. These studies are focusing on circRNA expression patterns, functional roles, and therapeutic interventions [170,171]. For example, in ovarian cancer, Zhu et al. demonstrated that circNUP50 was increased in DDP-resistant OC cells [172]. This would allow for the formation of platinum-resistant conditions in OC by promoting cell proliferation, changing the cell cycle, and reducing apoptosis. Thus, they proposed that circNUP50 is a new therapeutic target that modifies p53 ubiquitination to induce cisplatin resistance in OC.

According to these results, the regulation of circular RNAs is not constant and varies with the severity of diseases, supporting the potential therapeutic roles of circRNAs in cancer treatment. Table 2 explains the mechanism of action and research sources for the various cancer types where circRNAs can be used as therapeutic targets.

4.4. CircRNAs as a vaccine for cancer therapy

Recently, artificial circRNA has been created to discover how they might be used as a novel class of mRNA therapeutics and vaccines. According to several studies, since regular mRNAs were translated to function as antigens, circRNAs can be used to express proteins [12]. Due to this, circRNAs and mRNAs are suitable candidates to produce vaccines. For instance, novel clinical research has demonstrated that the translation of proteins could be achieved by introducing mRNAs into the dendritic cells to be used as antigen-producing molecules to create vaccines for cancer immunotherapy [208]. This makes it possible to translate circRNAs into proteins, which would then generate functional antigens and activate CD8⁺ T cells to develop vaccines from cancer stem cells. According to Sun et al., mature DCs with CD133⁺ hepatocellular carcinoma stem cell RNA can trigger vitro cytotoxic T lymphocyte responses against hepatocellular cancer stem cells [209]. Furthermore, when 9L glioma stem-like cell RNA was injected into dendritic cells, the tumor's growth was dramatically inhibited, and the survival rate of rats with gliomas increased [210].

Likewise, circRNAs can act as tumor antigens or vaccine adjuvants that stimulate innate and adaptive immune responses. According to Chen et al., the intracellular sensor retinoic-acid-inducible gene-I regulated the immune response to exogenous circRNAs, and transfecting HeLa cells in vitro to produce circRNA may trigger innate immunity gene expression [211]. Exogenous unmodified circRNA adjuvant might trigger antigen-specific T and B cell responses by interacting with and activating RIG-1 [212]. Li et al. showed that the RNA vaccine's stability was effectively increased by circularizing linear RNA molecules to produce stable circRNA molecules with long-lasting protein expression capabilities. Thus, by covering the antigen-coding circRNA in LNP to facilitate in vivo synthesis, scientists created a novel circRNA vaccination platform. This approach demonstrated superior anti-tumor effectiveness in many animal tumor models and was able to induce substantial innate and adaptive immune activation [12].

Despite the apparent benefits of circRNAs acting as novel vaccines,

Table 1
CircRNAs as potential indicators for cancer diagnosis and prognosis.

Type of cancer	circRNA	Source	Biomarker	Regulation	miRNA sponge	Mechanism	Refs
LC	hsa-circ-0013958	Tissue, plasma, cell line	Diagnostic in NSCLC	↑	miR-134	hsa-circ-0013958↑- miR-134↓-cyclin-D1↑	[49]
LC	hsa_circ_0075930	Cell line, tissue	Diagnostic in NSCLC	↑	miR-149-5p	–	[62]
LC	ciRS-7	Tissue	Prognostic biomarker in NSCLC	↑	miR-7	–	[63]
LC	circFARSA	Tissue, plasma	Diagnostic in NSCLC	↑	miR-330-5p and miR-326	–	[64]
LC	circPRMT5	Tissue, cell line	Diagnostic in NSCLC	↑	miR-377, miR-382, and miR-498	–	[65]
LC	circRNA100146	Tissue	Diagnostic	↑	miR-361-3p and miR-615-5p	circRNA100146↑- miR361-3p ↓/miR-615-5p↓- SF3B3↑	[66]
LC	circ_0005280	Tissue	Diagnostic	↓	–	Unknown	[67]
LC	circRNA_102231	Tissue	Diagnostic	↑	Unknown	Unknown	[68]
LC	circ-ITCH	Tissue	Diagnostic	↓	miR-7 and miR-214	circ-ITCH↓- miR-7↑/miR214↑-Wnt/β-catenin↑	[69]
LC	circPVT1	NSCLC tissue, cell line	Diagnostic	↑	miR-125b	circPVT1↑-miR-125b↓- E2F2 pathway↑	[70]
LC	circMET	MSCLC tissue, cell line	Diagnostic	↑	miR-145-5p	circMET↑-miR-145-5p↓- CXCL3↑	[71]
LC	circGFRA1	MSCLC tissue, cell line	Diagnostic	↑	miR-188-3p	circGFRA1↑-miR-188-3p↓- PI3K/AKT↑	[72]
LC	hsa_circ_0001946	Tissue, cell line	Diagnostic	↓	miR-135a-5p	hsa_circ_0001946↓-NER signaling pathway↑	[73, 74]
LC	hsa_circ_0030998	Tissue	Diagnostic	↓	miR –558	hsa_circ_0030998↓-miR –558↑-MMP1/MMP17↓	[75]
LC	circ-CCS	Tissue	Diagnostic	↑	miR-383	circ-CCS↑-miR-383↓- E2F7↑	[76]
LC	circ-IGF1R	Tissue, cell line	Diagnostic	↓	miR-1270	circ-IGF1R↓-miR-1270↑- VANGL2↓	[77]
LC	circRNA_102179	Tissue, cell line	Diagnostic	↑	miR-330-5p	circRNA_102179↑-miR -330-5p↓- HMGB3↑	[78]
LC	circ-ZKSCAN1	NSCLC sample, cell line	Diagnostic	↑	miR-330-5p	circ-ZKSCAN1↑-miR-330- 5p↓- FAM83A↑	[79]
LC	hsa_circ_0007059	Tissue	Diagnostic	↓	miR –378	hsa_circ_0007059↓-miR –378↑-Wnt/β-catenin↑/ERK1/2↑	[80]
LC	circ-PITX1	Tissue	Diagnostic	↑	miR-1248	circ-PITX1↑-miR-1248↓- CCND2↑	[81]
LC	circ_0000429	NSCLC tissue, cell line	Diagnostic	↑	miR-1197	circ_0000429↑-miR-1197↓- MADD↑	[82]
LC	circ_0001287	Tissue, cell line	Diagnostic	↓	miR-21	circ_0001287↓-miR-21↑- PTEN↓	[83]
LC	hsa_circ_0000064	Tissue, cell line	Diagnostic	↓	–	hsa_circ_0000064↑- caspase-3/9↑/ bax↑/p21↑/CDK6↑/cyclin D1↑/bcl-2↓/MMP-2/9↑	[84]
LC	circFGFR3	NSCLC tissue	Prognostic	↑	miR-22-3p	circFGFR3↑- miR-22-3p↓-Gal-1↑/p-AKT↑/p-ERK1/2↑	[85]
LC	circ_0003645	NSCLC tissue	Prognostic	↑	miR –1179	circ_0003645↑-miR –1179↓- TMEM14A↑	[86]
LC	CDR1as	Tissue	Prognostic	↑	miR-7	CDR1as↑-miR-7↓- EGFR↑/CCNE1↑/PIK3CD↑	[87]
LC	circ_POLA2	Tissue	Prognostic	↑	miR –326	circ_POLA2↑-miR –326↓-GNB1↑	[88]
LC	circ-FOXM1	Tissue	Prognostic	↑	miR -1304-5p	circ-FOXM1↑-miR -1304-5p↓-PPDPF↑/MACC1↑	[89]
LC	circPIP5K1A	Tissue, serum	Prognostic	↑	miR-600 and miR-101	circPIP5K1A↑- miR-600↓-HIF-1α↑	[90, 91]
LC	circRNA_010763	Tissue	Prognostic	↑	miR-715	circRNA_010763↑- miR-101↓-ABCC1↑	[92]
LC	circRNA_100876	Tissue	Prognostic	↑	Unknown	Unknown	[93]
LC	circ-ANXA7	Tissue	Prognostic	↑	miR-331	circ-ANXA7↑- miR-331↓-LAD1↑	[94]
LC	circ-PTEN	Tissue, serum	Prognostic	↓	miR –155 and miR-330-3p	circ-PTEN↓-miR –155↑/miR-330-3p↑- PTEN↓	[95]
LC	hsa_circ_0008003	Tissue	Prognostic	↑	miR-488	hsa_circ_0008003↑- miR-488↓- ZNF281↑	[96]
LC	circ-MTHFD2	Tissue	Prognostic	↑	Unknown	Unknown	[97]
GC	hsa_circ_0000745	Plasma	Diagnostic	↓	–	–	[98]
GC	hsa_circ_0000520	Tissue, plasma	Diagnostic	↓	–	–	[99]

(continued on next page)

Table 1 (continued)

Type of cancer	circRNA	Source	Biomarker	Regulation	miRNA sponge	Mechanism	Refs
GC	hsa_circ_0014717	Tissue	Diagnostic	↓	–	–	[100]
GC	hsa_circ_0000096	Tissue	Diagnostic	↓	–	–	[101]
GC	hsa_circ_0066444	Tissue	Diagnostic	↑	–	–	[102]
GC	hsa_circRNA_102958	Tissue	Diagnostic	↑	–	–	[50]
GC	circLARP4	Tissue	Prognostic	↓	miR-424-5p	circ-RNA-LARP4↓-miR-424-5p↑-LATS1↓	[103]
GC	hsa_circ_0001368	Tissue	Prognostic	↓	miR-6506-5p	hsa-circ-0001368↓-miR-6506↑-FOXO3↓	[104]
GC	circPVT1	Tissue	Prognostic	↑	–	–	[105]
GC	circNRIP1	Tissue	Prognostic	↑	miR-149-5p	circNRIP1↑- miR-149-5p↓-AKT1/mTOR↑	[106]
GC	circ-KIAA1244	Plasma	Prognosis	↓	–	–	[107]
GC	circAKT3	Tissue	Prognostic	↑	miR-198	circAKT3↑- miR-198↓-PIK3R1↑	[60]
GC	hsa_circ_0010882	Tissue	Prognostic	↑	–	hsa_circ_0010882↑- p-PI3K, p-Akt, and p-mTOR↑	[108]
GC	circ-ATAD1	Tissue	Prognostic	↑	miR-140-3p	circ-ATAD1↑-miR-140-3p↓-YY1↑	[109]
GC	circ-HIPK3	Tissue	Prognostic	↑	miR-124/29b	–	[110]
GC	circ-DCAF6	Tissue	Prognostic	↑	miR-1231/1256	–	[111]
GC	circPVT1	Tissue	Prognostic	↑	miR-125	–	[105]
GC	circ-OSBPL10	Tissue	Prognostic	↑	miR-136-5p	–	[112]
GC	circ-NHSL1	Tissue	Prognostic	↑	miR-1306-3p	–	[113]
GC	circ-NRIP1	Tissue	Prognostic	↑	miR-149-5p	circ-NRIP1↑-miR-149-5p↓- AKT1/mTOR↑	[106]
GC	circ-LARP4	Tissue	Prognostic	↓	miR-424-5p	circ-LARP4 ↓-miR-424-5p↑-LATS1↓	[103]
GC	circ-LMTK2	Tissue	Prognostic	↓	–	–	[114]
GC	circ-PVRL3	Tissue	Prognostic	↓	–	–	[115]
GC	circ-CCDC9	Tissue	Prognostic	↓	miR-6792-3p	circ-CCDC9↓- miR-6792-3p↑-CAV1↓	[116]
CRC	Circ-VAPA	Plasma	Diagnostic	↓	miR-101	circVAPA↑- miR-101↑-	[117]
CRC	circITGA7	Tissue	Diagnostic	↓	miR-370-3p	hsa_circ_0026782↓-miR-370-3p↑-NF1↓	[118]
CRC	circHIPK3	Tissue	Prognostic	↑	miR-7	circHIPK3↑-miR-7↓-FAK, IGF1R, EGFR, YY1↑	[119]
CRC	ciRS-7	Tissue	Prognostic	↑	miR-7	–	[120]
CRC	circRNA_100290	Tissue, cell line	Prognostic	↑	miR-5166	circRNA_100290↑-miR-5166↓-Wnt/β-catenin↑	[121]
CRC	CircCCDC66	Tissue, cell line	Diagnostic	↑	–	–	[122]
CRC	hsa-circ-0136666	Tissue, cell line	Prognostic and diagnostic	↑	miR-136	–	[123]
CRC	hsa-circ-0001649	Tissue, serum	Diagnostic	↓	–	–	[124]
CRC	hsa-circ-0000711	Tissue, cell line	Diagnostic and prognostic	↓	–	–	[125]
CRC	hsa-circ-0014717	Tissue, cell line	Prognostic	↓	–	–	[126]
CRC	hsa-circ-0000567	Tissue, cell line	Diagnostic	↓	–	–	[127]
CRC	hsa-circ-001988	Tissue	Diagnostic	↓	–	–	[128]
CRC	hsa-circ-26344	Tissue	Prognostic	↓	miR-21, miR-31	–	[129]
CRC	hsa-circ-003906	Tissue, cell line	Diagnostic	↓	–	–	[130]
CRC	hsa-circ-104700	Tissue	Diagnostic	↓	–	–	[131]
BC	hsa_circRNA_002178	Tissue	Prognostic	↑	miR-328-3p	hsa_circRNA_002178↑- miR-328-3p↓-COL1A1↑	[132]
BC	circ_0005230	Tissue	Prognostic	↑	miR-618	circ_0005230↑-miR-618↓-CBX8↑	[133]
BC	circKIF4A	Tissue	Prognostic	↑	miR-375	circKIF4A↑-miR-375↓- KIF4A↑	[134]
BC	circGFRA1	Tissue	Prognostic	↑	miR-34a	circGFRA1↑-miR-34a↓- GFRA1↑	[54]
HCC	Circ-CDYL	Tissue	Diagnostic	↑	–	–	[135]
HCC	circSMARCA5	Plasma	Diagnostic	↓	–	–	[136]
HCC	hsa_circ_0000976, hsa_circ_0007750 and hsa_circ_0139897	Plasma	Diagnostic	↑	–	–	[137]
HCC	circ_104075	Tissue	Diagnostic	↓	miR-582-3p	circ_104075 ↑-miR-582-3p↓-YAP↑	[138]
HCC	circZKSCAN1	Tissue	Diagnostic	↓	–	–	[139]
HCC	circ-10720	Tissue	Prognostic	↑	–	–	[140]
HCC	circRNA 101368	Tissue	Prognostic	↑	miR-200a	circRNA 101368↑-miR-200a↓-HMGB1/RAGE↓	[141]
HCC	circRNA-100338	Tissue	Prognostic	↑	miR-141-3p	circRNA-100338↑-miR-141-3p↓-MTSS1↑	[142]
HCC	SCD-circRNA 2	Tissue	Prognostic	↑	–	–	[143]
HCC	circADAMTS13	Tissue	Prognostic	↓	miR-484	–	[144]
PC	circ-LDLRAD3	Plasma	Diagnostic	↑	–	–	[145]
PC	circ-PDE8A	Plasma	Prognostic	↑	miR-338	circ-PDE8A↑- miR-338↓-MET↑	[55]
PC	circ-IARS	Tissue	Prognostic	↑	miR-122	circ-IARS↑- miR-122↓- RhoA and RhoA-GTP↑	[146]

(continued on next page)

Table 1 (continued)

Type of cancer	circRNA	Source	Biomarker	Regulation	miRNA sponge	Mechanism	Refs
PCa	CircTENM3	Tissue	Diagnostic	↓	miR-558	CircTENM3↓- miR-558↓- RUNX3↑	[147]
PCa	circFGFR ^{1int2}	Cell line/ Tissue	Diagnostic	↑	miR-4687-5p	circFGFR ^{1int2} ↑- miR-4687-5p↓-P65/ FUS↑	[148]
PCa	circROBO1	Cell line/ Tissue	Diagnostic	↑	miR-556-5p	circROBO1↑- miR-556-5p ↓- PGK1↑	
PCa	circ_0086722	Cell line/ Tissue	Prognostic	↑	miR-339-5p	circ_0086722↑-miR-339-5p↓-STAT5A↑	[149]
PCa	circDDIT4	Tissues and serum samples	Diagnostic	↑	ELAVL1/HuR	circDDIT4↑-ELAVL1/HuR↓ -ANO7↓	[150]
PCa OC	circWHSC1	Tissue	Prognostic	↑	miR-145 and miR-1182	circWHSC1↑- miR-145 and miR-1182↓- MUC1 and hTERT↑	[151]
OC	circPLEKHM3	Tissue	Prognostic	↓	miR-9	circPLEKHM3↓-miR-9↓ AKT1↓	[152]
BCa	cTFRC	Tissue	Prognostic	↑	miR-107	cTFRC↑- miR-107↓- TFRC↑	[153]
BCa	circMTO1	Tissue	Prognostic	↑	miR-221	–	[154]
Osteosarcoma	circPVT1	Serum	Diagnostic	↑	–	–	[155]
Glioma	circ_0034642	Tissue	Prognostic	↑	miR-1205	circ_0034642↑-miR-1205↓-BATF3↑	[156]
Nasopharyngeal carcinoma	CDR1as	Biopsy	Prognostic	↑	miR-7-5p	CDR1as↑-MiR-7-5P↓-E2F3↑	[157]

RCC renal cell carcinoma, LC lung cancer, GC gastric cancer, CRC colorectal cancer, BC breast cancer, HCC hepatocellular carcinoma, PC pancreatic cancer, OC ovarian cancer, NSCLC non-small cell lung cancer, BCa bladder cancer, SF3B3 splicing factor 3b subunit 3, PI3K phosphoinositide 3-kinase, NER nucleotide excision repair, MMP1 matrix metalloproteinase 1, MMP17 matrix metalloproteinase 17, HMGB3 high-mobility group box 3, FAM83A family with sequence similarity 83 member A, ERK1 extracellular signal-regulated kinase 1, PTEN phosphatase and tensin homolog, Bcl-2 B-cell lymphoma 2 protein, TMEM14A transmembrane protein 14A, GNB1 guanine nucleotide-binding protein subunit beta-1, EGFR epidermal growth factor receptor, PIK3CD phosphatidylinositol 3-kinase catalytic subunit delta, PPDPF protein phosphatase 1D magnesium-dependent delta isozyme, MACC1 metastasis-associated in colon cancer 1, ABCC1 ATP-binding cassette subfamily C member 1, LAD1 ladinin 1, ZNF281 zinc finger protein 281, LATS1 large tumor suppressor kinase 1, FOXO3 forkhead box O3, CBX8 chromobox protein homolog 8, YAP yes-associated protein, HMGB1 high-mobility group box 1, RAGE receptor for advanced glycation end products, MTSS1 metastasis suppressor 1, TFRC transferrin receptor, BATF3 basic leucine zipper ATF-like transcription factor 3.

circRNA investigations are still in their early stages; hence, all relevant clinical trials have concentrated on mRNA vaccines. Additionally, recombinant enzymes are still used to create circRNAs in vitro, which is expensive if circRNAs are needed for vaccines [213]. Thus, research on this model is necessary to clarify circRNA's application as a vaccine for cancer therapy.

5. Challenges of the use of circular RNA as a cancer therapy and strategies to overcome

Although the previous description shows that circRNA is an effective technique, its discovery and implementation in human tumors make it a targeting method with various limits and concerns that make its use in clinical trials challenging. Low circRNA copy number, delivery method, controlling circRNA expression in vivo, and off-targeting are the significant concerns with circRNAs that have been identified (Fig. 4).

5.1. Low copy number of circRNA

One of the most crucial metrics to shed light on the efficacy of circRNAs in cancer treatment is the quantity of circular RNA. If circRNAs are to be utilized as successful treatment strategies, it is crucial to identify their precise functional mechanism; however, this may be more challenging due to the low copy number of circRNA. For example, if a circRNA is numerous and contains a large number of miRNA binding sites, it may be able to function as an effective miRNA sponge [214]. However, if a circRNA has a low copy number, its sponging efficacy would be limited even if it has many miRNA binding sites. Similarly, a circRNA with a high abundance but few miRNA binding sites would have low sponging efficiency and is improbable to serve as a miRNA sponge [7]. Therefore, the copy quantity of circRNAs must be ascertained to conclude that they can act via protein binding or other processes such as miRNA sponging. The initial stage of circRNA-based therapeutic approaches should be considered the inadequate quantity of circRNA.

5.1.1. Strategy to overcome the low copy number of circRNA

CircRNA levels can be altered in particular tissues, organs, and animal systems [215]. An appropriate cancer therapy requires upregulating a specific gene expression through miRNA sponging, which depends on circRNA and miRNA interaction. For this reason, the circRNA to miRNA ratio should be relatively high as it can enhance the sponging of miRNA [7]. If this ratio is high, it shows the efficacy of circRNA as a molecule that can address gene dysregulation in cancer disease. However, a low ratio makes clinical applications less efficient or inefficient [7].

Genetic manipulation in cancer therapy has been significantly enhanced with the development of genome editing tools such as ZFN, TALEN, and CRISPR-Cas programs. As illustrated, CRISPR-Cas9 has made several genetic applications possible, like promoter editing for generating quantitative trait variation and its widespread use in knocking out protein-coding genes [216] (Fig. 5).

5.2. Delivery challenges of circRNA in cancer therapy

Delivery of therapeutic agents is a fundamental and essential step in cancer therapy, and any limitation in this system leads to a decrease in the efficiency of the cancer therapy strategy. In cancer therapy, the delivery of therapeutic molecules is challenging because of the increase in immunological activation, bio-incompatibility, and ineffectiveness [217,218]. Several practical and trustworthy techniques can address this limitation, such as lipid nanoparticle or exosome-based delivery.

5.2.1. Strategies to overcome challenges of circRNA delivery in cancer therapy

Many approaches and strategies have been proposed, but unfortunately, the growing new side effects limit their delivery efficiency in the treatment of cancer. Here, we demonstrate two feasible techniques that enable therapeutic approaches to work efficiently.

5.2.2. Lipid nanoparticle-based delivery of circRNA

Nanoparticles can carry drugs and deliver them to therapeutic targets [219], and their potential uses in therapeutic medications, molecular delivery imaging, and drug combinations that combine diagnostic

Table 2
CircRNAs as therapeutic targets in cancer.

Type of cancer	circRNAs	Source	Regulation	miRNA sponge	Mechanism	Ref.
LC	circUBAP2	Clinical sample, cell line	↑	miR-3182	circUBAP2↑-miR-3182↓-KLF4↑	[173]
LC	circRNA_102231	Tissue	↑	–	Unknown	[68]
LC	hsa_circ_100395	Tissue	↓	miR-1228	hsa_circ_100395↓-miR-1228↑-TCF21↓	[174]
LC	circFADS2	Tissue, cell line	↓	miR-498	circFADS2↓-miR-498↓-HMGA2↑	[175]
LC	circRNA-FOXO3	NSCLC specimen	↓	miR-155	circRNA-FOXO3↓-miR-155↑-FOXO3↓	[176]
LC	circCDYL	NSCLC tissue, plasma, cell line	↑	miR-185-5p	circCDYL↓-miR-185-5p↑/TNRC6A↓-ERK1/2↑	[177]
LC	circSEC31A	–	↑	miR-376a	circSEC31A↑-miR-376a↓-SEC31A↑	[178]
LC	hsa_circ_0020123	Tissue	↑	miR-144	hsa_circ_0020123↑-miR-144↓-ZEB1↑/EZH2↑	[179]
LC	circ_0003998	–	↑	miR136-5p	circ_0003998↑-miR136-5p↓-CORO1C	[180]
LC	circCCDC66	NSCLC cell line, human bronchial epithelial cell line	↑	miR-33a-5p	circCCDC66↑-miR-33a-5p↓-KPNA4↑	[181]
LC	hsa_circ_0002874	Cell line	↑	miR-1273f	hsa_circ_0002874↑-miR-1273f↓-MDM2/P53↑	[182]
LC	hsa_circ_11780	NSCLC tissue, cell line	↓	miR-544a	hsa_circ_11780↓-miR-544a↑-FBXW7↓	[183]
LC	hsa_circ_0038646	Cells, tissue	↑	miR-331-3p	hsa_circ_0038646↑-miR-331-3p↓-GRIK3↑	[184]
LC	hsa_circ_0010235	NSCLC tissue, cell line	↑	miR-433-3p	hsa_circ_0010235↑-miR-433-3p↓-TIPRL↑	[185]
LC	hsa_circ_0001073	Lung cancer cells	↓	miR-626	hsa_circ_0001073↓-miR-626↑-LIFR↓	[186]
LC	circRNA_103993	NSCLC cells, human bronchial epithelial cell line	↑	miR-1271	circRNA_103993↑-miR-1271↓-ERG↑	[187]
LC	circP4HB	NSCLC tissue, cell line	↑	miR-133a	circP4HB↑-miR-133a5p↓-vimentin↑	[188]
LC	circARHGAP10	NSCLC tissue, cell line	↑	miR-150-5p	circARHGAP10↑-miR-150-5p↓-GLUT1↑	[189]
LC	circVANGL1	NSCLC tissue, cell line	↑	miR-195	circVANGL1↑-miR-195↓-Bcl-2↑	[190]
BC	CircDENND4C	Tissue, cell line	↑	miR-200b, miR-200c	Functions in low-oxygen environments as a miR-200b and miR-200c sponge.	[191, 192]
BC	CircFBXW7	Cell line	↓	miR-197-3p	Sponging miR-197-3p upregulates FBXW7 expression.	[193]
BC	CircTADA2A	Tissue, cell line	↓	miR-203a-3p	Activates miR-203a-3p, which in turn increases cytokine signaling regulator SOCS3 expression.	[194]
BC	CircWWC3	Tissue	↓	miR-26b-3p, miR-660-3p	Increases expression of EGFR, GRB2, PAK4, MAPK1, and AKT1 while acting as a sponge for miR-26b-3p and miR-660-3p (Ras signaling pathway)	[195]
BC	CircRNF20	Tissue, cell line	↑	miR-487a	Suppresses miR-487a, which targets HIF1α's 3' UTR.	[196]
BC	Circ-CDYL	Tissue	↑	miR-1275	targets ULK1 and ATG7 mRNAs linked to autophagy and autophagosomes, acting as a miR-1275 mimic.	[197]
CRC	circACAP2	Tissue, cell line	↑	miR-21-5p	T lymphoma invasion and metastasis protein 1 expression was suppressed by circACAP2 silencing, which increased the expression of miR-21-5p.	[198]
CRC	hsa_circ_0020397	CRC cells	↑	miR-138	The expression of the miR-138 targets TERT and PD-L1 showed that has circ_0020397 did reduce miR-138 activity.	[199]
PCa	circABCC4	PCa cells and tissue	↑	IGF2BP2	circABCC4↑-IGF2BP2↑-CCAR1↑	[200]
PCa	circRBM33	PCa cells and tissue	↑	ARSI	circRBM33↑-ARSI↓-PDHA1↑	[201]
CRC	hsa_circ_0055625	Tissue	↓	miR-106b-5p	circ_0055625↑-miR-106b-5p (miR-106b) ↓-ITGB8↑	[202]
CRC	hsa_circ_0000523	Tissue, cell line	↓	miR-31	hsa_circ_0000523↓-miR-31↑-Wnt/β-catenin signaling pathway↑	[203]
CRC	has_circ_103809	Tissue, cell line	↓	miR-532-3P	hsa_circRNA_103809↓-miR-532-3p↑-FOXO4 axis↑	[204]
RCC	circPUM1	Tissue	↑	miR-340-5p	circPUM1↑-miR-340-5p↓-FABP7↑	[39]
RCC	circ_0005875	Tissue	↑	miR-502-5p	circ-0005875↑-miR-502-5p↓-ETS1↑	[205]
RCC	hsa_circ_0054537	Tissue	↑	miR-130a-3p	hsa_circ_0054537↑-miR-130a-3p↓-cMet↑	[206]
RCC	circPTCH1	Tissue, cell line	↑	miR-485-5p	circPTCH1↑-miR-485-5p↓MMP14 and EMT↑	[207]

LC lung cancer, BC breast cancer, CRC colorectal cancer, RCC renal cell carcinoma, NSCLC non-small cell lung cancer, KLF4 kruppel-like factor 4, TCF21 transcription factor 21, HMGA2 high mobility group at-hook 2, FOXO3 forkhead box o-3, TNRC6A trinucleotide repeat containing adaptor 6A, ERK1 extracellular signal regulated kinase 1, SEC31A secretory pathway component 31A, ZEB1 zinc finger E-Box binding homeobox 1, EZH2 enhancer of zeste homolog 2, KPNA4 karyopherin alpha 4, MDM2 mouse double minute 2, P53 protein 53, FBXW7 F-box/WD repeat containing protein 7, GRIK3 glutamate receptor ionotropic kainite 3, LIFR leukemia inhibitory factor receptor, GLUT1 glucose transporter 1, Bcl-2 B-cell lymphoma 2, SOCS3 suppressor cytokine signaling 3, EGFR epidermal growth factor receptor, GRB2 growth factor receptor bound protein 2, PAK4 p21-activated kinase 4, MAPK1 mitogen-activated protein kinase 1, AKT1 protein kinase B, HIF1 α hypoxia inducible factor, ULK1 unc-51 like autophagy activating kinase 1, ATG7 autophagy-related 7, TERT telomerase reverse transcription, PD-L1 programmed cell death ligand 1, ITGB8 integrin subunit beta 8, FOXO4 forkhead box 4, FABP7 fatty acid binding protein 7, ETS1 E26 transformation-specific-1, MMP14 matrix metalloproteinase 14, EMT epithelial mesenchymal transition.

and therapeutic properties are actively being investigated. To target specific cells with either endogenous or foreign ligands, lipid nanoparticles (LNPs) are the most advanced nanoparticle carriers of small interfering RNA (siRNA), messenger RNA (mRNA), and circular RNA (circRNA) [220]. LNPs are nanoparticles that can deliver medications to disease areas for enhanced therapy efficacy [221]. In response to either internal or external signals, they can encapsulate circRNA and transport

it to a targeted cell population; however, upon uptake, their membranes can become unstable, enabling circRNA to escape into the cytosol and reach their targets [220]. Li et al. demonstrated their successful and efficient use of lipid nanoparticles to encapsulate and deliver circRNA into the aggressive tumor in mice in a trial [12]. Several potential applications for these nanoparticles are currently under study, including delivering diagnostic and therapeutic compounds [222]. Based on the

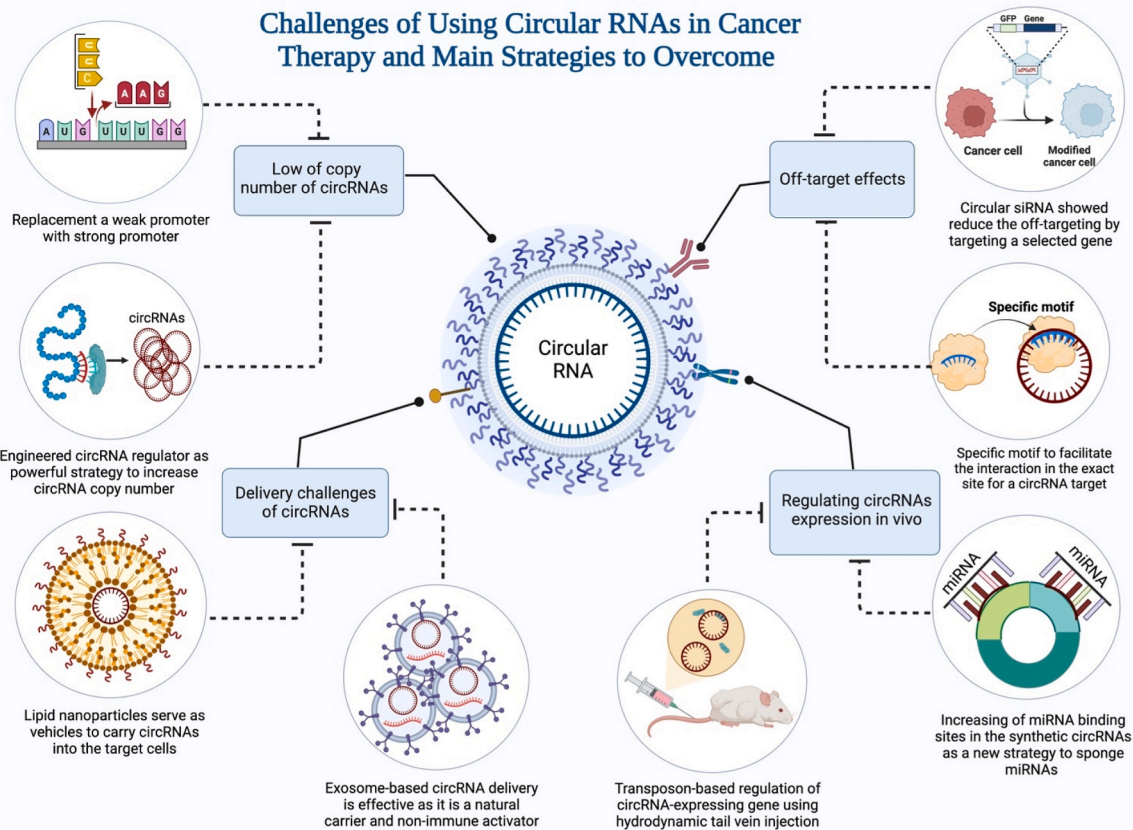


Fig. 4. The primary obstacles to employing circRNAs as therapeutic biomolecules and the methods to overcome them so that they can be utilized in cancer treatment as a potential replacement for traditional therapeutic biomolecules, which are currently less effective against most types of cancer.

results of these studies, lipid nanoparticles show significant promise as a delivery mechanism for circRNA-targeting therapeutics (Fig. 6a).

5.2.3. Gold nanoparticles (AuNPs) delivery

The use of nanoparticles as a delivery mechanism has significantly boosted the viability of circRNA-based therapeutics *in vivo*. Several studies have shown that gold nanoparticles (AuNPs) are a reliable delivery strategy in animals due to their great stability, purity, and easily adaptable surface [223]. For instance, in order to target the binding sites on circDnmt1 for Afl1 and p53 proteins, Du et al. administered AuNPs coupled with siRNA as a breast cancer therapeutic approach [224]. In addition, tumor growth was suppressed, and mouse survivability was prolonged by AuNP delivery of AONs inhibiting binding sites on circCcnb1 for Ccnb1 and Cdk1 [225]. According to this research, circRNA-targeting medicines may find a useful delivery method in nanoparticle form.

Moreover, circRNA expression plasmids have also been delivered *in vivo* via nanoparticles. Lu et al. found that the distribution of circEHMT1 plasmids via nanoparticles prevented the spread of breast cancer to the lungs in mice [226]. Similarly, the delivery of circFoxo3 plasmids with AuNP led to an increase in the rate of apoptosis in tumor cells and a reduction in the development of tumors [227].

Even though AuNPs effectively deliver circRNA targets in animal models, their safety in clinical settings remains largely unknown. Previous research has shown that AuNPs' hazardous effects are dependent on their size, with smaller AuNPs having more harmful effects [228]. As a result, AuNPs' characteristics can be adjusted to satisfy safety standards, offering circRNA-targeted medication delivery systems a bright future.

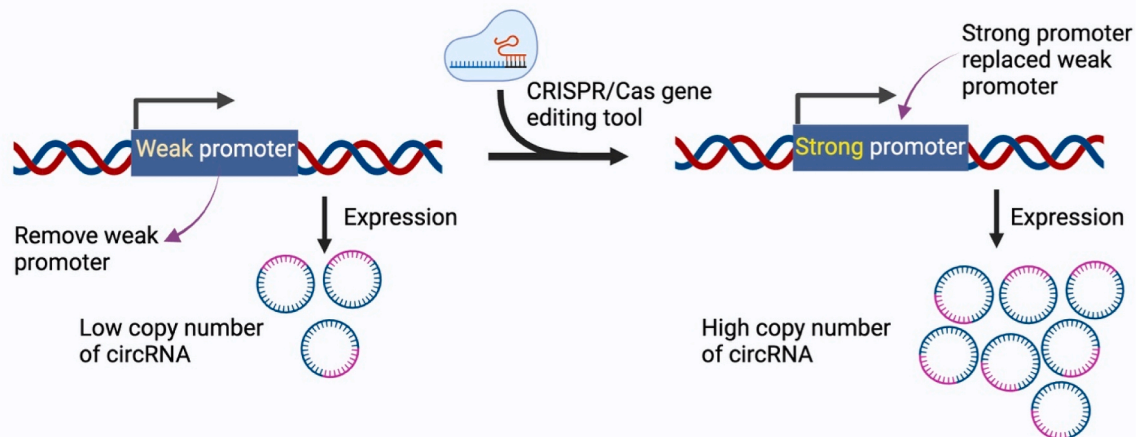
5.2.4. Exosome-based delivery of circRNA

Exosomes, also known as extracellular vesicles (EVs), can be found in nearly every cell, tissue, and bodily fluid. In disease pathobiology, they aid in intercellular signaling and keep tissues in a state of homeostasis [229,230]. Exosomal cargo contains 9769 proteins, 2838 microRNAs, 3408 messenger RNAs, and 1116 lipids, according to research [231].

Exosomes hold great promise as a therapeutic tool for various illnesses, including cancer, because they can effectively move tiny chemicals between cells [232]. Exosomes can be utilized to deliver treatments to cancer cells because of their unique characteristics, including their nano-size, double lipid membrane, capacity to serve as numerous carriers, good histocompatibility, high bioavailability, minimal cytotoxicity, and immunogenicity [233]. They can deliver a wide range of molecules, such as small interfering circRNAs (circRNAs) [4], microRNAs (miRNAs) [234], proteins [235], lipids [236], and DNA fragments [237] in their native state. Numerous studies have discussed the potential use of circRNAs as novel therapeutic targets and biomarkers in treating various illnesses, including cancer. Exo-circRNAs are circRNAs found in exosomes that can be found to support conventional diagnostic techniques and used to suppress the spread of malignancy [238].

Exosomal circRNA is taken by surrounding or distant cells and alters numerous physiological and pathological states, potentially boosting cell communications and spreading cancer. For the first time, in 2015, Li et al. discovered that exosomes are a rich source of circRNAs, and they were shown to be abundant in exosomes compared to parental cells, according to genome-wide RNA-seq investigations [239]. CircRNA sorting to exosomes can be controlled by adjusting parental cell levels of associated miRNA. This, in turn, can influence the biological activity of target cells, which are more stable and tolerant to their environment

A. Replace a weak promoter with a strong promoter to induce circRNA expression



B. Generating engineered circRNA regulators to induce circRNA expression

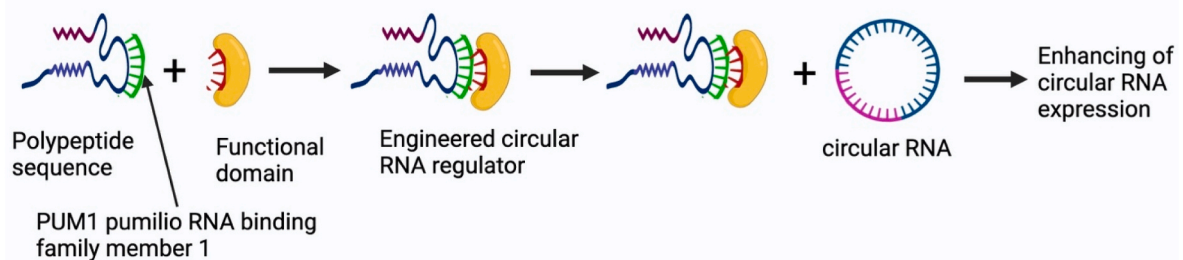


Fig. 5. shows the two strategies for increasing the copy number of circRNAs when used to treat cancer cells. The strategies are (A) replacing a weak promoter with a strong promoter by applying gene editing tools such as CRISPR/Cas technology to induce circRNA expression and (B) generating engineered circRNA regulators by using a gene encoding a member of the PUF family to induce circRNAs.

[240] (Fig. 6b).

This confirmed delivery ability of exosomes as carriers for circRNAs for cell-to-cell communication makes the exosomes more reliable and feasible carriers for circRNA delivery in cancer therapy-based circRNA strategy.

5.3. Regulating circRNA expression in vivo

To apply the effective therapeutic role of circRNAs in cancer, there should be approaches to regulate circRNA expression in vivo. Here, it can be concluded that new effective strategies can overcome the limitations of regulating circRNA expression in vivo.

5.3.1. Strategies to overcome regulating circRNA expression in vivo

Innovative genetic approaches for the sustained synthesis of prospective circRNAs hold great promise for controlling circRNA expression in vivo. For instance, transposons were employed to deliver a circRNA production cassette to the liver of mice by hydrodynamic tail vein injections (Fig. 7a). Mecozzi et al. inserted an expression cassette encoding the circRNA into the mouse genome using the recombination-mediated cassette exchange technique. Consequently, every mouse genome produces ectopic circRNA [241]. Further, to specifically promote circRNA expression in melanoma cells, researchers created a circRNA-expressing animal model of the disease using the same strategy [241]. These new methods and tools for functional analysis will alleviate some of the restrictions currently associated with circRNA expression systems.

Furthermore, circRNAs can be synthesized to increase their expression and functionality. The capacity of circRNA to act as a miRNA sponge is affected not only by the number of copies but also by the number of miRNA binding sites. To effectively sponge miRNAs, a synthetic circRNA with a high density of miRNA binding sites can be designed [242–244]. Therapeutic interventions may be possible through

this regulatory mechanism, which also has significance for gene expression (Fig. 7b).

Another new technology, clustered regularly interspaced short palindromic repeats (CRISPR), has entered the list of optimizers for circRNA regulation in vivo. Gao et al. used RNA-targeting type VI CRISPR effector RfxCas13 to suppress circRNA expression at the RNA level [245]. Similarly, Zhang et al. improved the strategy for creating CRISPR/Cas13 gRNAs with the ideal specificity and efficiency for reducing circRNA expression [246]. Additionally, Wang et al. explored essential circRNAs using optimized Cas13d technology and reported that knocking down of circRHOT1 leads to inhibition of HCC cell proliferation by depleting complementary sequences (CSs) [247] (Fig. 8).

Based on the aforementioned successful studies that control circRNA expression in vivo, this novel approach could propel circRNA-based cancer treatments forward and make them more viable. This will pave the way for other applications and the start of clinical trials.

5.4. Off-target effects in non-cancerous cells and tissues

While significant progress has been made in circRNA regulation in vivo, much more needs to be done before these technologies may be applied in clinical settings [248,249]. One obstacle is that a single circRNA molecule can function in a variety of tissues and diseases, for example, abnormal brain development [250], Parkinson's disease [251], myocardial infarction [252], diabetes [253], and femoral head necrosis [254], all have links with CDR1as. Similarly, circFoxo3 is implicated in the regulation of aging in the heart [255], as well as chemical resistance in prostate cancer [256]. Thus, using circRNA expression as a target for cancer therapy may have unexpected, adverse effects on other tissues or organs. To be a viable therapeutic option, circRNA-based therapies also need to increase therapeutic specificity and reduce off-target effects on other tissues or organs.

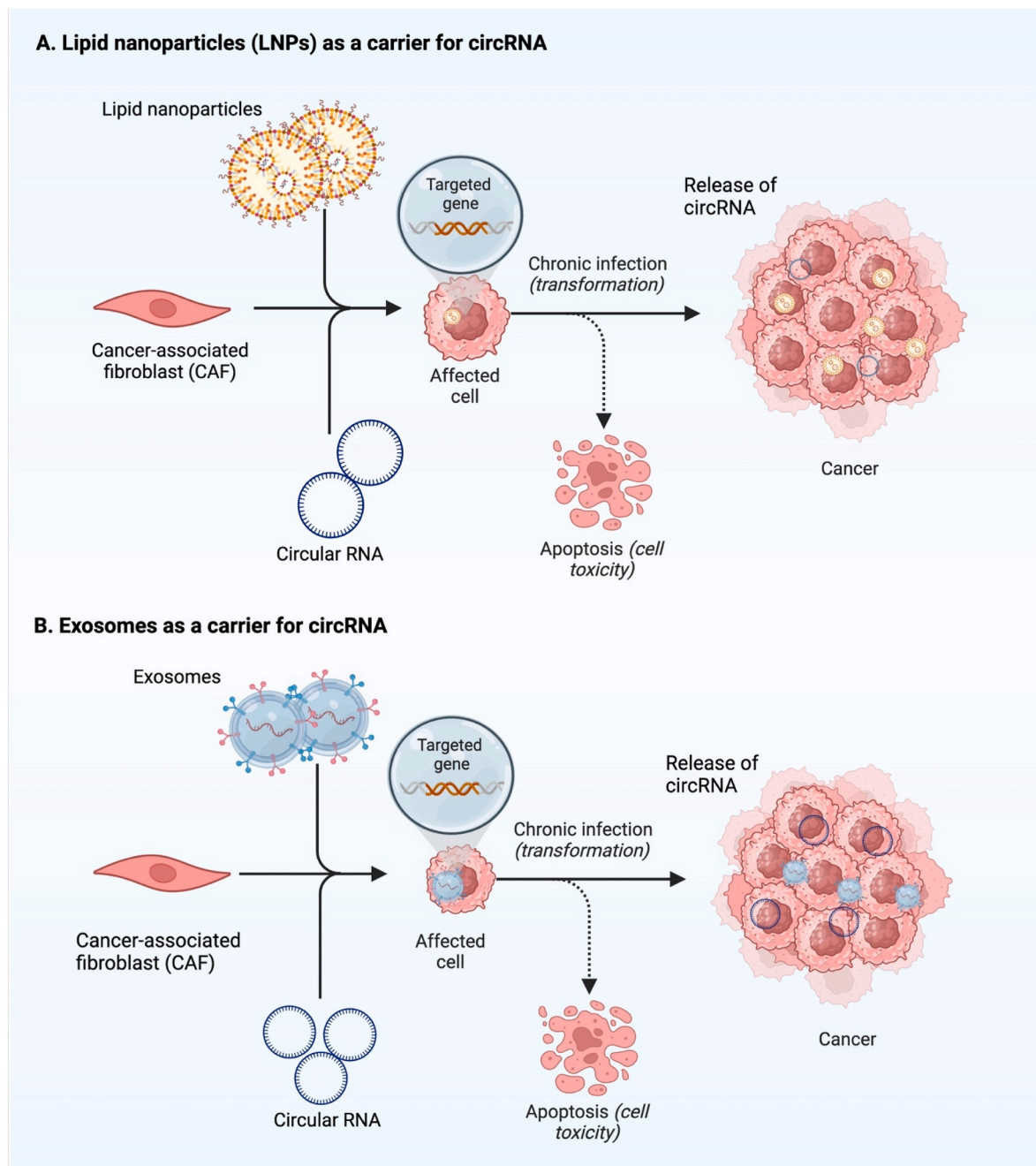


Fig. 6. Highlights the two practical approaches to address the limitations of the delivery method in delivering circRNAs for cancer treatment. (A) Lipid nanoparticles (LNPs) and (B) exosomes, both of which are carriers for circRNAs.

5.4.1. Strategies to overcome the circRNAs off-targeting effects

The base of the incorrect targeting in circRNA-based cancer therapy lacks a specific motif to facilitate the interaction in the exact site for a circRNA target. HEK293A cells were transfected with circular siRNAs targeting GFP or luciferase reporter genes with a circular sense and linear antisense strand. This allowed the siRNAs with the circular structural motif to effectively block the effects of sense strand RNA-induced off-target gene expression [257]. This discovery was made in response to the need to address the issue of circRNA off-targeting.

Circular siRNAs significantly reduced the off-target effect brought on by the sense strand of siRNA, as demonstrated by using RenillaXas, a rebuilt Renilla luciferase reporter gene. This new approach is significantly functional in reducing off-targeting in circRNA-based cancer therapy [257]. Moreover, off-target effects can be reduced to a minimum

through careful design of circRNA sequences. In order to improve the specificity of circRNAs and decrease the amount of non-specific binding to undesired targets, it is possible to optimize their length, secondary structure, and sequence composition [258]. Although, the stability and specificity of circRNAs can be improved through the introduction of chemical modifications, such as 2'-O-methyl or locked nucleic acid modifications. These alterations have the potential to either increase the affinity of circRNA for binding to target molecules or decrease the affinity for binding to non-specific targets [259,260]. Further, through the incorporation of cell-specific promoters into circRNA expression constructs, it is possible to restrict circRNA production to particular cell types, hence decreasing the impact of off-target expression in cells that are not the target. The utilization of this technique guarantees that circRNAs are preferentially expressed in the cell populations that are

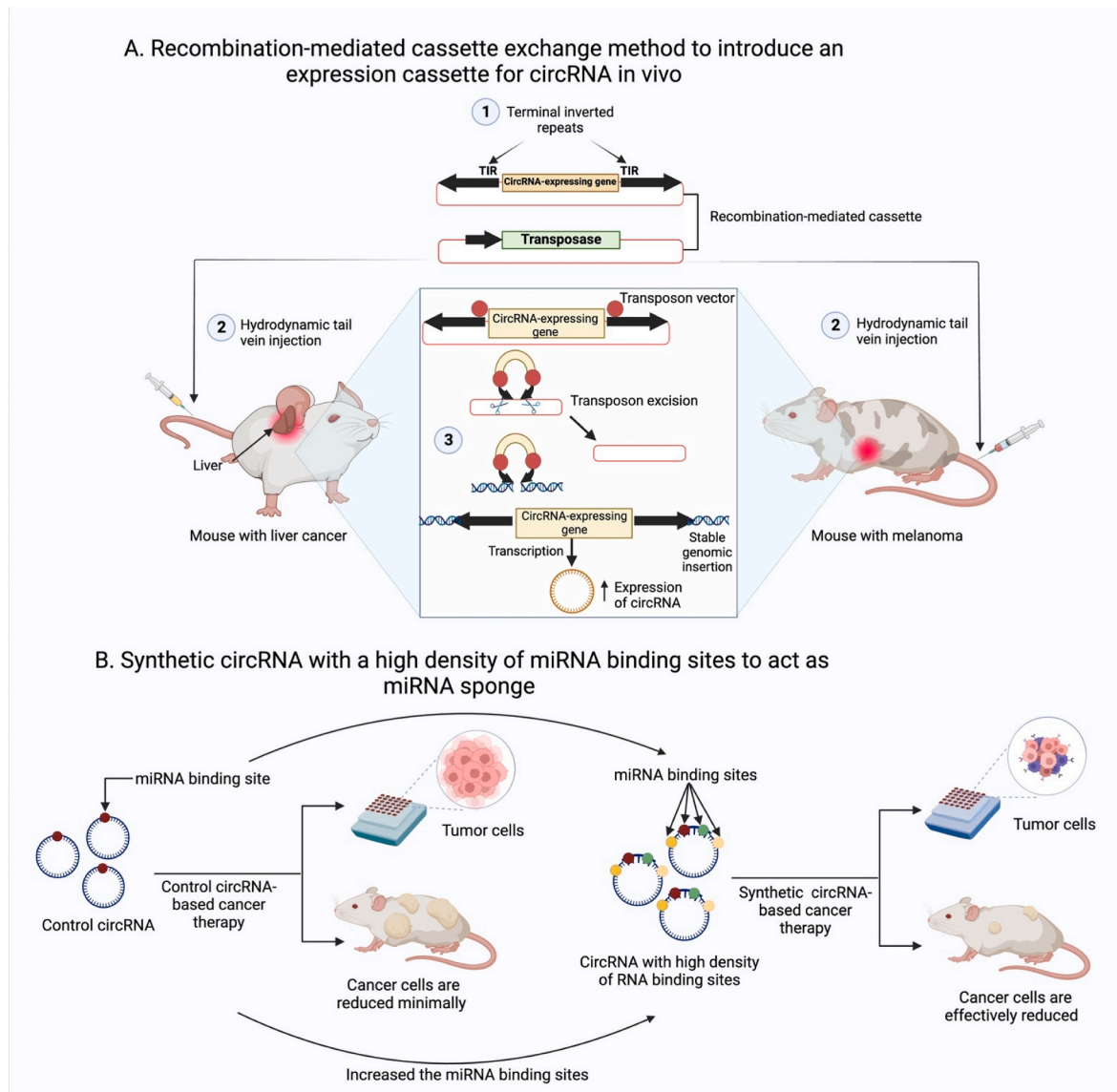


Fig. 7. Regulation of circRNA expression in vivo using new genetic methods for long-term production of potential circRNAs. Transposons are employed to deliver a circRNA production cassette to the livers of mice by hydrodynamic tail vein injections. At the same time, the recombination-mediated cassette exchange method is used to introduce an expression cassette for the circRNA into the mouse genome.

wanted, hence boosting therapeutic efficacy while simultaneously limiting off-target consequences [261]. Likewise, off-target effects can be reduced by selecting vector systems that are suitable for the delivery of circRNA. It is possible to increase circRNA localization and prevent unwanted interactions with off-target molecules by selecting vectors that have a high transfection efficiency and specificity for the target cell [262,263].

On the other hand, the CRISPR/Cas system is another developing and powerful strategy can be used to reduce off-target effects. For instance, using a machine learning methodology, Cheng et al. discovered that DeepCas13 effectively differentiates efficient sgRNAs from inefficient ones, and Cas13d's effect is linked to the guide's on-target cleavage effect in hepatocellular carcinoma [264]. Moreover, the CRISPR-dCas9 and CRISPR-Cas9 systems can also be used to demonstrate the function of this circular gRNA in vitro. Researchers have proposed a novel design for gRNA that might promote rapid circular gRNA assembly inside cells. The twister ribozymes are positioned on either side of the gRNA of interest in this circular pattern. It demonstrated a striking decrease in the off-target rate, together with decreased efficiency [265]. Furthermore, comparative investigations indicated that

RfxCas13-mediated circRNA knockdown exhibited significantly lower off-target effects on cognate mRNAs than shRNA/siRNA-mediated knockdown in mouse embryos [266]. Thus, the off-targeting effects of circRNAs during their usage in cancer therapy could be avoided or limited if the strategies listed above are used.

5.5. CircRNA-lncRNA sponge ceRNA interaction

Cancer study is greatly impacted by the complex mechanisms underlying the interactions between different ncRNAs, including lncRNAs and circRNAs. lncRNAs and circRNAs can interact and sponge miRNAs, which affects regular physiological functions and controls mRNA expression and, in turn, cell physiological state [267]. However, the significant implications for cancer may result from complex interactions between circRNAs and lncRNAs and brings therapeutic limitation. For instance, one of the primary mechanisms involves sponge regulation, where lncRNAs act as sponges for circRNAs or vice versa. These interactions can regulate gene expression precisely, which plays a role in the initiation and spread of cancer cells by sequestering miRNAs or other RNA-binding proteins [268].

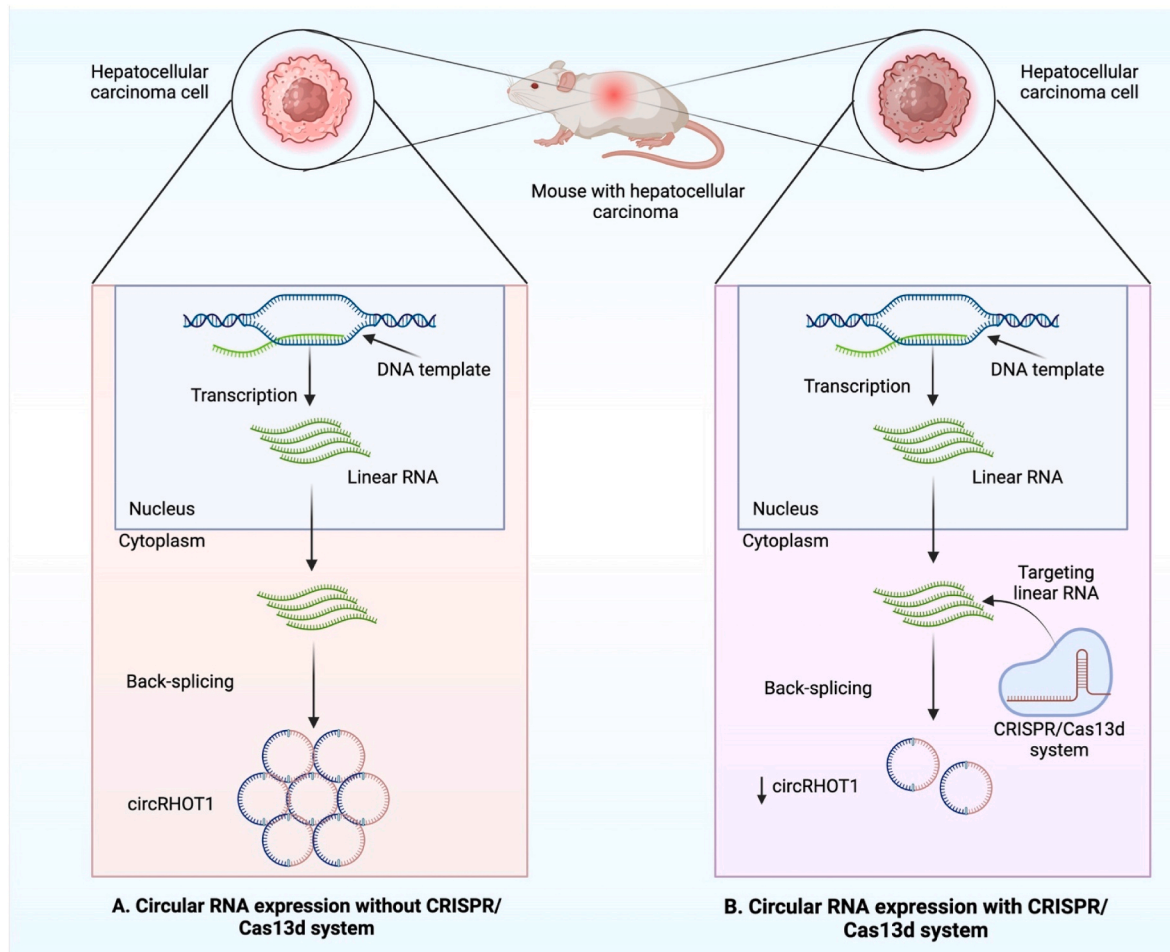


Fig. 8. Illustrates the expression of circular RNAs in an animal mouse model. (A) Overexpression of circRNA in mouse model hepatocellular carcinoma without CRISPR/Cas13. (B) Through the depletion of complementary sequences, the CRISPR/Cas13d technique-induced knockdown of oncogenic circRHOT1 in a mouse model inhibits the growth of HCC cells.

5.5.1. Strategies to overcome CircRNA-lncRNA interaction

The interplay between circRNAs and lncRNAs presents several challenges in the context of cancer therapy strategies. Creating synthetic RNA molecules that mimic the miRNA binding sites on the circRNAs or lncRNAs that mediate their interaction is one alternative [268]. These synthetic molecules may competitively bind to lncRNAs, preventing their interaction with the circRNA and disrupting the ceRNA regulatory network. Additionally, CRISPR/Cas-based genome editing technologies offer precision in targeting and disrupting the ceRNA interaction at the genetic level [269]. CRISPR/Cas systems can minimize the ceRNA effect by altering circRNA-lncRNA binding sequences, breaking the ceRNA connection. Further, accurate analysis and prediction of these interactions depend on sophisticated computer algorithms and bioinformatics tools. It is essential to achieve both specificity and selectivity when addressing circRNA-lncRNA sponge interactions to prevent any interference with regular cellular processes. Enhancing specificity can be accomplished by techniques such as creating tiny molecules or altered oligonucleotides that bind to target ceRNAs only while sparing non-targeted RNAs [270].

6. Conclusion and future perspectives

CircRNAs are new RNAs recently found in numerous species by high-throughput sequencing. CircRNAs play critical roles in cancer therapy and have been identified as possible attractive non-coding molecules. Their structural stability and resistance to RNase enzymes make them

promising diagnostics and therapeutic options.

CircRNAs are abundant in tumors; they bind to proteins, control transcription, and translation, and act as sponges for miRNAs. Therefore, circRNAs have applications in prognosis, drug-delivery design for target therapy, therapeutic response, and early diagnosis. Their remarkable properties and powerful functions have attracted the attention of researchers and clinicians in their study and potential therapeutic applications.

Despite advanced studies, several open concerns remain about the clinical use of circRNAs in cancer therapy. This study highlights various novel developments and therapeutic possibilities of circRNA-based cancer therapy. These include the utilization of circRNAs as a diagnostic and prognostic biomarker in cancer patients, the use of adjuvant-based vaccines with sponge oncogenic miRNAs, and the potential use of carcinogenic circRNAs as a therapeutic target for siRNAs. Further, it explores the main challenges of using circular RNA as a cancer therapy, like low copy numbers of circRNAs, delivery challenges, regulation expression, and off-targeting effects. We also highlighted the primary strategies to overcome these limitations.

Future studies should further investigate the efficacy and safety of using circRNA-based therapies. Critically, in vivo research is necessary to determine whether CRISPR/Cas technology is viable for efficient and specifically knocking down circRNAs. We assume that future discussions will revolve around methods to overcome the primary obstacles to using circRNAs in place of conventional cancer treatments and that additional research will be necessary to confirm the therapeutic efficacy of these

molecules.

Ethics approval and consent to participant

Not applicable.

Consent of publication

Not applicable.

Availability of data and materials

Not applicable.

CRedit authorship contribution statement

Goran Sedeeq Hama Faraj: Methodology, Investigation. **Bashdar Mahmud Husen:** Methodology, Investigation. **Snur Rasool Abdullah:** Methodology, Investigation. **Mohammed Fatih Rasul:** Resources, Investigation. **Yasaman Hajiesmaeili:** Methodology, Investigation. **Aria Baniahmad:** Methodology, Investigation. **Mohammad Taheri:** Writing – original draft, Supervision, Investigation.

Declaration of competing interest

The authors declare they have no conflict of interest.

Acknowledgement

Not applicable.

Funding

Not applicable.

Abbreviations

ATG7	Autophagy-related 7
BC	Breast cancer
CRC	Colorectal cancer
EGFR	Epidermal growth factor receptor
EMT	Epithelial-mesenchymal transition
ERK1	Extracellular signal-regulated kinase 1
EZH2	Enhancer of Zeste homolog 2
FABP7	Fatty acid binding protein 7
FOXO3	Forkhead box o-3
FOXO4	Forkhead box 4
GLUT1	Glucose transporter 1
GRB2	Growth factor receptor bound protein 2
GRIK3	Glutamate receptor ionotropic kainite 3
HMGA2	High mobility group at-hook 2
ITGB8	Integrin subunit beta 8
KLF4	Kruppel-like factor 4
KPNA4	Karyopherin alpha 4
LC	Lung cancer
LIFR	Leukemia inhibitory factor receptor
MAPK1	Mitogen-activated protein kinase 1
MDM2	Mouse double minute 2
MMP14	Matrix metalloproteinase 14
NSCLS	Non-small cell lung cancer
P53	Protein 53
PD-L1	Programmed cell death ligand 1
RCC	Renal cell carcinoma
SEC31A	Secretory pathway component 31A
SOCS3	Suppressor cytokine signaling 3
TCF21	Transcription factor 21

TERT	Telomerase reverse transcription
TNRC6A	Trinucleotide repeat containing adaptor 6A
ZEB1	Zinc finger E-Box binding homeobox 1

References

- [1] V. Mansouri, N. Beheshtizadeh, M. Gharibshahian, L. Sabouri, M. Varzandeh, N. Rezaei, Recent advances in regenerative medicine strategies for cancer treatment, *Biomed. Pharmacother.* 141 (2021) 111875.
- [2] J.-J. Wang, K.-F. Lei, F. Han, Tumor microenvironment: recent advances in various cancer treatments, *Eur. Rev. Med. Pharmacol. Sci.* 22 (12) (2018).
- [3] J. Nam, S. Son, K.S. Park, W. Zou, L.D. Shea, J.J. Moon, Cancer nanomedicine for combination cancer immunotherapy, *Nat. Rev. Mater.* 4 (6) (2019) 398–414.
- [4] B.M. Hussien, S.R. Abdullah, G.S. Hama Faraj, M.F. Rasul, A. Salihi, S. Ghafouri-Fard, et al., Exosomal circular RNA: a signature for lung cancer progression, *Cancer Cell Int.* 22 (1) (2022) 1–24.
- [5] X.O. Zhang, H.B. Wang, Y. Zhang, X. Lu, L.L. Chen, L. Yang, Complementary sequence-mediated exon circularization, *Cell* 159 (1) (2014) 134–147.
- [6] B. Xu, Y. Meng, Y. Jin, RNA structures in alternative splicing and back-splicing, *Wiley Interdiscip. Rev. RNA* 12 (1) (2021) e1626.
- [7] Circular RNAs in cancer: limitations in functional studies and diagnostic potential, in: F. Li, Q. Yang, A.T. He, B.B. Yang (Eds.), *Seminars in Cancer Biology*, Elsevier, 2021.
- [8] W.R. Jeck, J.A. Sorrentino, K. Wang, M.K. Slevin, C.E. Burd, J. Liu, et al., Circular RNAs are abundant, conserved, and associated with ALU repeats, *RNA* 19 (2) (2013) 141–157.
- [9] J. Salzman, C. Gawad, P.L. Wang, N. Lacayo, P.O. Brown, Circular RNAs are the predominant transcript isoform from hundreds of human genes in diverse cell types, *PLoS One* 7 (2) (2012) e30733.
- [10] S. Misir, N. Wu, B.B. Yang, Specific expression and functions of circular RNAs, *Cell Death Differ.* 29 (3) (2022) 481–491.
- [11] R.A. Wesselhoeft, *Synthetic Circular RNA for Protein Expression*, Massachusetts Institute of Technology, 2019.
- [12] H. Li, K. Peng, K. Yang, W. Ma, S. Qi, X. Yu, et al., Circular RNA cancer vaccines drive immunity in hard-to-treat malignancies, *Theranostics* 12 (14) (2022) 6422.
- [13] R. Liu, Y. Ma, T. Guo, G. Li, Identification, biogenesis, function, and mechanism of action of circular RNAs in plants, *Plant Communications* (2022) 100430.
- [14] Y. Liu, X. Ao, W. Yu, Y. Zhang, J. Wang, Biogenesis, functions, and clinical implications of circular RNAs in non-small cell lung cancer, *Mol. Ther. Nucleic Acids* 27 (2022) 50–72.
- [15] Y. Huang, Y. Li, W. Lin, S. Fan, H. Chen, J. Xia, et al., Promising roles of circular RNAs as biomarkers and targets for potential diagnosis and therapy of tuberculosis, *Biomolecules* 12 (9) (2022) 1235.
- [16] X. Wang, J. Zhang, G. Cao, J. Hua, G. Shan, W. Lin, Emerging roles of circular RNAs in gastric cancer metastasis and drug resistance, *J. Exp. Clin. Cancer Res.* 41 (1) (2022) 1–13.
- [17] R. Liu, L. Zhang, X. Zhao, J. Liu, W. Chang, L. Zhou, et al., circRNA: regulatory factors and potential therapeutic targets in inflammatory dermatoses, *J. Cell Mol. Med.* 26 (16) (2022) 4389–4400.
- [18] Y. Zeng, Y. Zou, G. Gao, S. Zheng, S. Wu, X. Xie, et al., The biogenesis, function and clinical significance of circular RNAs in breast cancer, *Cancer Biology & Medicine* 19 (1) (2022) 14.
- [19] L.S. Kristensen, M.S. Andersen, L.V. Stagsted, K.K. Ebbesen, T.B. Hansen, J. Kjems, The biogenesis, biology and characterization of circular RNAs, *Nat. Rev. Genet.* 20 (11) (2019) 675–691.
- [20] X. Liu, X. Wang, J. Li, S. Hu, Y. Deng, H. Yin, et al., Identification of meircRNAs and their roles in the mitochondrial entry of proteins, *Sci. China Life Sci.* 63 (10) (2020) 1429–1449.
- [21] Q. Wang, Y. Sun, Q. Zhao, W. Wu, L. Wang, Y. Miao, et al., Circular RNAs in pulmonary hypertension: emerging biological concepts and potential mechanism, *Animal Models and Experimental Medicine* 5 (1) (2022) 38–47.
- [22] K. Nishikura, Functions and regulation of RNA editing by ADAR deaminases, *Annu. Rev. Biochem.* 79 (2010) 321–349.
- [23] H. Shen, O. An, X. Ren, Y. Song, S.J. Tang, X.-Y. Ke, et al., ADARs act as potent regulators of circular transcriptome in cancer, *Nat. Commun.* 13 (1) (2022) 1508.
- [24] D. Xiong, R. He, Y. Dang, H. Wu, Z. Feng, G. Chen, The latest overview of circRNA in the progression, diagnosis, prognosis, treatment, and drug resistance of hepatocellular carcinoma, *Front. Oncol.* 10 (2021) 608257.
- [25] L. Yu, H. Zhu, Z. Wang, J. Huang, Y. Zhu, G. Fan, et al., Circular RNA circFIRRE drives osteosarcoma progression and metastasis through tumorigenic-angiogenic coupling, *Mol. Cancer* 21 (1) (2022) 1–20.
- [26] H.L. Sanger, G. Klotz, D. Riesner, H.J. Gross, A.K. Kleinschmidt, Viroids are single-stranded covalently closed circular RNA molecules existing as highly base-paired rod-like structures, *Proc. Natl. Acad. Sci. USA* 73 (11) (1976) 3852–3856.
- [27] M.-T. Hsu, M. Coca-Prados, Electron microscopic evidence for the circular form of RNA in the cytoplasm of eukaryotic cells, *Nature* 280 (5720) (1979) 339–340.
- [28] J.M. Nigro, K.R. Cho, E.R. Fearon, S.E. Kern, J.M. Ruppert, J.D. Oliner, et al., Scrambled exons, *Cell* 64 (3) (1991) 607–613.
- [29] C. Cocquerelle, P. Daubersies, M.-A. Majerus, J.-P. Kerckaert, B. Bailleul, Splicing with inverted order of exons occurs proximal to large introns, *EMBO J.* 11 (3) (1992) 1095–1098.
- [30] C. Cocquerelle, B. Mascrez, D. Hétiuin, B. Bailleul, Mis-splicing yields circular RNA molecules, *Faseb. J.* 7 (1) (1993) 155–160.

- [31] B. Capel, A. Swain, S. Nicolis, A. Hacker, M. Walter, P. Koopman, et al., Circular transcripts of the testis-determining gene *Sry* in adult mouse testis, *Cell* 73 (5) (1993) 1019–1030.
- [32] Z. Pasman, M.D. Been, M.A. Garcia-Blanco, Exon circularization in mammalian nuclear extracts, *RNA* 2 (6) (1996) 603.
- [33] C. Schindewolf, S. Braun, H. Domdey, In vitro generation of a circular exon from a linear pre-mRNA transcript, *Nucleic Acids Res.* 24 (7) (1996) 1260–1266.
- [34] P.G. Zaphiropoulos, Circular RNAs from transcripts of the rat cytochrome P450 2C24 gene: correlation with exon skipping, *Proc. Natl. Acad. Sci. USA* 93 (13) (1996) 6536–6541.
- [35] P.G. Zaphiropoulos, Exon skipping and circular RNA formation in transcripts of the human cytochrome P-450 2C18 gene in epidermis and of the rat androgen binding protein gene in testis, *Mol. Cell Biol.* 17 (6) (1997) 2985–2993.
- [36] A. Surono, Y. Takeshima, T. Wibawa, M. Ikezawa, I. Nonaka, M. Matsuo, Circular dystrophin RNAs consisting of exons that were skipped by alternative splicing, *Hum. Mol. Genet.* 8 (3) (1999) 493–500.
- [37] C.E. Burd, W.R. Jeck, Y. Liu, H.K. Sanoff, Z. Wang, N.E. Sharpless, Expression of linear and novel circular forms of an *INK4/ARF*-associated non-coding RNA correlates with atherosclerosis risk, *PLoS Genet.* 6 (12) (2010) e1001233.
- [38] Y.-H. Luo, Y.-P. Yang, C.-S. Chien, A. Yarmishyn, A. Ishola, Y. Chien, et al., Plasma level of circular RNA *hsa_circ_0000190* correlates with tumor progression and poor treatment response in advanced lung cancers, *Cancers* 12 (2020) 1740.
- [39] F. Zeng, L. Luo, M. Song, D. Li, Silencing of circular RNA *PUM1* inhibits clear cell renal cell carcinoma progression through the *miR-340-5p/FABP7* axis, *J. Recept. Signal Transduction* 42 (2) (2022) 141–150.
- [40] X. Zang, J. Jiang, J. Gu, Y. Chen, M. Wang, Y. Zhang, et al., Circular RNA *EIF4G3* suppresses gastric cancer progression through inhibition of β -catenin by promoting δ -catenin ubiquitin degradation and upregulating *SIK1*, *Mol. Cancer* 21 (1) (2022) 1–16.
- [41] M. Taheri, S. Najafi, A. Basiri, B.M. Hussien, A. Baniahmad, E. Jamali, et al., The role and clinical potentials of circular RNAs in prostate cancer, *Front. Oncol.* 11 (2021).
- [42] S. Ghafouri-Fard, B.M. Hussien, M. Taheri, S.A. Ayatollahi, Emerging role of circular RNAs in breast cancer, *Pathol. Res. Pract.* 223 (2021) 153496.
- [43] Y. Xi, Y. Shen, D. Wu, J. Zhang, C. Lin, L. Wang, et al., *CircBCAR3* accelerates esophageal cancer tumorigenesis and metastasis via sponging *miR-27a-3p*, *Mol. Cancer* 21 (1) (2022) 1–20.
- [44] A.A. Ishola, C.-S. Chien, Y.-P. Yang, Y. Chien, A.A. Yarmishyn, P.-H. Tsai, et al., Oncogenic circRNA *C190* promotes non-small cell lung cancer via modulation of the *EGFR/ERK* Pathway/*C190* participates in *EGFR* pathway activation in lung cancer, *Cancer Res.* 82 (1) (2022) 75–89.
- [45] R. Guo, X. Cui, X. Li, W. Zang, M. Chang, Z. Sun, et al., *CircMANIA2* is upregulated by *Helicobacter pylori* and promotes development of gastric cancer, *Cell Death Dis.* 13 (4) (2022) 1–11.
- [46] R. Yang, H. Chen, L. Xing, B. Wang, M. Hu, X. Ou, et al., Hypoxia-induced circWSB1 promotes breast cancer progression through destabilizing p53 by interacting with *USP10*, *Mol. Cancer* 21 (1) (2022) 1–20.
- [47] Z. Fu, P. Zhang, R. Zhang, B. Zhang, S. Xiang, Y. Zhang, et al., Novel hypoxia-induced *HIF1 α -circTDRD3*-positive feedback loop promotes the growth and metastasis of colorectal cancer, *Oncogene* (2022) 1–15.
- [48] S. Ghafouri-Fard, M. Taheri, B.M. Hussien, J. Vafaimehanesh, A. Abak, R. Vafaee, Function of circular RNAs in the pathogenesis of colorectal cancer, *Biomed. Pharmacother.* 140 (2021) 111721.
- [49] X. Zhu, X. Wang, S. Wei, Y. Chen, Y. Chen, X. Fan, et al., *hsa_circ_0013958*: a Circular RNA and Potential Novel Biomarker for Lung Adenocarcinoma, Wiley Online Library, 2017, pp. 2170–2182.
- [50] J. Wei, W. Wei, H. Xu, Z. Wang, W. Gao, T. Wang, et al., Circular RNA *hsa_circRNA_102958* may serve as a diagnostic marker for gastric cancer, *Cancer Biomarkers* 27 (2) (2020) 139–145.
- [51] J. Huang, Q. Zhou, Y. Li, Circular RNAs in gynecological diseases: promising biomarkers and diagnostic targets, *Biosci. Rep.* 39 (5) (2019).
- [52] F. Li, L. Zhang, W. Li, J. Deng, J. Zheng, M. An, et al., Circular RNA *ITCH* has inhibitory effect on ESCC by suppressing the *Wnt/ β -catenin* pathway, *Oncotarget* 6 (8) (2015) 6001.
- [53] W-h Li, Y-c Song, H. Zhang, Z-j Zhou, X. Xie, Q-n Zeng, et al., Decreased expression of *Hsa_circ_00001649* in gastric cancer and its clinical significance, *Dis. Markers* 2017 (2017).
- [54] R. He, P. Liu, X. Xie, Y. Zhou, Q. Liao, W. Xiong, et al., *circGFRA1* and *GFRA1* act as ceRNAs in triple negative breast cancer by regulating *miR-34a*, *J. Exp. Clin. Cancer Res.* 36 (1) (2017) 1–12.
- [55] Z. Li, W. Yanfang, J. Li, P. Jiang, T. Peng, K. Chen, et al., Tumor-released exosomal circular RNA *PDE8A* promotes invasive growth via the *miR-338/MAC1/MET* pathway in pancreatic cancer, *Cancer Lett.* 432 (2018) 237–250.
- [56] J. Liu, X. Zhang, M. Yan, H. Li, Emerging role of circular RNAs in cancer, *Front. Oncol.* 10 (2020) 663.
- [57] H. Xu, C. Wang, H. Song, Y. Xu, G. Ji, RNA-Seq profiling of circular RNAs in human colorectal Cancer liver metastasis and the potential biomarkers, *Mol. Cancer* 18 (1) (2019) 1–6.
- [58] Z.Q. Hu, S.L. Zhou, J. Li, Z.J. Zhou, P.C. Wang, H.Y. Xin, et al., Circular RNA sequencing identifies *CircASAP1* as a key regulator in hepatocellular carcinoma metastasis, *Hepatology* 72 (3) (2020) 906–922.
- [59] R. Persiani, S. Rausei, C. Pozzo, A. Biondi, C. Barone, F. Cananzi, et al., 7-Year survival results of perioperative chemotherapy with epirubicin, etoposide, and cisplatin (EEP) in locally advanced resectable gastric cancer: up-to-date analysis of a phase-II study, *Ann. Surg. Oncol.* 15 (8) (2008) 2146–2152.
- [60] X. Huang, Z. Li, Q. Zhang, W. Wang, B. Li, L. Wang, et al., Circular RNA *AKT3* upregulates *PIK3R1* to enhance cisplatin resistance in gastric cancer via *miR-198* suppression, *Mol. Cancer* 18 (1) (2019) 1–20.
- [61] X. Wang, H. Wang, H. Jiang, L. Qiao, C. Guo, Circular RNA *circ_0076305* promotes cisplatin (DDP) resistance of non-small cell lung cancer cells by regulating *ABCC1* through *miR-186-5p*, *Cancer Biother. Radiopharm.* 38 (5) (2023 Jun 1) 293–304. X.
- [62] J. Li, J. Wang, Z. Chen, Y. Chen, M. Jin, *Hsa_circ_0079530* promotes cell proliferation and invasion in non-small cell lung cancer, *Gene* 665 (2018) 1–5.
- [63] B. Yan, W. Zhang, X. Mao, L. Jiang, Circular RNA *circS-7* correlates with advance disease and poor prognosis, and its down-regulation inhibits cells proliferation while induces cells apoptosis in non-small cell lung cancer, *Eur. Rev. Med. Pharmacol. Sci.* 22 (24) (2018) 8712–8721.
- [64] D. Hang, J. Zhou, N. Qin, W. Zhou, H. Ma, G. Jin, et al., A novel plasma circular RNA *circ FARSA* is a potential biomarker for non-small cell lung cancer, *Cancer Med.* 7 (6) (2018) 2783–2791.
- [65] Y. Wang, Y. Li, H. He, F. Wang, Circular RNA *circ-PRMT5* facilitates non-small cell lung cancer proliferation through upregulating *EZH2* via sponging *miR-377/382/498*, *Gene* 720 (2019) 144099.
- [66] L. Chen, A. Nan, N. Zhang, Y. Jia, X. Li, Y. Ling, et al., Circular RNA 100146 functions as an oncogene through direct binding to *miR-361-3p* and *miR-615-5p* in non-small cell lung cancer, *Mol. Cancer* 18 (1) (2019) 1–8.
- [67] L. Li, M. Du, C. Wang, P. He, Reduced expression of circRNA *novel_circ_0005280* and its clinical value in the diagnosis of non-small cell lung cancer, *J. Thorac. Dis.* 12 (12) (2020) 7281.
- [68] L. Zong, Q. Sun, H. Zhang, Z. Chen, Y. Deng, D. Li, et al., Increased expression of circRNA_102231 in lung cancer and its clinical significance, *Biomed. Pharmacother.* 102 (2018) 639–644.
- [69] L. Wan, L. Zhang, K. Fan, Z.-X. Cheng, Q.-C. Sun, J.-J. Wang, Circular RNA-*ITCH* suppresses lung cancer proliferation via inhibiting the *Wnt/ β -catenin* pathway, *BioMed Res. Int.* 2016 (2016).
- [70] X. Li, Z. Zhang, H. Jiang, Q. Li, R. Wang, H. Pan, et al., Circular RNA *circPVT1* promotes proliferation and invasion through sponging *miR-125b* and activating *E2F2* signaling in non-small cell lung cancer, *Cell. Physiol. Biochem.* 51 (5) (2018) 2324–2340.
- [71] X. Pei, S.-W. Chen, X. Long, S.-Q. Zhu, B.-Q. Qiu, K. Lin, et al., *circMET* promotes NSCLC cell proliferation, metastasis, and immune evasion by regulating the *miR-145-5p/CXCL3* axis, *Aging (Albany NY)* 12 (13) (2020) 13038.
- [72] J. Yao, G. Xu, L. Zhu, H. Zheng, *circGFRA1* enhances NSCLC progression by sponging *miR-188-3p*, *OncoTargets Ther.* 13 (2020) 549.
- [73] M.-S. Huang, J.-Y. Liu, X.-B. Xia, Y.-Z. Liu, X. Li, J.-Y. Yin, et al., *Hsa_circ_0001946* inhibits lung cancer progression and mediates cisplatin sensitivity in non-small cell lung cancer via the nucleotide excision repair signaling pathway, *Front. Oncol.* 9 (2019) 508.
- [74] Y. Yao, Q. Hua, Y. Zhou, H. Shen, CircRNA *has_circ_0001946* promotes cell growth in lung adenocarcinoma by regulating *miR-135a-5p/SIRT1* axis and activating *Wnt/ β -catenin* signaling pathway, *Biomed. Pharmacother.* 111 (2019) 1367–1375.
- [75] X. Li, Y. Feng, B. Yang, T. Xiao, H. Ren, X. Yu, et al., A novel circular RNA, *hsa_circ_0030998* suppresses lung cancer tumorigenesis and Taxol resistance by sponging *miR-558*, *Mol. Oncol.* 15 (8) (2021) 2235–2248.
- [76] Y. Yuan, X. Zhou, Y. Kang, H. Kuang, Q. Peng, B. Zhang, et al., *Circ-CCS* is identified as a cancer-promoting circRNA in lung cancer partly by regulating the *miR-383/E2F7* axis, *Life Sci.* 267 (2021) 118955.
- [77] Z. Xu, W. Xiang, W. Chen, Y. Sun, F. Qin, J. Wei, et al., *Circ-IGF1R* inhibits cell invasion and migration in non-small cell lung cancer, *Thoracic cancer* 11 (4) (2020) 875–887.
- [78] Z-f Zhou, Z. Wei, J.-C. Yao, S-y Liu, F. Wang, Z. Wang, et al., CircRNA_102179 promotes the proliferation, migration and invasion in non-small cell lung cancer cells by regulating *miR-330-5p/HMGB3* axis, *Pathol. Res. Pract.* 216 (11) (2020) 153144.
- [79] Y. Wang, R. Xu, D. Zhang, T. Lu, W. Yu, Y. Wo, et al., *Circ-ZKSCAN1* regulates *FAM83A* expression and inactivates *MAPK* signaling by targeting *miR-330-5p* to promote non-small cell lung cancer progression, *Transl. Lung Cancer Res.* 8 (6) (2019) 862.
- [80] S. Gao, Y. Yu, L. Liu, J. Meng, G. Li, Circular RNA *hsa_circ_0007059* restrains proliferation and epithelial-mesenchymal transition in lung cancer cells via inhibiting microRNA-378, *Life Sci.* 233 (2019) 116692.
- [81] Q. Yue, Y. Xu, X. Deng, S. Wang, J. Qiu, B. Qian, et al., *Circ-PITX1* promotes the progression of non-small cell lung cancer through regulating the *miR-1248/CCND2* axis, *OncoTargets Ther.* 14 (2021) 1807.
- [82] J.-Y. Wang, F. Zhang, L. Hong, S.-J. Wei, CircRNA_0000429 regulates development of NSCLC by acting as a sponge of *miR-1197* to control *MADD*, *Cancer Manag. Res.* 13 (2021) 861.
- [83] C.-C. Zhang, Y. Li, X.-Z. Feng, D.-B. Li, Circular RNA *circ_0001287* inhibits the proliferation, metastasis, and radiosensitivity of non-small cell lung cancer cells by sponging microRNA *miR-21* and up-regulating phosphatase and tensin homolog expression, *Bioengineered* 12 (1) (2021) 414–425.
- [84] Y.-H. Luo, X.-Z. Zhu, K.-W. Huang, Q. Zhang, Y.-X. Fan, P.-W. Yan, et al., Emerging roles of circular RNA *hsa_circ_0000064* in the proliferation and metastasis of lung cancer, *Biomed. Pharmacother.* 96 (2017) 892–898.
- [85] B.Q. Qiu, P.F. Zhang, D. Xiong, J.J. Xu, X. Long, S.Q. Zhu, et al., CircRNA fibroblast growth factor receptor 3 promotes tumor progression in non-small cell lung cancer by regulating *Galactin1-1AKT/ERK1/2* signaling, *J. Cell. Physiol.* 234 (7) (2019) 11256–11264.

- [86] J. An, H. Shi, N. Zhang, S. Song, Elevation of circular RNA circ_0003645 forecasts unfavorable prognosis and facilitates cell progression via miR-1179/TMEM14A pathway in non-small cell lung cancer, *Biochem. Biophys. Res. Commun.* 511 (4) (2019) 921–925.
- [87] X. Zhang, D. Yang, Y. Wei, Overexpressed CDR1as functions as an oncogene to promote the tumor progression via miR-7 in non-small-cell lung cancer, *Oncotargets Ther.* 11 (2018) 3979.
- [88] Z. Fan, Y. Bai, Q. Zhang, P. Qian, CircRNA circ_POLA2 promotes lung cancer cell stemness via regulating the miR-326/GNB1 axis, *Environ. Toxicol.* 35 (10) (2020) 1146–1156.
- [89] G. Liu, H. Shi, L. Deng, H. Zheng, W. Kong, X. Wen, et al., Circular RNA circ-FOXM1 facilitates cell progression as ceRNA to target PDPDF and MACC1 by sponging miR-1304-5p in non-small cell lung cancer, *Biochem. Biophys. Res. Commun.* 513 (1) (2019) 207–212.
- [90] Y. Chi, Q. Luo, Y. Song, F. Yang, Y. Wang, M. Jin, et al., Circular RNA circPIP5K1A promotes non-small cell lung cancer proliferation and metastasis through miR-600/HIF-1 α regulation, *J. Cell. Biochem.* 120 (11) (2019) 19019–19030.
- [91] N. Shao, L. Song, X. Sun, Exosomal circ_PIP5K1A regulates the progression of non-small cell lung cancer and cisplatin sensitivity by miR-101/ABCC1 axis, *Mol. Cell. Biochem.* 476 (6) (2021) 2253–2267.
- [92] P. Zhang, X. Xue, X. Ling, Q. Yang, Y. Yu, J. Xiao, et al., CircRNA_010763 promotes growth and invasion of lung cancer through serving as a molecular sponge of miR-715 to induce c-Myc expression, *Eur. Rev. Med. Pharmacol. Sci.* 24 (13) (2020) 7310–7319.
- [93] J.-T. Yao, S.-H. Zhao, Q.-P. Liu, M.-Q. Lv, D.-X. Zhou, Z.-J. Liao, et al., Overexpression of CircRNA_100876 in non-small cell lung cancer and its prognostic value, *Pathol. Res. Pract.* 213 (5) (2017) 453–456.
- [94] Y. Wang, circ-ANXA7 facilitates lung adenocarcinoma progression via miR-331/LAD1 axis, *Cancer Cell Int.* 21 (1) (2021) 1–15.
- [95] Y. Wang, Z. Wang, J. Lu, H. Zhang, Circular RNA circ-PTEN elevates PTEN inhibiting the proliferation of non-small cell lung cancer cells, *Hum. Cell* 34 (4) (2021) 1174–1184.
- [96] R. Gu, K. Shao, Q. Xu, X. Zhao, H. Qiu, H. Hu, Circular RNA hsa_circ_0008003 facilitates tumorigenesis and development of non-small cell lung carcinoma via modulating miR-488/ZNF281 axis, *J. Cell Mol. Med.* 26 (6) (2022) 1754–1765.
- [97] Q. Geng, Q. Wu, Y. Zhang, G. Zhang, J. Fu, N. Chen, Clinical significance of circ-MTHFD2 in diagnosis, pathological staging and prognosis of NSCLC, *Eur. Rev. Med. Pharmacol. Sci.* 24 (18) (2020) 9473–9479.
- [98] M. Huang, Y.-R. He, L.-C. Liang, Q. Huang, Z.-Q. Zhu, Circular RNA hsa_circ_0000745 may serve as a diagnostic marker for gastric cancer, *World J. Gastroenterol.* 23 (34) (2017) 6330.
- [99] H. Sun, W. Tang, D. Rong, H. Jin, K. Fu, W. Zhang, et al., Hsa_circ_0000520, a potential new circular RNA biomarker, is involved in gastric carcinoma, *Cancer Biomarkers* 21 (2) (2018) 299–306.
- [100] Y. Shao, J. Li, R. Lu, T. Li, Y. Yang, B. Xiao, et al., Global circular RNA expression profile of human gastric cancer and its clinical significance, *Cancer Med.* 6 (6) (2017) 1173–1180.
- [101] P. Li, H. Chen, S. Chen, X. Mo, T. Li, B. Xiao, et al., Circular RNA 0000096 affects cell growth and migration in gastric cancer, *Br. J. Cancer* 116 (5) (2017) 626–633.
- [102] D. Rong, C. Dong, K. Fu, H. Wang, W. Tang, H. Cao, Upregulation of circ_0066444 promotes the proliferation, invasion, and migration of gastric cancer cells, *Oncotargets Ther.* 11 (2018) 2753.
- [103] J. Zhang, H. Liu, L. Hou, G. Wang, R. Zhang, Y. Huang, et al., Circular RNA_LARP4 inhibits cell proliferation and invasion of gastric cancer by sponging miR-424-5p and regulating LATS1 expression, *Mol. Cancer* 16 (1) (2017) 1–16.
- [104] J. Lu, P.-y Zhang, P. Li, J.-w Xie, J.-b Wang, J.-x Lin, et al., Circular RNA hsa_circ_0001368 suppresses the progression of gastric cancer by regulating miR-6506-5p/FOXO3 axis, *Biochem. Biophys. Res. Commun.* 512 (1) (2019) 29–33.
- [105] J. Chen, Y. Li, Q. Zheng, C. Bao, J. He, B. Chen, et al., Circular RNA profile identifies circPVT1 as a proliferative factor and prognostic marker in gastric cancer, *Cancer Lett.* 388 (2017) 208–219.
- [106] X. Zhang, S. Wang, H. Wang, J. Cao, X. Huang, Z. Chen, et al., Circular RNA circNRP1 acts as a microRNA-149-5p sponge to promote gastric cancer progression via the AKT1/mTOR pathway, *Mol. Cancer* 18 (1) (2019) 1–24.
- [107] W. Tang, K. Fu, H. Sun, D. Rong, H. Wang, H. Cao, CircRNA microarray profiling identifies a novel circulating biomarker for detection of gastric cancer, *Mol. Cancer* 17 (1) (2018) 1–6.
- [108] Y. Peng, K. Pu, H. Su, J. Zhang, Y. Zheng, R. Ji, et al., Circular RNA hsa_circ_0010882 promotes the progression of gastric cancer via regulation of the PI3K/Akt/mTOR signaling pathway, *Eur. Rev. Med. Pharmacol. Sci.* 24 (3) (2020) 1142–1151.
- [109] L. Zhang, X. Chang, T. Zhai, J. Yu, W. Wang, A. Du, et al., A novel circular RNA, circ-ATAD1, contributes to gastric cancer cell progression by targeting miR-140-3p/YY1/PCIF1 signaling axis, *Biochem. Biophys. Res. Commun.* 525 (4) (2020) 841–849.
- [110] J. Cheng, H. Zhuo, M. Xu, L. Wang, H. Xu, J. Peng, et al., Regulatory network of circRNA-miRNA-mRNA contributes to the histological classification and disease progression in gastric cancer, *J. Transl. Med.* 16 (1) (2018) 1–14.
- [111] L. Wu, D. Liu, Y. Yang, Enhanced expression of circular RNA circ-DCAF6 predicts adverse prognosis and promotes cell progression via sponging miR-1231 and miR-1256 in gastric cancer, *Exp. Mol. Pathol.* 110 (2019) 104273.
- [112] S. Wang, X. Zhang, Z. Li, W. Wang, B. Li, X. Huang, et al., Circular RNA profile identifies circOSBPL10 as an oncogenic factor and prognostic marker in gastric cancer, *Oncogene* 38 (44) (2019) 6985–7001.
- [113] Z. Zhu, Z. Rong, Z. Luo, Z. Yu, J. Zhang, Z. Qiu, et al., Circular RNA circNHSL1 promotes gastric cancer progression through the miR-1306-3p/SIX1/vimentin axis, *Mol. Cancer* 18 (1) (2019) 1–20.
- [114] J. He, J. Chen, B. Ma, L. Jiang, G. Zhao, CircLMTK2 acts as a novel tumor suppressor in gastric cancer, *Biosci. Rep.* 39 (5) (2019).
- [115] H.-D. Sun, Z.-P. Xu, Z.-Q. Sun, B. Zhu, Q. Wang, J. Zhou, et al., Down-regulation of circPVR13 promotes the proliferation and migration of gastric cancer cells, *Sci. Rep.* 8 (1) (2018) 1–13.
- [116] Z. Luo, Z. Rong, J. Zhang, Z. Zhu, Z. Yu, T. Li, et al., Circular RNA circCCDC9 acts as a miR-6792-3p sponge to suppress the progression of gastric cancer through regulating CAV1 expression, *Mol. Cancer* 19 (1) (2020) 1–21.
- [117] X.-N. Li, Z.-J. Wang, C.-X. Ye, B.-C. Zhao, X.-X. Huang, L. Yang, Circular RNA circVAPA is up-regulated and exerts oncogenic properties by sponging miR-101 in colorectal cancer, *Biomed. Pharmacother.* 112 (2019) 108611.
- [118] X. Li, J. Wang, C. Zhang, C. Lin, J. Zhang, W. Zhang, et al., Circular RNA circITGA7 inhibits colorectal cancer growth and metastasis by modulating the Ras pathway and upregulating transcription of its host gene ITGA7, *J. Pathol.* 246 (2) (2018) 166–179.
- [119] K. Zeng, X. Chen, M. Xu, X. Liu, X. Hu, T. Xu, et al., CircHIPK3 promotes colorectal cancer growth and metastasis by sponging miR-7, *Cell Death Dis.* 9 (4) (2018) 1–15.
- [120] W. Weng, Q. Wei, S. Toden, K. Yoshida, T. Nagasaka, T. Fujiwara, et al., Circular RNA ciRS-7—a promising prognostic biomarker and a potential therapeutic target in colorectal Cancer ciRS-7 in colorectal cancer, *Clin. Cancer Res.* 23 (14) (2017) 3918–3928.
- [121] G. Fang, B.-L. Ye, B.-R. Hu, X.-J. Ruan, Y.-X. Shi, CircRNA_100290 promotes colorectal cancer progression through miR-516b-induced downregulation of FZD4 expression and Wnt/ β -catenin signaling, *Biochem. Biophys. Res. Commun.* 504 (1) (2018) 184–189.
- [122] K.-Y. Hsiao, Y.-C. Lin, S.K. Gupta, N. Chang, L. Yen, H.S. Sun, et al., Noncoding effects of circular RNA CCDC66 promote colon cancer growth and Metastasis CircCCDC66 promotes colon cancer progression, *Cancer Res.* 77 (9) (2017) 2339–2350.
- [123] C. Jin, A. Wang, L. Liu, G. Wang, G. Li, Hsa_circ_0136666 promotes the proliferation and invasion of colorectal cancer through miR-136/SH2B1 axis, *J. Cell. Physiol.* 234 (5) (2019) 7247–7256.
- [124] W. Ji, C. Qiu, M. Wang, N. Mao, S. Wu, Y. Dai, Hsa_circ_0001649: a circular RNA and potential novel biomarker for colorectal cancer, *Biochem. Biophys. Res. Commun.* 497 (1) (2018) 122–126.
- [125] J. Li, S. Ni, C. Zhou, M. Ye, The expression profile and clinical application potential of hsa_circ_0000711 in colorectal cancer, *Cancer Manag. Res.* 10 (2018) 2777.
- [126] F. Wang, J. Wang, X. Cao, L. Xu, L. Chen, Hsa_circ_0014717 is downregulated in colorectal cancer and inhibits tumor growth by promoting p16 expression, *Biomed. Pharmacother.* 98 (2018) 775–782.
- [127] J. Wang, X. Li, L. Lu, L. He, H. Hu, Z. Xu, Circular RNA hsa_circ_0000567 can be used as a promising diagnostic biomarker for human colorectal cancer, *J. Clin. Lab. Anal.* 32 (5) (2018) e22379.
- [128] X. Wang, Y. Zhang, L. Huang, J. Zhang, F. Pan, B. Li, et al., Decreased expression of hsa_circ_001988 in colorectal cancer and its clinical significances, *Int. J. Clin. Exp. Pathol.* 8 (12) (2015) 16020.
- [129] Y. Yuan, W. Liu, Y. Zhang, Y. Zhang, S. Sun, CircRNA circ_0026344 as a prognostic biomarker suppresses colorectal cancer progression via microRNA-21 and microRNA-31, *Biochem. Biophys. Res. Commun.* 503 (2) (2018) 870–875.
- [130] F. Zhuo, H. Lin, Z. Chen, Z. Huang, J. Hu, The expression profile and clinical significance of circRNA0003906 in colorectal cancer, *Oncotargets Ther.* 10 (2017) 5187.
- [131] P. Zhang, Z. Zuo, W. Shang, A. Wu, R. Bi, J. Wu, et al., Identification of differentially expressed circular RNAs in human colorectal cancer, *Tumor Biol.* 39 (3) (2017) 1010428317694546.
- [132] T. Liu, P. Ye, Y. Ye, S. Lu, B. Han, Circular RNA hsa_circRNA_002178 silencing retards breast cancer progression via microRNA-328-3p-mediated inhibition of COL1A1, *J. Cell Mol. Med.* 24 (3) (2020) 2189–2201.
- [133] Y. Xu, Y. Yao, K. Leng, D. Ji, L. Qu, Y. Liu, et al., Increased expression of circular RNA circ_0005230 indicates dismal prognosis in breast cancer and regulates cell proliferation and invasion via miR-618/CBX8 signal pathway, *Cell. Physiol. Biochem.* 51 (4) (2018) 1710–1722.
- [134] H. Tang, X. Huang, J. Wang, L. Yang, Y. Kong, G. Gao, et al., circKIF4A acts as a prognostic factor and mediator to regulate the progression of triple-negative breast cancer, *Mol. Cancer* 18 (1) (2019) 1–9.
- [135] Y. Wei, X. Chen, C. Liang, Y. Ling, X. Yang, X. Ye, et al., A noncoding regulatory RNAs network driven by Circ-CDYL acts specifically in the early stages hepatocellular carcinoma, *Hepatology* 71 (1) (2020) 130–147.
- [136] Z. Li, Y. Zhou, G. Yang, S. He, X. Qiu, L. Zhang, et al., Using circular RNA SMARCA5 as a potential novel biomarker for hepatocellular carcinoma, *Clin. Chim. Acta* 492 (2019) 37–44.
- [137] J. Yu, Wb Ding, Mc Wang, Xg Guo, J. Xu, Qg Xu, et al., Plasma circular RNA panel to diagnose hepatitis B virus-related hepatocellular carcinoma: a large-scale, multicenter study, *Int. J. Cancer* 146 (6) (2020) 1754–1763.
- [138] X. Zhang, Y. Xu, Z. Qian, W. Zheng, Q. Wu, Y. Chen, et al., circRNA_104075 stimulates YAP-dependent tumorigenesis through the regulation of HNF4a and may serve as a diagnostic marker in hepatocellular carcinoma, *Cell Death Dis.* 9 (11) (2018) 1–14.
- [139] Z. Yao, J. Luo, K. Hu, J. Lin, H. Huang, Q. Wang, et al., Zkscan 1 gene and its related circular rna (circ zkscan 1) both inhibit hepatocellular carcinoma cell

- growth, migration, and invasion but through different signaling pathways, *Mol. Oncol.* 11 (4) (2017) 422–437.
- [140] J. Meng, S. Chen, J.-X. Han, B. Qian, X.-R. Wang, W.-L. Zhong, et al., Twist1 regulates vimentin through Cui2 circular RNA to promote EMT in hepatocellular carcinoma, *Twist1 regulates vimentin through circRNA-Cui2 in HCC*, *Cancer Res.* 78 (15) (2018) 4150–4162.
- [141] S. Li, H. Gu, Y. Huang, Q. Peng, R. Zhou, P. Yi, et al., Circular RNA 101368/miR-200a axis modulates the migration of hepatocellular carcinoma through HMGB1/RAGE signaling, *Cell Cycle* 17 (19–20) (2018) 2349–2359.
- [142] X.-Y. Huang, Z.-L. Huang, Y.-H. Xu, Q. Zheng, Z. Chen, W. Song, et al., Comprehensive circular RNA profiling reveals the regulatory role of the circRNA-100338/miR-141-3p pathway in hepatitis B-related hepatocellular carcinoma, *Sci. Rep.* 7 (1) (2017) 1–12.
- [143] W. Dong, Z.-h Dai, F.-c Liu, X.-g Guo, C.-m Ge, J. Ding, et al., The RNA-binding protein RBM3 promotes cell proliferation in hepatocellular carcinoma by regulating circular RNA SCD-circRNA 2 production, *EBioMedicine* 45 (2019) 155–167.
- [144] L. Qiu, Y. Huang, Z. Li, X. Dong, G. Chen, H. Xu, et al., Circular RNA profiling identifies circ ADAMTS 13 as a miR-484 sponge which suppresses cell proliferation in hepatocellular carcinoma, *Mol. Oncol.* 13 (2) (2019) 441–455.
- [145] F. Yang, D.-Y. Liu, J.-T. Guo, N. Ge, P. Zhu, X. Liu, et al., Circular RNA circ-LDLRAD3 as a biomarker in diagnosis of pancreatic cancer, *World J. Gastroenterol.* 23 (47) (2017) 8345.
- [146] J. Li, Z. Li, P. Jiang, M. Peng, X. Zhang, K. Chen, et al., Circular RNA IARS (circ-IARS) secreted by pancreatic cancer cells and located within exosomes regulates endothelial monolayer permeability to promote tumor metastasis, *J. Exp. Clin. Cancer Res.* 37 (1) (2018) 1–16.
- [147] L. Lu, F. Wang, J. Chen, C. Zhao, S. Guo, D. Dong, et al., CircTENM3 inhibits tumor progression via the miR-558/RUNX3 axis in prostate cancer, *J. Transl. Med.* 21 (1) (2023) 850.
- [148] R. Wang, J. Zhong, X. Pan, Z. Su, Y. Xu, M. Zhang, et al., A novel intronic circular RNA circFGFR1(int2) up-regulates FGFR1 by recruiting transcriptional activators P65/FUS and suppressing miR-4687-5p to promote prostate cancer progression, *J. Transl. Med.* 21 (1) (2023) 840.
- [149] W. Deng, X. Zhou, K. Zhu, R. Chen, X. Liu, L. Chen, et al., Novel circular RNA circ_0086722 drives tumor progression by regulating the miR-339-5p/STAT5A axis in prostate cancer, *Cancer Lett.* 533 (2022) 215606.
- [150] Z. Kong, Y. Lu, Y. Yang, K. Chang, Y. Lin, Y. Huang, et al., m6A-Mediated biogenesis of circDDIT4 inhibits prostate cancer progression by sequestering ELAVL1/HuR, *Mol. Cancer Res.* 21 (12) (2023) 1342–1355.
- [151] Z.-H. Zong, Y.-P. Du, X. Guan, S. Chen, Y. Zhao, CircWHSC1 promotes ovarian cancer progression by regulating MUC1 and hTERT through sponging miR-145 and miR-1182, *J. Exp. Clin. Cancer Res.* 38 (1) (2019) 1–10.
- [152] L. Zhang, Q. Zhou, Q. Qiu, L. Hou, M. Wu, J. Li, et al., CircPLEKHM3 acts as a tumor suppressor through regulation of the miR-9/BRCA1/DNAJB6/KLF4/AKT1 axis in ovarian cancer, *Mol. Cancer* 18 (1) (2019) 1–19.
- [153] H. Su, T. Tao, Z. Yang, X. Kang, X. Zhang, D. Kang, et al., Circular RNA cTFR3 acts as the sponge of MicroRNA-107 to promote bladder carcinoma progression, *Mol. Cancer* 18 (1) (2019) 1–15.
- [154] Y. Li, B. Wan, L. Liu, L. Zhou, Q. Zeng, Circular RNA circMTO1 suppresses bladder cancer metastasis by sponging miR-221 and inhibiting epithelial-to-mesenchymal transition, *Biochem. Biophys. Res. Commun.* 508 (4) (2019) 991–996.
- [155] Z. Kun-Peng, M. Xiao-Long, Z. Chun-Lin, Overexpressed circPVT1, a potential new circular RNA biomarker, contributes to doxorubicin and cisplatin resistance of osteosarcoma cells by regulating ABCB1, *Int. J. Biol. Sci.* 14 (3) (2018) 321.
- [156] M. Yang, G. Li, L. Fan, G. Zhang, J. Xu, J. Zhang, Circular RNA circ_0034642 elevates BATEF3 expression and promotes cell proliferation and invasion through miR-1205 in glioma, *Biochem. Biophys. Res. Commun.* 508 (3) (2019) 980–985.
- [157] Q. Zhong, J. Huang, J. Wei, R. Wu, Circular RNA CDR1as sponges miR-7-5p to enhance E2F3 stability and promote the growth of nasopharyngeal carcinoma, *Cancer Cell Int.* 19 (1) (2019) 1–13.
- [158] D.W. Thomson, M.E. Dinger, Endogenous microRNA sponges: evidence and controversy, *Nat. Rev. Genet.* 17 (5) (2016) 272–283.
- [159] J. Bi, H. Liu, W. Dong, W. Xie, Q. He, Z. Cai, et al., Circular RNA circ-ZKSCAN1 inhibits bladder cancer progression through miR-1178-3p/p21 axis and acts as a prognostic factor of recurrence, *Mol. Cancer* 18 (1) (2019) 1–14.
- [160] T.B. Hansen, E.D. Wiklund, J.B. Bramsen, S.B. Villadsen, A.L. Statham, S.J. Clark, et al., miRNA-dependent gene silencing involving Ago2-mediated cleavage of a circular antisense RNA, *EMBO J.* 30 (2011) 4414–4422.
- [161] X.-O. Zhang, H.-B. Wang, Y. Zhang, X. Lu, L.-L. Chen, L. Yang, Complementary sequence-mediated exon circularization, *Cell* 159 (1) (2014) 134–147.
- [162] Y. Chen, F. Yang, E. Fang, W. Xiao, H. Mei, H. Li, et al., Circular RNA circAGO2 drives cancer progression through facilitating HuR-repressed functions of AGO2-miRNA complexes, *Cell Death Differ.* 26 (7) (2019) 1346–1364.
- [163] L. Fang, W.W. Du, J. Lyu, J. Dong, C. Zhang, W. Yang, et al., Enhanced breast cancer progression by mutant p53 is inhibited by the circular RNA circ-Ccnb1, *Cell Death Differ.* 25 (12) (2018) 2195–2208.
- [164] X. Zhang, J. Zhang, Q. Liu, Y. Zhao, W. Zhang, H. Yang, Circ-CUX1 accelerates the progression of neuroblastoma via miR-16-5p/DMRT2 axis, *Neurochem. Res.* 45 (12) (2020) 2840–2855.
- [165] K. Han, F.-W. Wang, C.-H. Cao, H. Ling, J.-W. Chen, R.-X. Chen, et al., CircLONP2 enhances colorectal carcinoma invasion and metastasis through modulating the maturation and exosomal dissemination of microRNA-17, *Mol. Cancer* 19 (1) (2020) 1–18.
- [166] W. Li, C. Zhong, J. Jiao, P. Li, B. Cui, C. Ji, et al., Characterization of hsa_circ_0004277 as a new biomarker for acute myeloid leukemia via circular RNA profile and bioinformatics analysis, *Int. J. Mol. Sci.* 18 (3) (2017) 597.
- [167] P. Fang, Q. Jiang, S. Liu, J. Gu, K. Hu, Z. Wang, Circ_0002099 is a novel molecular therapeutic target for bladder cancer, *Drug Dev. Res.* 83 (8) (2022) 1890–1905.
- [168] W. Weng, Q. Wei, S. Toden, K. Yoshida, T. Nagasaka, T. Fujiwara, et al., Circular RNA circRS7—a promising prognostic biomarker and a potential therapeutic target in colorectal cancer, *Clin. Cancer Res.* 23 (14) (2017) 3918–3928.
- [169] H. Yang, X. Li, Q. Meng, H. Sun, S. Wu, W. Hu, et al., CircPTK2 (hsa_circ_0005273) as a novel therapeutic target for metastatic colorectal cancer, *Mol. Cancer* 19 (2020) 1–15.
- [170] Y. Zhang, J. Luo, W. Yang, W.-C. Ye, CircRNAs in colorectal cancer: potential biomarkers and therapeutic targets, *Cell Death Dis.* 14 (6) (2023) 353.
- [171] S.I.S. Al-Hawary, W. Asghar, A. Amin, Y.F. Mustafa, A. Hjazji, A.F. Almulla, et al., Circ_0067934 as a novel therapeutic target in cancer: from mechanistic to clinical perspectives, *Pathol. Res. Pract.* (2023) 154469.
- [172] Y. Zhu, L. Liang, Y. Zhao, J. Li, J. Zeng, Y. Yuan, et al., CircNUP50 is a novel therapeutic target that promotes cisplatin resistance in ovarian cancer by modulating p53 ubiquitination, *J. Nanobiotechnol.* 22 (1) (2024) 35.
- [173] G. Zheng, J. Huang, W. Chen, P. You, Y. Ding, P. Tu, circUBAP2 exacerbates malignant capabilities of NSCLC by targeting KLF4 through miR-3182 modulation, *Aging (Albany NY)* 13 (8) (2021) 11083.
- [174] D. Chen, W. Ma, Z. Ke, F. Xie, CircRNA hsa_circ_100395 regulates miR-1228/TCF21 pathway to inhibit lung cancer progression, *Cell Cycle* 17 (16) (2018) 2080–2090.
- [175] F. Zhao, Y. Han, Z. Liu, Z. Zhao, Z. Li, K. Jia, circFADS2 regulates lung cancer cells proliferation and invasion via acting as a sponge of miR-498, *Biosci. Rep.* 38 (4) (2018).
- [176] Y. Zhang, H. Zhao, L. Zhang, Identification of the tumor-suppressive function of circular RNA FOXO3 in non-small cell lung cancer through sponging miR-155, *Mol. Med. Rep.* 17 (6) (2018) 7692–7700.
- [177] W.-X. Bian, F. Xue, L.-Y. Wang, X.-F. Xing, Circular RNA CircCDYL regulates proliferation and apoptosis in non-small cell lung cancer cells by sponging miR-185-5p and upregulating TNRC6A, *Cancer Manag. Res.* 13 (2021) 633.
- [178] F. Cheng, J. Yu, X. Zhang, Z. Dai, A. Fang, CircSEC31A promotes the malignant progression of non-small cell lung cancer through regulating SEC31A expression via sponging miR-376a, *Cancer Manag. Res.* 12 (2020) 11527.
- [179] D. Qu, B. Yan, R. Xin, T. Ma, A novel circular RNA hsa_circ_0020123 exerts oncogenic properties through suppression of miR-144 in non-small cell lung cancer, *Am. J. Cancer Res.* 8 (8) (2018) 1387.
- [180] W. Zhang, C. Song, X. Ren, Circ_0003998 regulates the progression and docetaxel sensitivity of DTX-resistant non-small cell lung cancer cells by the miR-136-5p/CORO1C axis, *Technol. Cancer Res. Treat.* 20 (2021) 1533033821990040.
- [181] Y. Wang, W. Zhao, S. Zhang, STAT3-induced upregulation of circCCDC66 facilitates the progression of non-small cell lung cancer by targeting miR-33a-5p/KPNA4 axis, *Biomed. Pharmacother.* 126 (2020) 110019.
- [182] J. Xu, L. Ni, F. Zhao, X. Dai, J. Tao, J. Pan, et al., Overexpression of hsa_circ_0002874 promotes resistance of non-small cell lung cancer to paclitaxel by modulating miR-1273f/MDM2/p53 pathway, *Aging (Albany NY)* 13 (4) (2021) 5986.
- [183] Y. Liu, C. Yang, C. Cao, Q. Li, X. Jin, H. Shi, Hsa_circ_RNA_0011780 represses the proliferation and metastasis of non-small cell lung cancer by decreasing FBXW7 via targeting miR-544a, *OncoTargets Ther.* 13 (2020) 745.
- [184] H. Du, Z. He, F. Feng, D. Chen, L. Zhang, J. Bai, et al., Hsa_circ_0038646 promotes cell proliferation and migration in colorectal cancer via miR-331-3p/GRIK3, *Oncol. Lett.* 20 (1) (2020) 266–274.
- [185] F. Zhang, R. Cheng, P. Li, C. Lu, G. Zhang, Hsa_circ_0010235 functions as an oncogenic drive in non-small cell lung cancer by modulating miR-433-3p/TIPRL axis, *Cancer Cell Int.* 21 (1) (2021) 1–14.
- [186] Q. Liu, G. Cao, Y. Wan, C. Xu, Y. He, G. Li, Hsa_circ_0001073 targets miR-626/LIFR axis to inhibit lung cancer progression, *Environ. Toxicol.* 36 (6) (2021) 1052–1060.
- [187] Y. Lv, C. Wang, L. Li, S. Han, Y. Li, Effects of circRNA_103993 on the proliferation and apoptosis of NSCLC cells through miR-1271/ERG signaling pathway, *Eur. Rev. Med. Pharmacol. Sci.* 24 (16) (2020) 8384–8393.
- [188] T. Wang, X. Wang, Q. Du, N. Wu, X. Liu, Y. Chen, et al., The circRNA circP4HB promotes NSCLC aggressiveness and metastasis by sponging miR-133a-5p, *Biochem. Biophys. Res. Commun.* 513 (4) (2019) 904–911.
- [189] M. Jin, C. Shi, C. Yang, J. Liu, G. Huang, Upregulated circRNA ARHGAP10 predicts an unfavorable prognosis in NSCLC through regulation of the miR-150-5p/GLUT-1 axis, *Mol. Ther. Nucleic Acids* 18 (2019) 219–231.
- [190] L. Wang, H. Ma, W. Kong, B. Liu, X. Zhang, Up-regulated circular RNA VANG1 contributes to progression of non-small cell lung cancer through inhibition of miR-195 and activation of Bcl-2, *Biosci. Rep.* 39 (6) (2019).
- [191] G. Liang, Z. Liu, L. Tan, A. Su, W.G. Jiang, C. Gong, HIF1 α -associated circDENND4C promotes proliferation of breast cancer cells in hypoxic environment, *Anticancer Res.* 37 (8) (2017) 4337–4343.
- [192] S. Ren, J. Liu, Y. Feng, Z. Li, L. He, L. Li, et al., Knockdown of circDENND4C inhibits glycolysis, migration and invasion by up-regulating miR-200b/c in breast cancer under hypoxia, *J. Exp. Clin. Cancer Res.* 38 (1) (2019) 1–12.
- [193] F. Ye, G. Gao, Y. Zou, S. Zheng, L. Zhang, X. Ou, et al., circFBXW7 inhibits malignant progression by sponging miR-197-3p and encoding a 185-aa protein in triple-negative breast cancer, *Mol. Ther. Nucleic Acids* 18 (2019) 88–98.
- [194] J.-Z. Xu, C.-C. Shao, X.-J. Wang, X. Zhao, J.-Q. Chen, Y.-X. Ouyang, et al., circTADA2As suppress breast cancer progression and metastasis via targeting miR-203a-3p/SOCS3 axis, *Cell Death Dis.* 10 (3) (2019) 1–16.

- [195] L. Meng, S. Liu, F. Liu, M. Sang, Y. Ju, X. Fan, et al., ZEB1-mediated transcriptional upregulation of circWWC3 promotes breast cancer progression through activating ras signaling pathway, *Mol. Ther. Nucleic Acids* 22 (2020) 124–137.
- [196] L. Cao, M. Wang, Y. Dong, B. Xu, J. Chen, Y. Ding, et al., Circular RNA circRNF20 promotes breast cancer tumorigenesis and Warburg effect through miR-487a/HIF-1 α /HK2, *Cell Death Dis.* 11 (2) (2020) 1–10.
- [197] G. Liang, Y. Ling, M. Mehrpour, P.E. Saw, Z. Liu, W. Tan, et al., Autophagy-associated circRNA circCDYL augments autophagy and promotes breast cancer progression, *Mol. Cancer* 19 (1) (2020) 1–16.
- [198] J.-H. He, Y.-G. Li, Z.-P. Han, J.-B. Zhou, W.-M. Chen, Y.-B. Lv, et al., The CircRNA-ACAP2/Hsa-miR-21-5p/Tiam1 regulatory feedback circuit affects the proliferation, migration, and invasion of colon cancer SW480 cells, *Cell. Physiol. Biochem.* 49 (4) (2018) 1539–1550.
- [199] Xi Zhang, Li Xu, F. Wang, Hsa_circ_0020397 regulates colorectal cancer cell viability, apoptosis and invasion by promoting the expression of the miR-138 targets TERT and PD-L1, *Cell Biol. Int.* 41 (9) (2017) 1056–1064.
- [200] C. Huang, R. Xu, X. Zhu, H. Jiang, m6A-modified circABCC4 promotes stemness and metastasis of prostate cancer by recruiting IGF2BP2 to increase stability of CCAR1, *Cancer Gene Ther.* 30 (10) (2023) 1426–1440.
- [201] C. Zhong, Z. Long, T. Yang, S. Wang, W. Zhong, F. Hu, et al., M6A-modified circRBM33 promotes prostate cancer progression via PDHA1-mediated mitochondrial respiration regulation and presents a potential target for ARSI therapy, *Int. J. Biol. Sci.* 19 (5) (2023) 1543–1563.
- [202] J. Zhang, H. Liu, P. Zhao, H. Zhou, T. Mao, Has_circ.0055625 from circRNA profile increases colon cancer cell growth by sponging miR-106b-5p, *J. Cell. Biochem.* 120 (3) (2019) 3027–3037.
- [203] Y. Jin, L. Yu, B. Zhang, C. Liu, Y. Chen, Circular RNA hsa_circ_0000523 regulates the proliferation and apoptosis of colorectal cancer cells as miRNA sponge, *Braz. J. Med. Biol. Res.* 51 (2018).
- [204] L. Bian, X. Zhi, L. Ma, J. Zhang, P. Chen, S. Sun, et al., Hsa_circRNA_103809 regulated the cell proliferation and migration in colorectal cancer via miR-532-3p/FOXO4 axis, *Biochem. Biophys. Res. Commun.* 505 (2) (2018) 346–352.
- [205] S. Luo, F. Deng, N. Yao, F. Zheng, Circ.0005875 sponges miR-502-5p to promote renal cell carcinoma progression through upregulating E26 transformation specific-1, *Anti Cancer Drugs* 33 (1) (2022) e286–e298.
- [206] R. Li, S. Luo, D. Zhang, Circular RNA hsa_circ_0054537 sponges miR-130a-3p to promote the progression of renal cell carcinoma through regulating cMet pathway, *Gene* 754 (2020) 144811.
- [207] H. Liu, G. Hu, Z. Wang, Q. Liu, J. Zhang, Y. Chen, et al., circPTCH1 promotes invasion and metastasis in renal cell carcinoma via regulating miR-485-5p/MMP14 axis, *Theranostics* 10 (23) (2020) 10791.
- [208] J. Lee, D. Boczkowski, S. Nair, Programming human dendritic cells with mRNA, *Synthetic Messenger RNA and Cell Metabolism Modulation* (2013) 111–125.
- [209] J.-c Sun, K. Pan, M.-S. Chen, Q.-j Wang, H. Wang, H.-q Ma, et al., Dendritic cell-mediated CTLs targeting hepatocellular carcinoma stem cells, *Cancer Biol. Ther.* 10 (4) (2010) 368–375.
- [210] Z.-y Xiao, H. Tang, Z.-m Xu, Z.-j Yan, P. Li, Y.-q Cai, et al., An experimental study of dendritic cells transfected with cancer stem-like cells RNA against 9L brain tumors, *Cancer Biol. Ther.* 11 (11) (2011) 974–980.
- [211] Y. Chen, I. Noda, Y.M. Jung, Influence of polyethylene glycol on the phase transition of poly (butylene adipate) thin films: polymorphic control, *Spectrochim. Acta Mol. Biomol. Spectrosc.* 184 (2017) 228–234.
- [212] Y.G. Chen, R. Chen, S. Ahmad, R. Verma, S.P. Kasturi, L. Amaya, et al., N6-methyladenosine modification controls circular RNA immunity, *Mol. Cell* 76 (1) (2019) 96–109.e9.
- [213] L.M. Holdt, A. Kohlmaier, D. Teupser, Circular RNAs as therapeutic agents and targets, *Front. Physiol.* 9 (2018) 1262.
- [214] H.M. Li, X.L. Ma, H.G. Li, Intriguing circles: conflicts and controversies in circular RNA research, *Wiley Interdisciplinary Reviews: RNA*. 10 (5) (2019) e1538.
- [215] E. Kim, Y.K. Kim, S.-J.V. Lee, Emerging functions of circular RNA in aging, *Trends Genet.* 37 (9) (2021) 819–829.
- [216] D. Rodríguez-Leal, Z.H. Lemmon, J. Man, M.E. Bartlett, Z.B. Lippman, Engineering quantitative trait variation for crop improvement by genome editing, *Cell* 171 (2) (2017), 470–80.e8.
- [217] N. Babel, C. Hugo, T.H. Westhoff, Vaccination in patients with kidney failure: lessons from COVID-19, *Nat. Rev. Nephrol.* (2022) 1–16.
- [218] P. Sanchez-Moreno, J.L. Ortega-Vinuesa, J.M. Peula-Garcia, J.A. Marchal, H. Boulaiz, Smart drug-delivery systems for cancer nanotherapy, *Curr. Drug Targets* 19 (4) (2018) 339–359.
- [219] A.M. Ito, S.L. Vemula, M.T. Gupta, M.V. Giram, S.A. Kumar, B. Ghosh, et al., Multifunctional graphene oxide nanoparticles for drug delivery in cancer, *J. Contr. Release* 350 (2022) 26–59.
- [220] X. Zhao, Y. Zhong, X. Wang, J. Shen, W. An, Advances in circular RNA and its applications, *Int. J. Med. Sci.* 19 (6) (2022) 975–985.
- [221] A.Z. Wang, R. Langer, O.C. Farokhzad, Nanoparticle delivery of cancer drugs, *Annu. Rev. Med.* 63 (2012) 185–198.
- [222] X. Ma, Y. Zhao, X.-J. Liang, Theranostic nanoparticles engineered for clinic and pharmaceuticals, *Accounts Chem. Res.* 44 (10) (2011) 1114–1122.
- [223] A.C. Oliveira, J. Fernandes, A. Gonçalves, A.C. Gomes, M. Oliveira, Lipid-based nanocarriers for siRNA delivery: challenges, strategies and the lessons learned from the DODAX: MO liposomal system, *Curr. Drug Targets* 20 (1) (2019) 29–50.
- [224] W.W. Du, W. Yang, X. Li, F.M. Awan, Z. Yang, L. Fang, et al., A circular RNA circDNMT1 enhances breast cancer progression by activating autophagy, *Oncogene* 37 (44) (2018) 5829–5842.
- [225] L. Fang, W.W. Du, F.M. Awan, J. Dong, B.B. Yang, The circular RNA circ-Ccnb1 dissociates Ccnb1/Cdk1 complex suppressing cell invasion and tumorigenesis, *Cancer Lett.* 459 (2019) 216–226.
- [226] M. Lu, Y. Wu, B. Zeng, J. Sun, Y. Li, J. Luo, et al., CircEHMT1 inhibits metastatic potential of breast cancer cells by modulating miR-1233-3p/KLF4/MMP2 axis, *Biochem. Biophys. Res. Commun.* 526 (2) (2020) 306–313.
- [227] W.W. Du, L. Fang, W. Yang, N. Wu, F.M. Awan, Z. Yang, et al., Induction of tumor apoptosis through a circular RNA enhancing Foxo3 activity, *Cell Death Differ.* 24 (2) (2017) 357–370.
- [228] A. Sani, C. Cao, D. Cui, Toxicity of gold nanoparticles (AuNPs): a review, *Biochemistry and biophysics reports* 26 (2021) 100991.
- [229] B.M. Hussien, G.S.H. Faraj, M.F. Rasul, H.J. Hidayat, A. Salihi, A. Baniahmad, et al., Strategies to overcome the main challenges of the use of exosomes as drug carrier for cancer therapy, *Cancer Cell Int.* 22 (1) (2022) 323.
- [230] B.M. Hussien, S. Mohamad-tahr, S.R. Abdullah, H.J. Hidayat, M.F. Rasul, G. S. Hama Faraj, et al., Exosomal circular RNAs: new player in breast cancer progression and therapeutic targets, *Front. Genet.* 14 (2023).
- [231] S. Muthu, A. Bapat, R. Jain, N. Jeyaraman, M. Jeyaraman, Exosomal therapy—a new frontier in regenerative medicine, *Stem Cell Invest.* 8 (2021) 7.
- [232] G. Hu, K.M. Drescher, X.-M. Chen, Exosomal miRNAs: biological properties and therapeutic potential, *Front. Genet.* 3 (2012) 56.
- [233] W. Ke, K.A. Afonin, Exosomes as natural delivery carriers for programmable therapeutic nucleic acid nanoparticles (NANPs), *Adv. Drug Deliv. Rev.* 176 (2021) 113835.
- [234] J. Qian, Q. Zhang, M. Liu, Y. Wang, M. Lu, A portable system for isothermal amplification and detection of exosomal microRNAs, *Biosens. Bioelectron.* 196 (2022) 113707.
- [235] X. Wang, J. Huang, W. Chen, G. Li, Z. Li, J. Lei, The updated role of exosomal proteins in the diagnosis, prognosis, and treatment of cancer, *Exp. Mol. Med.* (2022) 1–11.
- [236] F. Hüllin-Matsuda, P. Colosetti, M. Rabia, C. Luquain-Costaz, I. Delton, Exosomal lipids from membrane organization to biomarkers: Focus on an endolysosomal-specific lipid, *Biochimie* 203 (2022 Dec 1) 77–92; Hüllin-Matsuda F., Colosetti P., Rabia M., Luquain-Costaz C., Delton I. Exosomal lipids from membrane organization to biomarkers: Focus on an endolysosomal-specific lipid. *Biochimie*. 2022 Dec 1;203:77-92.
- [237] Y. Li, J. Wu, E. Li, Z. Xiao, J. Lei, F. Zhou, et al., TP53 mutation detected in circulating exosomal DNA is associated with prognosis of patients with hepatocellular carcinoma, *Cancer Biol. Ther.* 23 (1) (2022) 439–445.
- [238] H. Bai, K. Lei, F. Huang, Z. Jiang, X. Zhou, Exo-circRNAs: a new paradigm for anticancer therapy, *Mol. Cancer* 18 (1) (2019) 1–10.
- [239] T. Lener, M. Gimona, L. Aigner, V. Börger, E. Buzas, G. Camussi, et al., Applying extracellular vesicles based therapeutics in clinical trials—an ISEV position paper, *J. Extracell. Vesicles* 4 (1) (2015) 30087.
- [240] R. Wu, H. Li, C. Sun, J. Liu, D. Chen, H. Yu, et al., Exosome-based strategy for degenerative disease in orthopedics: recent progress and perspectives, *Journal of Orthopaedic Translation* 36 (2022) 8–17.
- [241] N. Mecozzi, A. Nenci, O. Vera, I. Bok, A. Falzone, G.M. DeNicola, et al., Genetic tools for the stable overexpression of circular RNAs, *RNA Biol.* 19 (1) (2022) 353–363.
- [242] I. Jost, L.A. Shalamova, G.K. Gerresheim, M. Niepmann, A. Bindereif, O. Rossbach, Functional sequestration of microRNA-122 from Hepatitis C Virus by circular RNA sponges, *RNA Biol.* 15 (8) (2018) 1032–1039.
- [243] O. Rossbach, Artificial circular RNA sponges targeting MicroRNAs as a novel tool in molecular biology, *Mol. Ther. Nucleic Acids* 17 (2019) 452–454.
- [244] Y. Shu, K. Wu, Z. Zeng, S. Huang, X. Ji, C. Yuan, et al., A simplified system to express circularized inhibitors of miRNA for stable and potent suppression of miRNA functions, *Mol. Ther. Nucleic Acids* 13 (2018) 556–567.
- [245] X. Gao, X.-K. Ma, X. Li, G.-W. Li, C.-X. Liu, J. Zhang, et al., Knockout of circRNAs by base editing back-splice sites of circularized exons, *Genome Biol.* 23 (1) (2022) 1–22.
- [246] Y. Zhang, T.M. Nguyen, X.-O. Zhang, L. Wang, T. Phan, J.G. Clohessy, et al., Optimized RNA-targeting CRISPR/Cas13d technology outperforms shRNA in identifying functional circRNAs, *Genome Biol.* 22 (1) (2021) 1–22.
- [247] T.N. Aung, Z. Qu, R.D. Kortschak, D.L. Adelson, Understanding the effectiveness of natural compound mixtures in cancer through their molecular mode of action, *Int. J. Mol. Sci.* 18 (3) (2017) 656.
- [248] O.O. Abudayyeh, J.S. Gootenberg, P. Essletzbichler, S. Han, J. Joung, J.J. Belanto, et al., RNA targeting with CRISPR-Cas13, *Nature* 550 (7675) (2017) 280–284.
- [249] D.B. Cox, J.S. Gootenberg, O.O. Abudayyeh, B. Franklin, M.J. Kellner, J. Joung, et al., RNA editing with CRISPR-Cas13, *Science* 358 (6366) (2017) 1019–1027.
- [250] B. Kleaveland, C.Y. Shi, J. Stefano, D.P. Bartel, A network of noncoding regulatory RNAs acts in the mammalian brain, *Cell* 174 (2) (2018) 350–362.e17.
- [251] S. Ghosal, S. Das, R. Sen, P. Basak, J. Chakrabarti, Circ2Traits: a comprehensive database for circular RNA potentially associated with disease and traits, *Front. Genet.* 4 (2013) 283.
- [252] Y. Zhang, L. Sun, L. Xuan, Z. Pan, K. Li, S. Liu, et al., Reciprocal changes of circulating long non-coding RNAs ZFAS1 and CDR1AS predict acute myocardial infarction, *Sci. Rep.* 6 (1) (2016) 1–11.
- [253] L. Stoll, J. Sobel, A. Rodriguez-Trejo, C. Guay, K. Lee, M.T. Venø, et al., Circular RNAs as novel regulators of β -cell functions in normal and disease conditions, *Mol. Metabol.* 9 (2018) 69–83.
- [254] G. Chen, Q. Wang, Z. Li, Q. Yang, Y. Liu, Z. Du, et al., Circular RNA CDR1as promotes adipogenic and suppresses osteogenic differentiation of BMSCs in steroid-induced osteonecrosis of the femoral head, *Bone* 133 (2020) 115258.

- [255] W.W. Du, W. Yang, Y. Chen, Z.-K. Wu, F.S. Foster, Z. Yang, et al., Foxo3 circular RNA promotes cardiac senescence by modulating multiple factors associated with stress and senescence responses, *Eur. Heart J.* 38 (18) (2017) 1402–1412.
- [256] Z. Shen, L. Zhou, C. Zhang, J. Xu, Reduction of circular RNA Foxo3 promotes prostate cancer progression and chemoresistance to docetaxel, *Cancer Lett.* 468 (2020) 88–101.
- [257] L. Zhang, D. Liang, C. Chen, Y. Wang, G. Amu, J. Yang, et al., Circular siRNAs for reducing off-target effects and enhancing long-term gene silencing in cells and mice, *Mol. Ther. Nucleic Acids* 10 (2018) 237–244.
- [258] P. Obi, Y.G. Chen, The design and synthesis of circular RNAs, *Methods* 196 (2021) 85–103.
- [259] S.T. Croke, J.L. Witztum, C.F. Bennett, B.F. Baker, RNA-targeted therapeutics, *Cell Metabol.* 27 (4) (2018) 714–739.
- [260] K.H. Lee, S. Kim, J. Song, S.R. Han, J.H. Kim, S.-W. Lee, Efficient circular RNA engineering by end-to-end self-targeting and splicing reaction using Tetrahymena group I intron ribozyme, *Mol. Ther. Nucleic Acids* 33 (2023) 587–598.
- [261] Y. Qi, W. Han, D. Chen, J. Zhao, L. Bai, F. Huang, et al., Engineering circular RNA regulators to specifically promote circular RNA production, *Theranostics* 11 (15) (2021) 7322.
- [262] T.L. Young, K.C. Wang, A.J. Varley, B. Li, Clinical delivery of circular RNA: lessons learned from RNA drug development, *Adv. Drug Deliv. Rev.* (2023) 114826.
- [263] R. Luo, H. Le, Q. Wu, C. Gong, Nanoplatform-based in vivo gene delivery systems for cancer therapy, *Small* (2024) 2312153.
- [264] X. Cheng, Z. Li, R. Shan, Z. Li, S. Wang, W. Zhao, et al., Modeling CRISPR-Cas13d on-target and off-target effects using machine learning approaches, *Nat. Commun.* 14 (1) (2023) 752.
- [265] M. Wang, J. Xu, J. Meng, X. Huang, Synthetic circular gRNA mediated biological function of CRISPR-d Cas9 system, *Front. Cell Dev. Biol.* 10 (2022) 863431.
- [266] S. Li, X. Li, W. Xue, L. Zhang, L.-Z. Yang, S.-M. Cao, et al., Screening for functional circular RNAs using the CRISPR-Cas13 system, *Nat. Methods* 18 (1) (2021) 51–59.
- [267] X. Yin, H. Lin, L. Lin, L. Miao, J. He, Z. Zhuo, LncRNAs and CircRNAs in cancer, *MedComm* 3 (2) (2022) e141.
- [268] B. Ma, S. Wang, W. Wu, P. Shan, Y. Chen, J. Meng, et al., Mechanisms of circRNA/lncRNA-miRNA interactions and applications in disease and drug research, *Biomed. Pharmacother.* 162 (2023) 114672.
- [269] B. Zhang, CRISPR/Cas gene therapy, *J. Cell. Physiol.* 236 (4) (2021) 2459–2481.
- [270] R. Zhao, J. Fu, L. Zhu, Y. Chen, B. Liu, Designing strategies of small-molecule compounds for modulating non-coding RNAs in cancer therapy, *J. Hematol. Oncol.* 15 (1) (2022) 14.